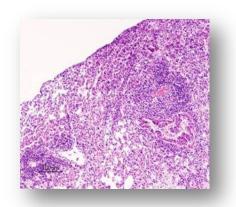
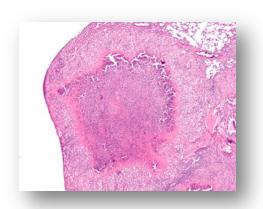
# Pre-clinical tools for evaluating new components of TB regimens

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July 19, 2017









# "All models are wrong, some are useful."



George Box



# In vitro and in vivo models Pros & cons

#### *In vitro* models

#### **Pros**

- controlled microenvironment
- unlimited range of doses, schedules
- more precise measurement of drug conc. to which Mtb is exposed
- simple, serial sampling of Mtb products

#### Cons

 difficult to account for host effects on lesion microenvironment, microbial growth and susceptibility, and drug exposures at site of infection

### In vivo models



#### **Pros**

- embody dynamic interaction b/w host, drug and microbe & represent impact of pathology
- enable simultaneous study of multiple sub-populations, perhaps in clinically relevant proportions

#### Cons

- limitations in dose size and schedules
- often difficult to mimic human PK
- may or may not represent diverse human disease states well

June 26, 2014

### **EMA** qualification opinion on the HFS-TB

- HFS-TB qualified for use in drug development programs as an additional and complementary tool
- HFS-TB can be used in regulatory submissions, esp. for informed design and interpretation of clinical studies
- HFS-TB is recommended to be useful as follows:
  - To provide preliminary proof of concept for developing a specific drug or combination to treat tuberculosis
  - To select the pharmacodynamic target (e.g. T<sub>>MIC</sub>, AUC/MIC)
  - To provide data to support PK/PD analyses leading to initial dose selection for non-clinical and clinical studies
  - To assist in confirming dose regimens for later clinical trials taking into account human PK data and exposure-response relationships

### Some unanswered questions for HFS-TB

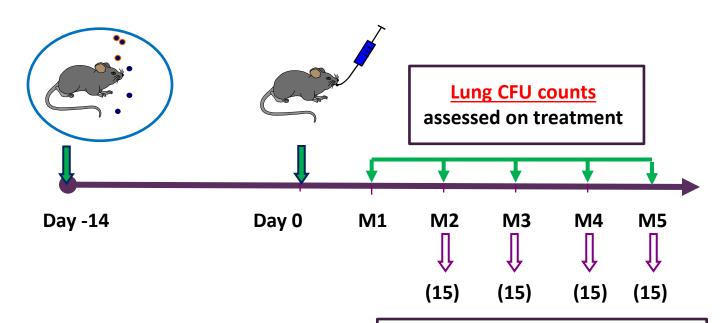
- Reproducibility?
- Obstacles to technology transfer and uptake?
- Reliability of estimates of drug exposures at site of infection (eg, using free drug fraction, ELF penetration ratios)?
- Predictive accuracy for efficacy of regimens?
  - rank ordering existing and novel regimens
  - estimating absolute or relative treatment durations
- Optimal integration of log phase and sterilizing effect models to predict regimen efficacy?

### **Caveat**

"Correlations between drug concentration and pathogen survival that are based on in vitro models cannot be expected to reiterate all aspects of in vivo antimycobacterial treatment."

Chilukuri et al, CID 2015; 61(S1):S32

### Scheme for relapse-based experiments in mice

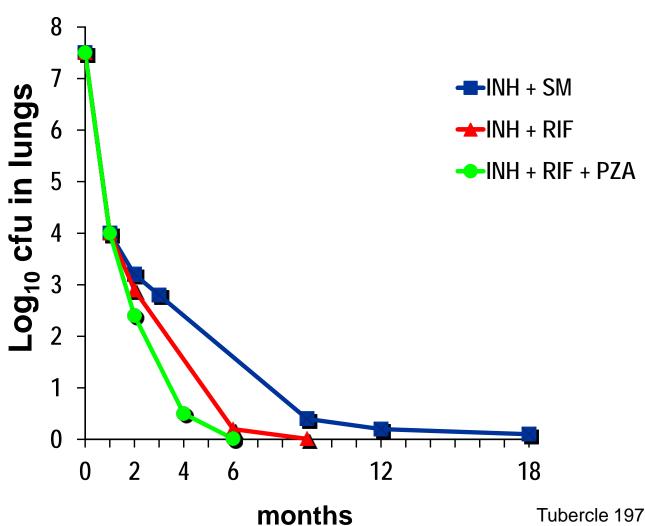


Relapse-free cure (absence of cultivable bacilli) assessed after holding mice without treatment for 3-6 months

# Current uses of mouse models in the context of TB regimen development

- Derive (or confirm) PK/PD relationships for selecting optimal doses of component drugs
- Rank order drug combinations on the basis of efficacy
- Estimate treatment-shortening potential
- Assess impact of caseous pathology (eg, in Kramnik mice)
- Estimate potential for selection of drug-resistant mutants

## Recapitulation of the short-course regimen in the mouse...as in humans



Tubercle 1978: 59:287 & 1986;67:5 AAC (2016); 60:1091

# Performance of novel regimens in BALB/c mice (HDA model)

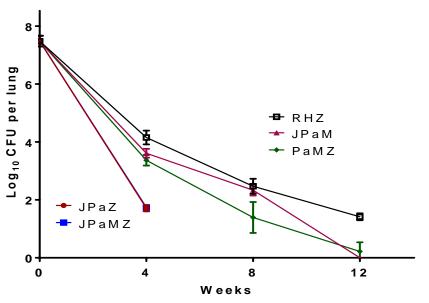
Regimen	Treatment-shortening effect (in months) relative to RHZ(E)	
RMZ(E)	1 - 1.5	
PaMZ	0 - 1	
BPaMZ	3 - 3.5	
BPaL	1-1.5 months	

Pa = pretomanid; M = moxifloxacin; Z = pyrazinamide; B = bedaquiline; L = linezolid

- The PCS working group of CPTR has embarked on an effort to quantify the predictive accuracy of the "sterilizing" mouse model
- New regimens in, or advancing to, phase 3 trials will provide additional opportunities to evaluate the correspondence

## Contribution of component drugs to the efficacy of the BPaMZ regimen

### Bactericidal effect



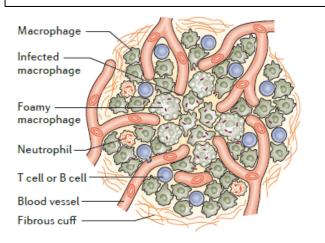
### Sterilizing effect

	Proportion of mice relapsing by time point:				
Regimen	M1.5 (+3)	M2 (+3)	M3 (+3)	M4 (+3)	M5 (+3)
RHZ				10/15	2/15
PaMZ			10/14	3/15	
JPaM			2/15	0/14	
JPaZ	13/14	0/15	0/15		
JPaMZ	3/15	0/15	0/15		

## Challenges in translating mouse model results to the clinic

### Cellular granuloma

BALB/c, C3HeB/FeJ mice



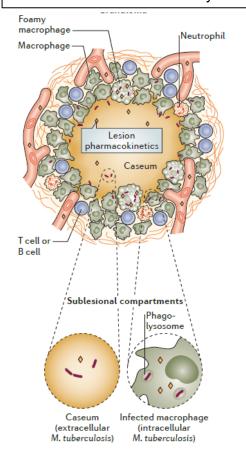
response and lung pathology



#### Differences in:

Mtb growth rate
Intra/extracellular residence
Drug distribution
Lesion microenvironment

### Caseating granuloma C3HeB/FeJ mice only



# Examples of different outcomes in C3HeB/FeJ mice compared to BALB/c mice

- Lack of <u>pyrazinamide</u> bactericidal effect in large caseous lesions, where caseum has neutral pH<sup>1</sup>
  - yet, addition of PZA to RIF+INH+EMB still shortens treatment duration<sup>2</sup>
- Lack of <u>clofazimine</u> bactericidal effect in large caseous lesions, where CFZ diffuses poorly and caseum has neutral pH and is hypoxic<sup>3</sup>
- Reduced <u>bedaquiline</u> effect in large caseous lesions, where BDQ diffuses poorly<sup>4</sup>
  - 1. Lanoix et al, AAC 2016; 60:735
  - 2. Lanoix et al, AAC 2016; 60:1091
  - 3. Irwin et al, AAC 2014; 58:4026
  - 4. Irwin et al, ACS Infect Dis 2016; 2:251

# Limited experience comparing regimens in other animal models with caseous pathology

### Guinea pigs

- Replacing RIF with RPT had no significant treatment-shortening effect<sup>1</sup>
- PaMZ had no significant treatment shortening effect compared to RHZ<sup>2</sup>

#### Rabbits

No regimen comparisons found

#### Marmosets

 RHZE reduced the extent of disease on PET-CT and lowered CFU counts in cavities compared to HS after 6 weeks of treatment<sup>3</sup>

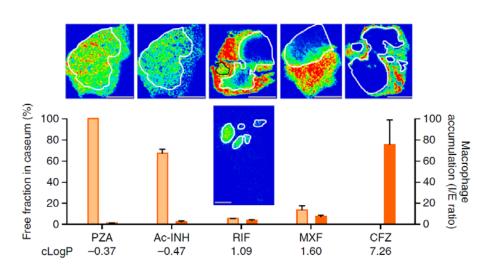
### Macaques

Metronidazole did not increase the bactericidal effect of RH

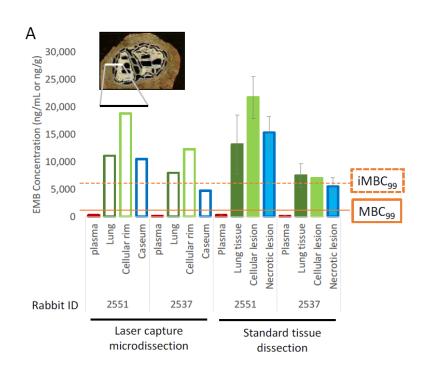
- 1. Ahmad et al, AAC 2012; 56:3726
- 2. Dutta et al, AAC 2013; 57:3910
- 3. Via et al, AAC 2014; 59:4181

### **Evaluating drug partitioning into TB lesions**

#### **MALDI-MSI**



### Laser-capture microdissection and LC/MS-MS



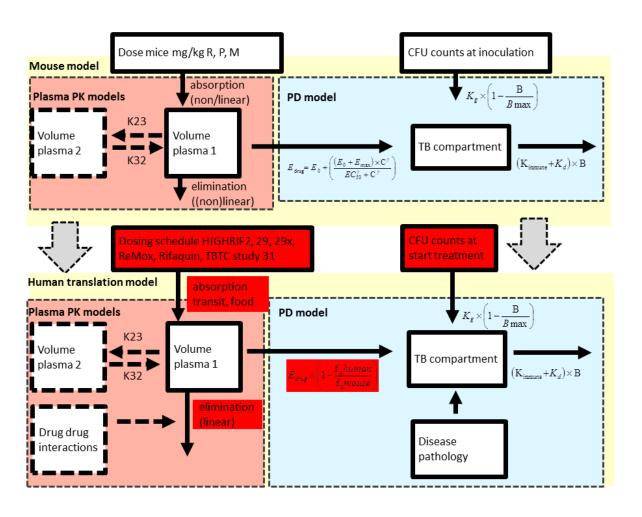
# Additional challenges in translating mouse model results to the clinic

- Inter-species differences in drug PK
- Experiments in <u>inbred mice</u> infected with <u>1 Mtb strain</u> and treated with <u>identical drug doses</u> cannot recapitulate the many sources of heterogeneity in human TB:
  - PK variability
  - Severity of disease (eg, presence of cavities, cavity size)
  - Immune status
  - Adherence to treatment
  - Mtb drug susceptibility

## Translational (mouse → human) PK/PD Model Objectives

- Develop a translational PK/PD model that utilizes:
  - mouse PK data for RIF, RPT and MXF
  - mouse PD data (CFU counts) for RIF, RPT and MXF alone and in combinations including PZA + INH or EMB
  - human PK data, including rifamycin-MXF interaction
  - an immune effect on bacterial death derived from CFU differences between nude and BALB/c mice
  - inter-species differences in protein binding
  - effect of caseation and cavitation on lesion distribution of rifamycins
- Perform clinical trial simulations to predict trial outcomes
  - sputum culture status at 8 wks
  - relapse status at 1 yr

### Translational (Mouse → Human) PK/PD Model



R= rifampin

P= rifapentine

M= moxifloxacin

B<sub>MAX</sub>= maximum number of bacteria

K<sub>growth</sub>= bacterial growth constant

K<sub>death</sub>= bacterial death constant

 $IT_{50}$  = time of 50% of max. immune response

 $\theta_{KIND}$ = max. immune kill rate (untreated mice)

γ<sub>immune response</sub> = sigmoidicity factor, defines shape of immune response effect

 $\theta_{\text{KDOI.0}}$ = immune kill rate (treated mice) at average incubation time

 $\theta_{\text{KDOI.t}}$ = increase in kill rate (treated mice) in expts w/above average incubation time

 $E_{drug} = drug effect$ 

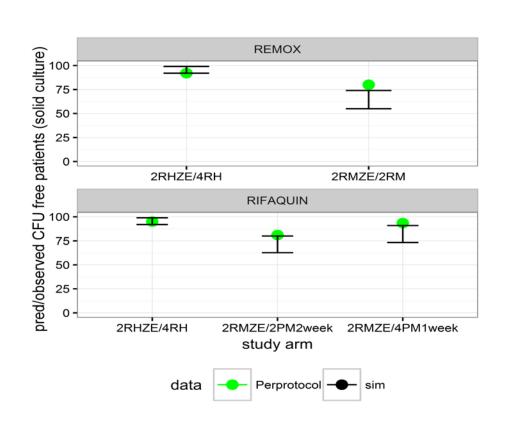
 $E_{max}$  = max. achievable drug effect

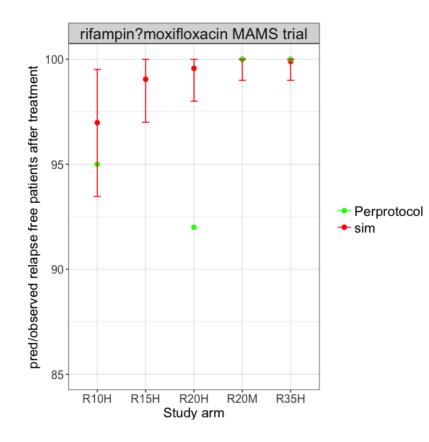
 $EC_{50}$  = antibiotic conc. producing 50% of  $E_{max}$ 

γ = sigmoidicity factor, defines the shape of drug effect

### Using the final translational PK/PD model:

### Predicted vs. observed trial results



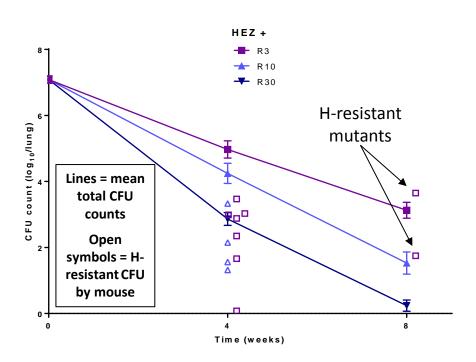


### Translational PK/PD Model - conclusions

- The model performed reasonably well, especially in predicting higher relapse rates for 4-month arms
- Work is ongoing to:
  - incorporate individual PK/PD and dose-response for all drugs in regimens
  - simulate phase 3 trials with more novel regimens
  - incorporate drug-resistant sub-populations to predict rates of resistance emergence
  - merge PK/PD model with mechanistic within-host model to gain greater insight into factors driving regimen performance

### Assessing the risk of resistance amplification

## Impact of simulated RIF PK variability in C3HeB/FeJ mice



Low R exposures lead to acquired INH resistance in 2 of 10 mice

### Impact of intermittency and immunodeficiency in nude mice

Recapitulating the arms of TBTC Study 22, selection of AHR and ARR were associated with:

- <u>immunosuppression</u>
  - nude mice more likely than BALB/c to have AHR (8.5% vs 0%, p= 0.001) and ARR (3.5% vs. 0%, p= 0.06)
- intermittent vs daily initial phase therapy
  - 30% vs 2.7% for AHR/ARR (p< 0.001)</li>
  - 20% vs 2.7% for ARR only (p< 0.01)</li>
- once-weekly RPT vs RH in contin. phase
  - 18% vs 3.3% for ARR (p< 0.05)</li>

### **Take-home points**

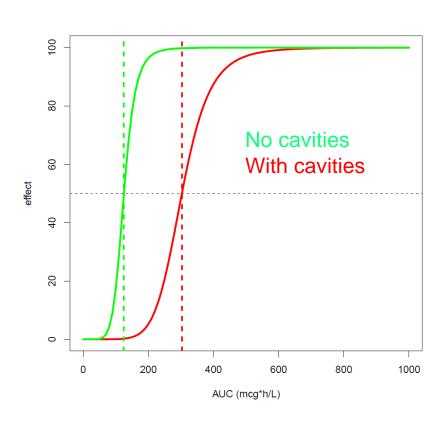
- In vitro hollow fiber models are qualified as useful tools for exploring PK/PD relationships under controlled conditions
- Mouse models have an established track record in estimating the treatment-shortening potential of novel regimens
- The impact of certain variables that modify the effect of some drugs may require elucidation in caseous disease models:
- Emerging data from clinical trials with novel regimens will provide a great opportunity for further evaluating the predictive accuracy of these and other preclinical models
- Some factors are more difficult to account for in pre-clinical models and may be best address with more predictive PK/PD-based translational models:
  - o inter-species PK differences in PK, protein binding, etc
  - human PK variability
  - heterogeneity in human host (eg, cavitation, immune response)
  - heterogeneity in bacterial pathogen (eg, MIC distribution)

### Acknowledgements

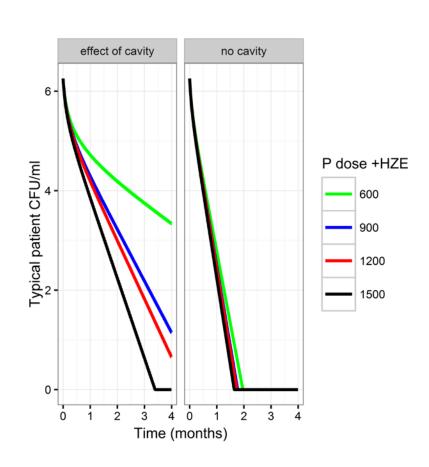
- Members of the lab and the JHU Center for TB Research
- Collaborators
  - Veronique Dartois (Rutgers)
  - Tawanda Gumbo (Baylor)
  - Debra Hanna and CPTR PCS-WG
  - Anne Lenaerts, Scott Irwin (CSU)
  - Chuck Peloquin (UF)
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     Nan Zhang (UCSF)
- Funding
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  - Global Alliance for TB Drug Development
  - FDA (U18-FD004004)
  - Gates Foundation (TBDA #42581, OPP1037174)
  - C-Path
  - CDC TB Trials Consortium



## Incorporating the effect of disease pathology into the model



A 4-fold higher  $EC_{50}$  for P in pts with cavitary disease.

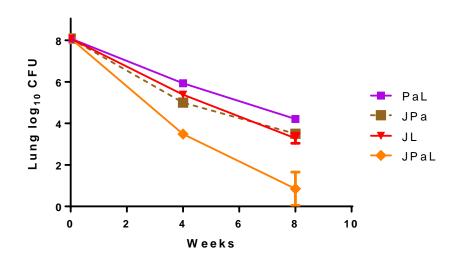


A 4-fold higher  $EC_{50}$  for P in the cavity compartment compared to plasma was used in the simulation.

Bartelink, I. et al. CPT 2017

# Contribution of each component to the efficacy of the BPaL regimen

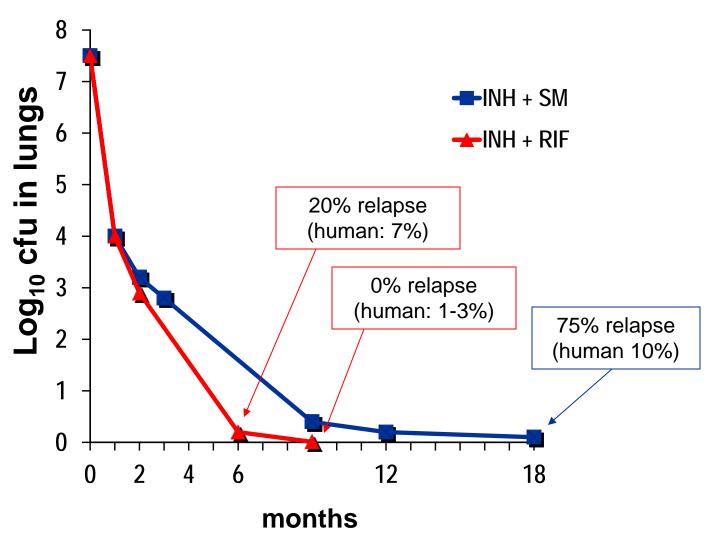
#### Bactericidal effect



### Sterilizing effect

	Proportion relapsing	Proportion relapsing after treatment for:		
Regimen	2 months	3 months		
2RHZ/RH		8/14		
JPa		3/14		
JPaL		0/15		
2JPaL/1JPa	6/15	0/15		
1JPaL/2JPa	9/15	0/15		

## Recapitulation of the short-course regimen in the mouse...as in humans



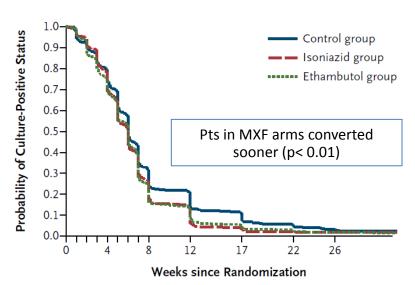
# Recapitulating the evolution of short-course therapy in mice and humans\*

		Proportion Relapsing after Treatment:	
Regimen	Months	Mice	Humans
INH+SM	6	100%	29%
INH+SM	18	75%	~10%
INH+RIF	6	20%	6-7%
INH+RIF	9	0-5%	1-3%
INH+RIF+PZA	4	70-90%	11-15%
INH+RIF+PZA	6	0-5%	1-3%

<sup>\*</sup>From Mitchison; and Grosset & Ji; in Gangadharam & Jenkins, Chapman & Hall, 1998

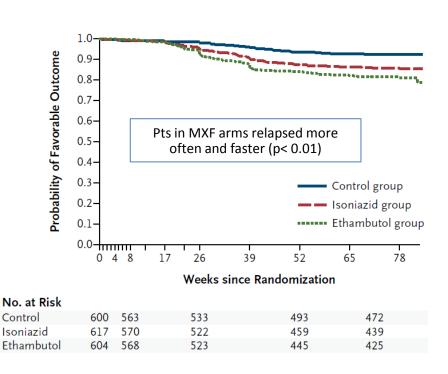
# Moxifloxacin for treatment shortening: The REMoxTB phase 3 trial

#### Time to sputum conversion

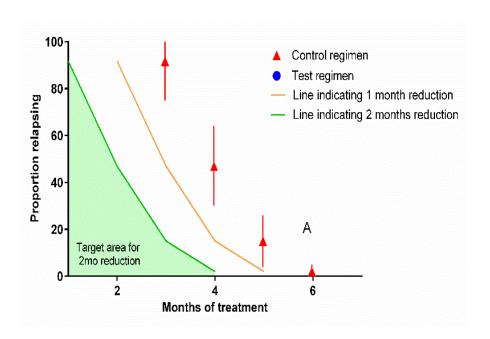


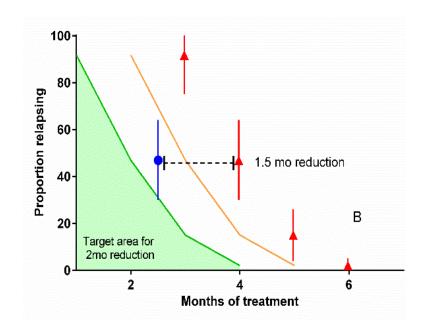
#### No. at Risk Control Isoniazid Ethambutol

#### Time to unfavorable outcome



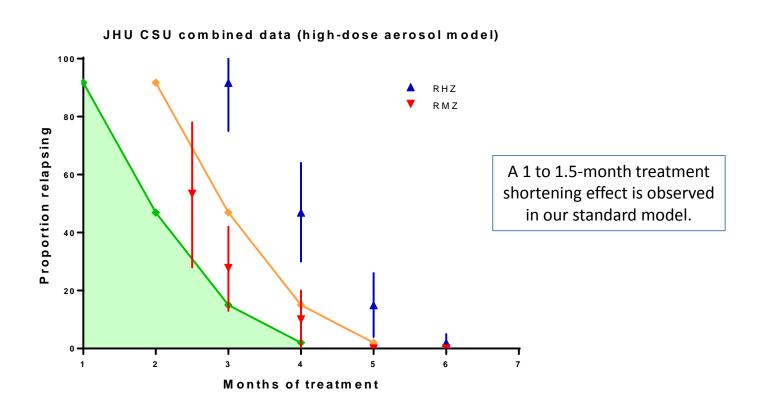
## Measuring the treatment-shortening effect of a test regimen relative to a control regimen in mice





Colored symbols represent the proportion of mice relapsing after receiving the indicated regimen for various durations (error bars represent the 95%CI).

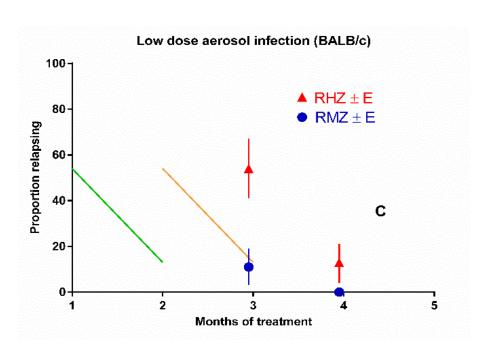
## Treatment-shortening effect of substituting moxifloxacin for isoniazid in the 1<sup>st</sup>-line regimen in BALB/c mice

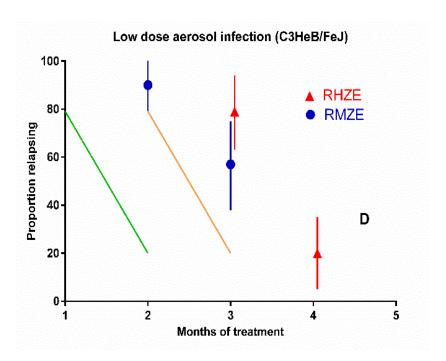


### **Outcomes with REMox-TB regimens in mice**

- In REMox-TB, substitution of M for H or E resulted in faster sputum conversion but did not permit shortening the duration of treatment by 2 months
- In mice, <u>substitution of M for H</u> resulted in:
  - treatment shortening of <u>1-1.5 months</u> in high-dose infection models
  - treatment shortening of <u>0-1 month</u> in low-dose infection models
- In mice, <u>substitution of M for E</u> resulted in:
  - treatment shortening of <u>0-1 month</u> in low-dose infection models
- Results in mice are not inconsistent with those of REMox-TB

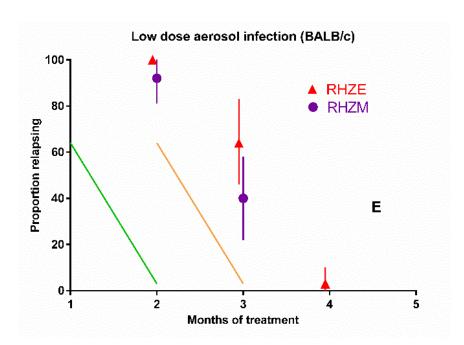
## Treatment-shortening effect of substituting moxifloxacin for isoniazid in the 1<sup>st</sup>-line regimen

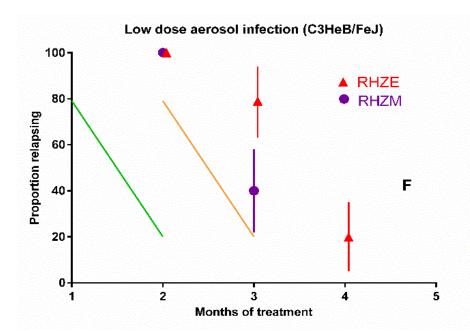




The treatment shortening effect is between 0-1 month in low-dose aerosol infection models in BALB/c and Kramnik mice.

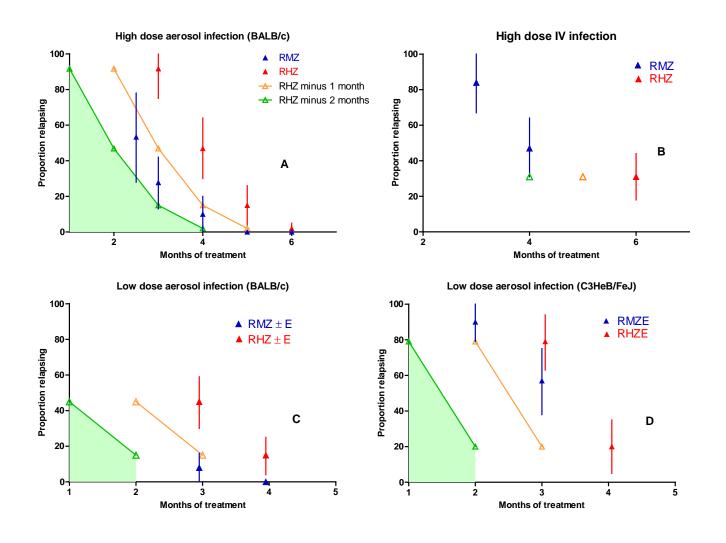
## Treatment-shortening effect of substituting moxifloxacin for ethambutol in the 1<sup>st</sup>-line regimen



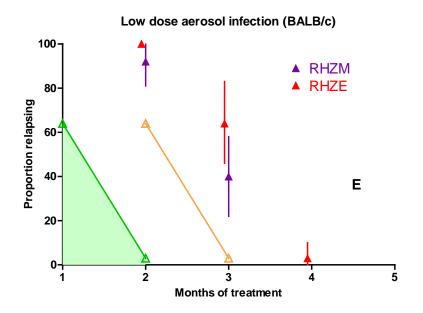


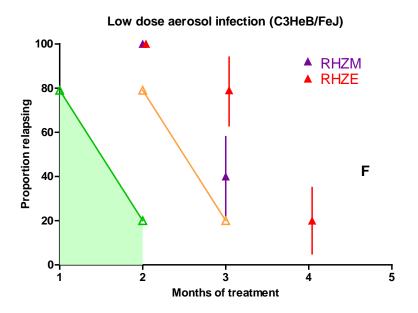
The treatment shortening effect is between 0-1 month in low-dose aerosol infection models in BALB/c and Kramnik mice.

## Substitution of M for H in the RHZ regimen in mice – data from 3 institutions, using low- & high-dose infection models

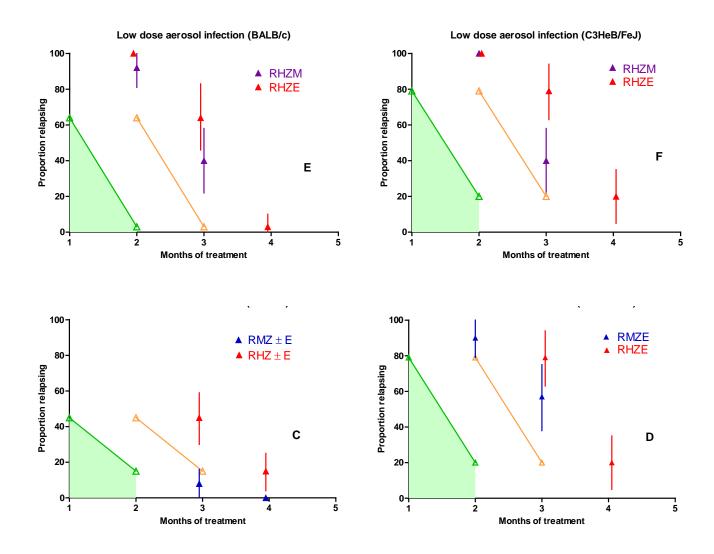


## Substitution of M for E in RHZE – data from 2 institutions in chronic infections in 2 mouse strains





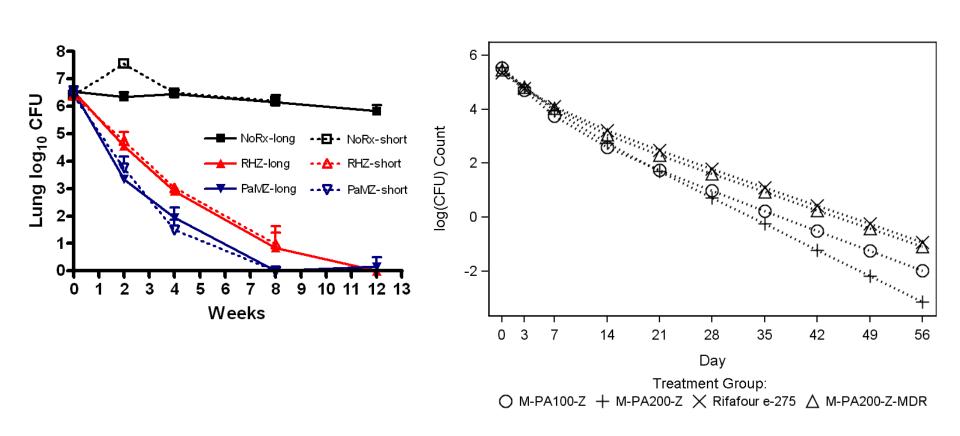
### Substitution of M for H in the RHZ regimen in mice – data from 3 institutions, using low- & high-dose infection models



## Correspondence between results of REMox-TB trial and those in mice

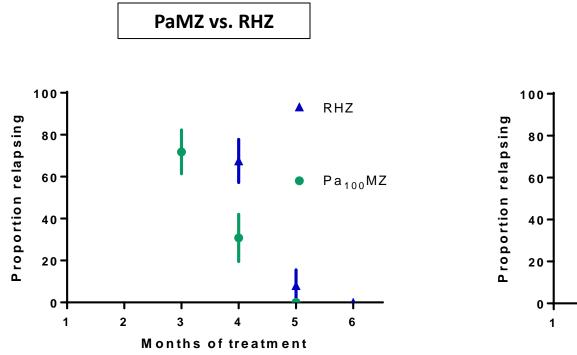
- In REMox-TB, substitution of M for H or E resulted in faster sputum conversion but did not permit shortening the duration of treatment to 4 months
- In mice, substitution of M for H or E reduced CFU cts more rapidly, but relapse rates were inevitably higher when RMZ(E) or RHZM duration was reduced by 2 months relative to RHZ(E)
- Results in C3HeB/FeJ mice may be closer to those of REMox-TB
  - smaller difference for RMZE relative to RHZE
  - no difference between RHZM and RMZE
- The most severely affected C3HeB/FeJ mice may best represent pts most likely to relapse

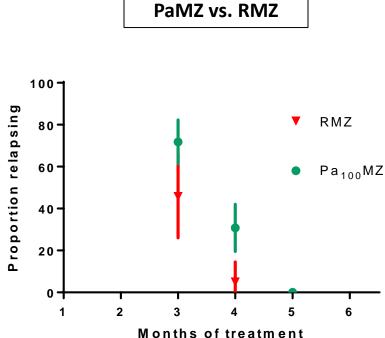
## Bacterial killing with Pa<sub>50</sub>MZ vs. RHZ: mouse vs. NC-002 results



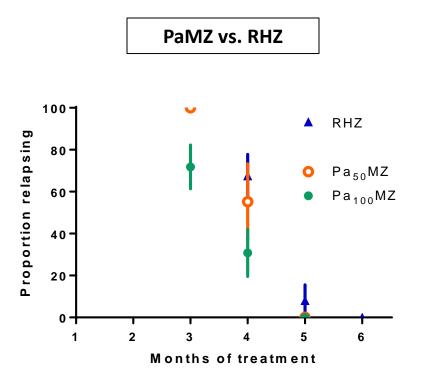
Long = 6 wks from low-dose infxn to treatment onset Short = 2 wks from high-dose infxn to treatment onset

### PaMZ vs. RHZ, RMZ HDA model in BALB/c mice – relapse data

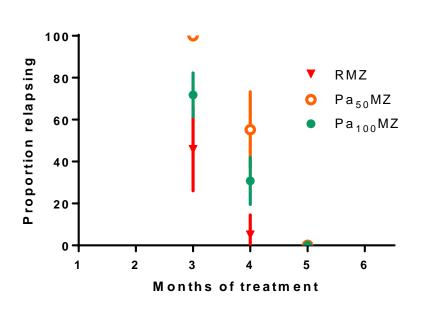




### PaMZ vs. RHZ, RMZ HDA model in BALB/c mice – relapse data



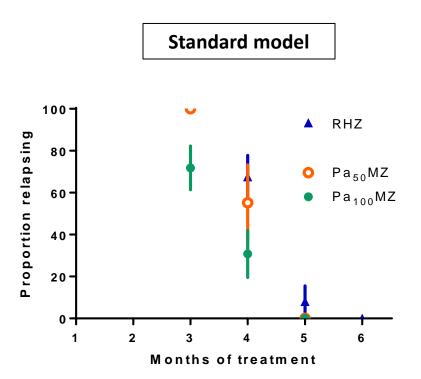


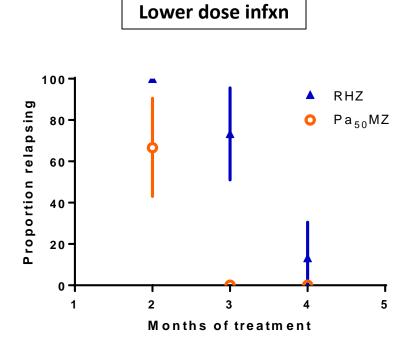


The treatment shortening effect is between 0-1 month when compared to RHZ and depends on the Pa dose.

PaMZ is no more effective than RMZ and appears less effective than RMZ when the Pa dose is 50 mpk.

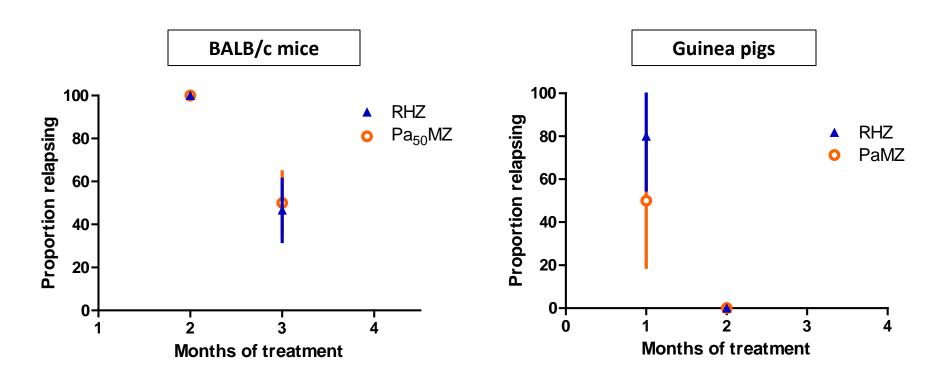
### PaMZ vs. RHZ HDA model in BALB/c mice – relapse data





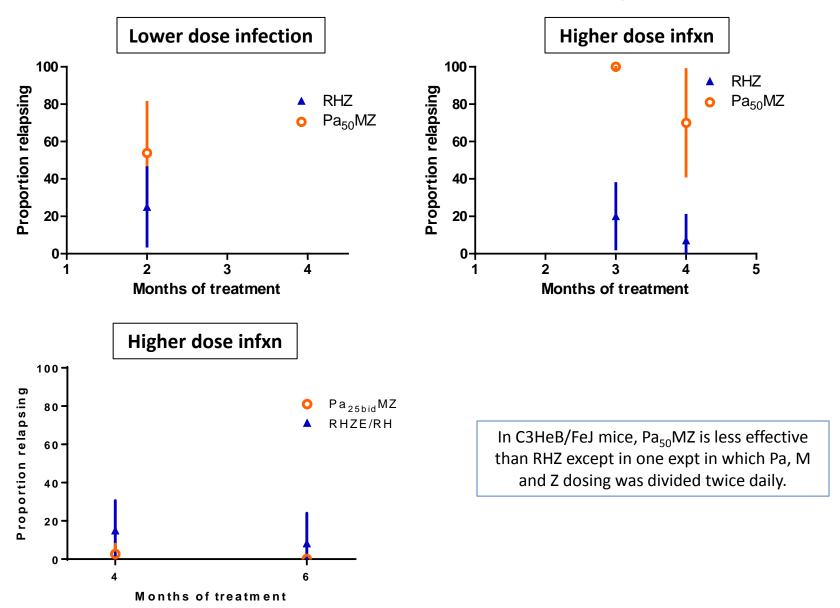
In one experiment in which a lower-dose aerosol infection was used in a 14-day incubation model to match Day 0 CFU counts with a low-dose chronic (42-day) infection model (right panel), the treatment shortening effect of PaMZ was at least 1 month.

### PaMZ vs. RHZ Chronic LDA model in BALB/c mice, guinea pigs



In the chronic low-dose aerosol model in BALB/c mice, Pa<sub>50</sub>MZ is equivalent to RHZ. In the chronic guinea pig model, the treatment shortening effect of PaMZ is between 0 and 1 month.

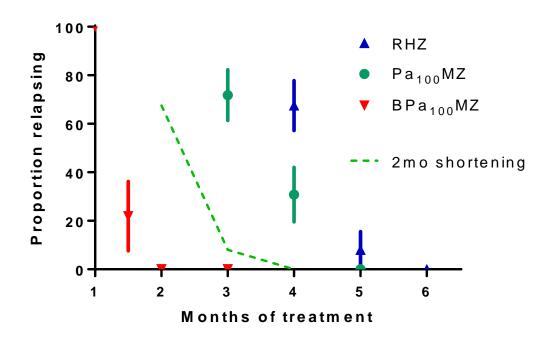
#### PaMZ vs. RHZ LDA model in C3HeB/FeJ mice – relapse data



#### **Conclusions re: PaMZ**

- Results of the abbreviated STAND trial comparing 2RHZE/4RH to 4PaMZ and 6PaMZ should be available soon & will provide a basis for comparison of mouse and human results
- In our standard high-dose aerosol model in BALB/c mice:
  - PaMZ requires <u>0-1 month less treatment</u> to cure compared to RHZ, and this effect is somewhat dose-dependent
  - PaMZ requires <u>0-1 month more treatment</u> to cure compared to RMZ

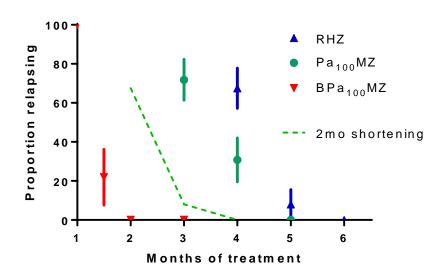
### BPaMZ vs. PaMZ vs. RHZ HDA model in BALB/c mice



In the high-dose aerosol model in BALB/c mice, BPa<sub>100</sub>MZ shortens the duration of treatment by 3-3.5 months compared to RHZ.

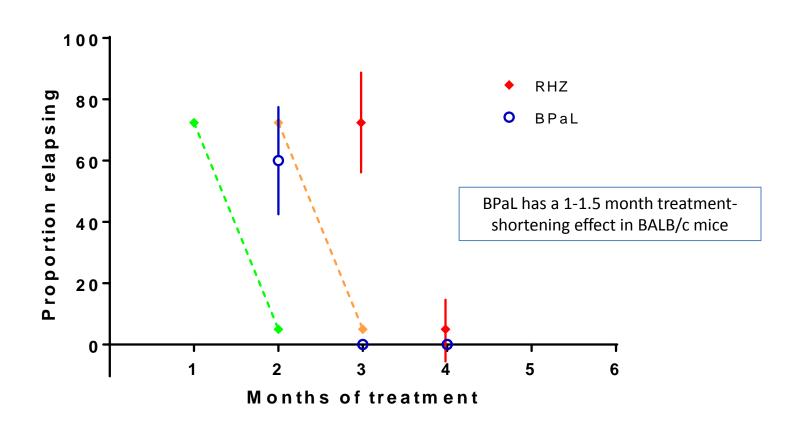
### BPaMZ vs. PaMZ vs. RHZ HDA model in BALB/c mice

#### All expts evaluating PaMZ and/or BPaMZ

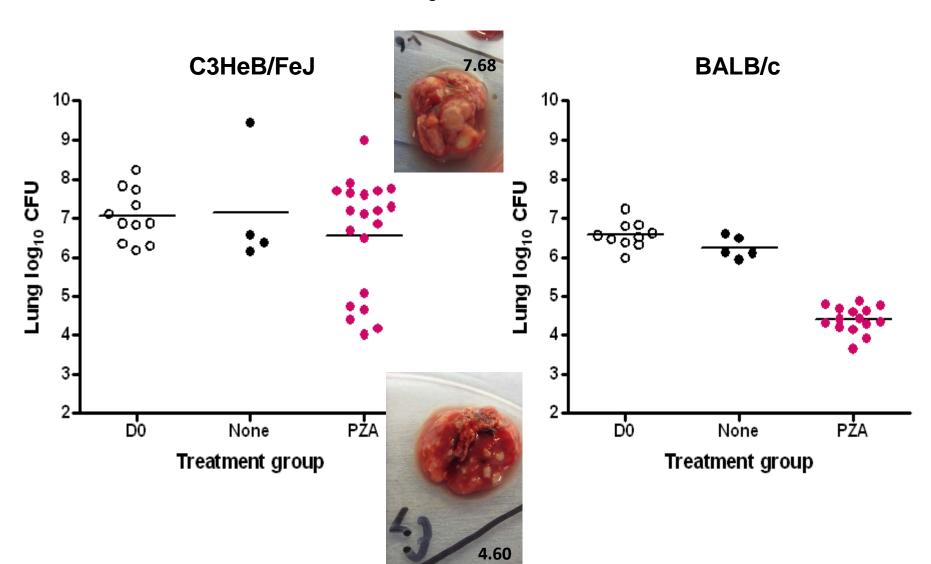


In the high-dose aerosol model in BALB/c mice, BPa<sub>100</sub>MZ shortens the duration of treatment by 3-3.5 months compared to RHZ.

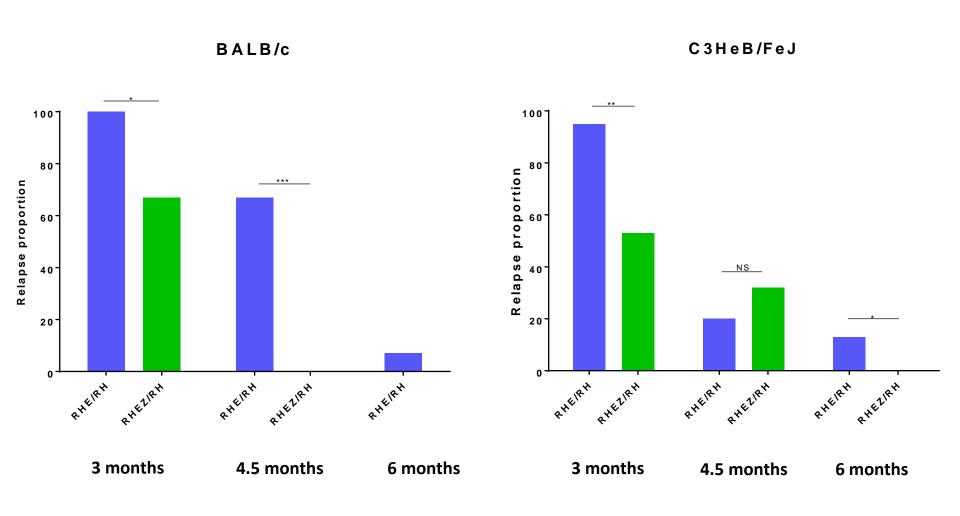
### BPaL vs. RHZ HDA model in BALB/c mice



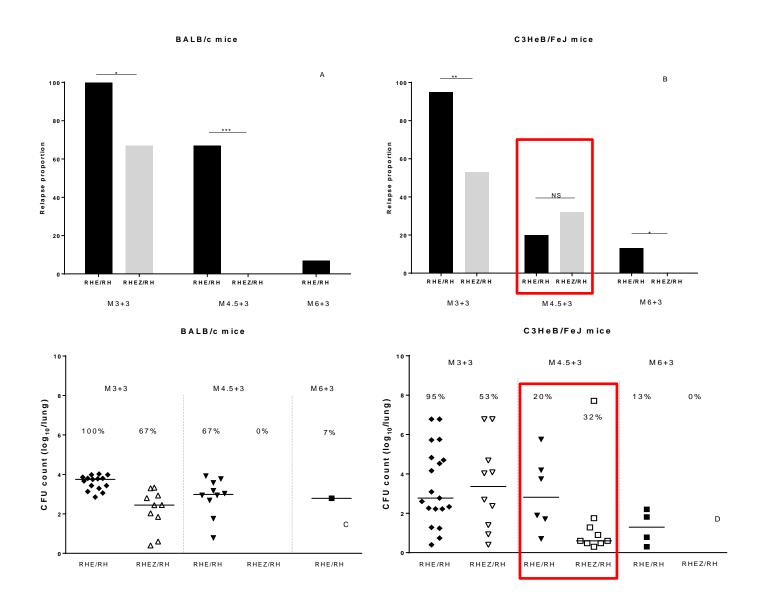
## "Dichotomous" activity of PZA in C3HeB/FeJ mice



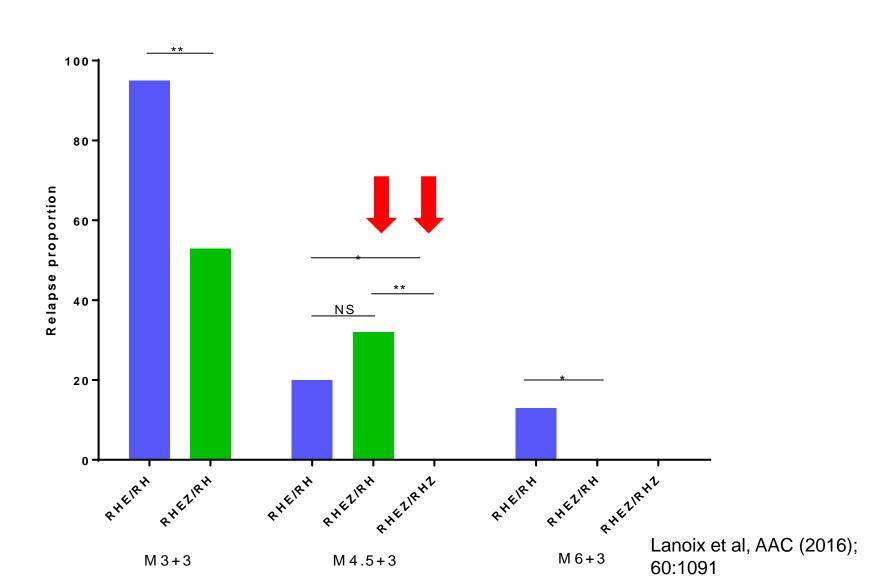
# Z adds sterilizing activity to RHE in BALB/c and C3HeB/FeJ mice



## Sterilizing activity of Z in 1<sup>st</sup>-line regimen in 2 mouse strains



## Increasing duration of PZA increases sterilizing effect in C3HeB/FeJ mice



## Z Adds Sterilizing Activity to RHE in BALB/c and C3HeB/FeJ Mice

	Does use of Z for 2 months reduce relapses compared to no Z?			Does extending Z beyond 2 months reduce relapses?
	3 mo.	4.5 mo.	6 mo.	3-4.5 mo.
BALB/c	Yes	Yes	Yes	No
Kramnik	Yes	No*	Yes	Yes†

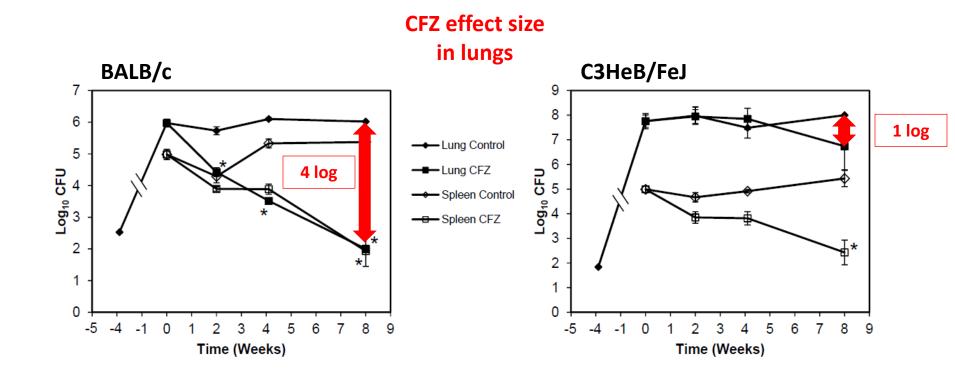
<sup>\*</sup>Z was associated with lower mean CFU count among relapsing mice †2RHEZ/2.5RHZ was associated with fewer relapses than 2RHEZ/2.5RH

Lanoix et al, AAC (2016); 60:1091

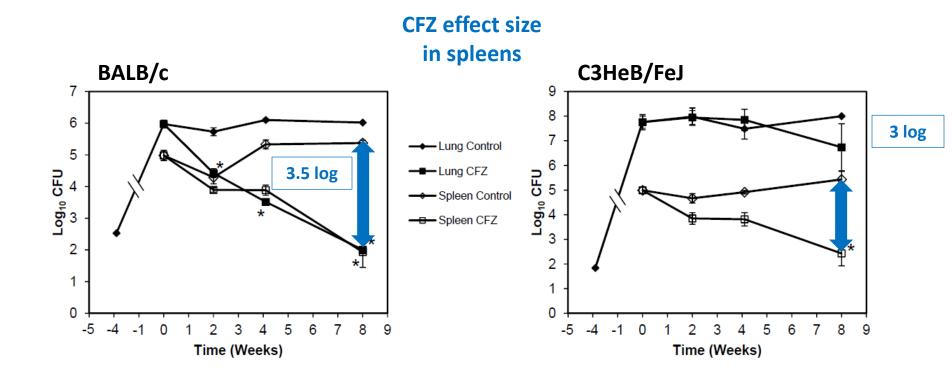
Study	Regimen*	Follow-up (months)	Patients assessed	Bacterio- logical relapses
East African/BMRC (current)	2SHRZ/HRZ 2SHRZ/HR	24 24	40 40	0 1 (2%)
Singapore/BMRC (1981)	2SHRZ/HRZ 2SHRZ/HR	24	78 80	0 2 (2%)

Can we really conclude that continuing Z beyond 2 months would not benefit the most difficult-to-cure patients?

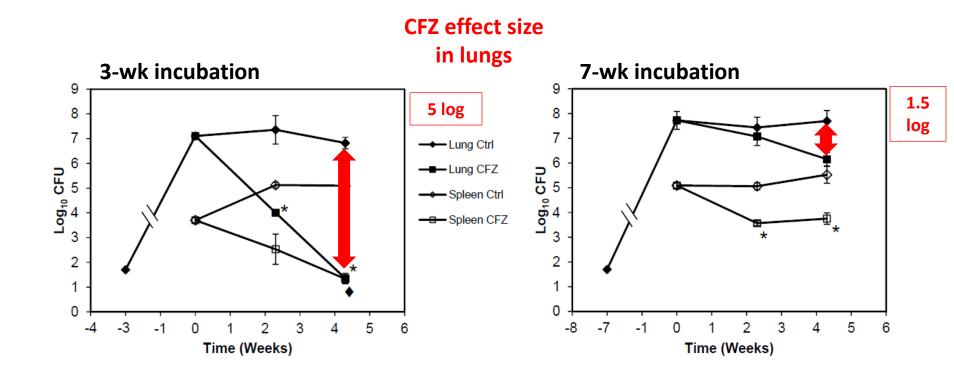
#### Activity of clofazimine in C3HeB/FeJ mice



#### Activity of clofazimine in C3HeB/FeJ mice

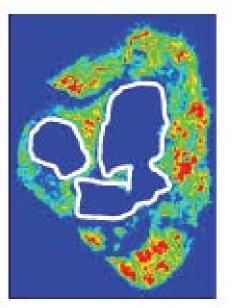


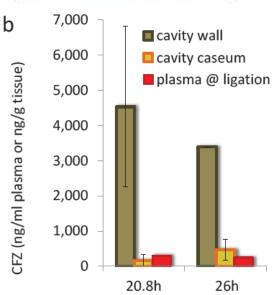
#### Activity of clofazimine in C3HeB/FeJ mice



## Compartmentalized activity of CFZ – PK/distribution

- Slow, steady accumulation in adipose tissue & macrophages
  - pH-dependent ion trapping in lysosomes
- Poor distribution into caseum relative to cavity wall





#### **Data inventory**

- Focus first on mouse strains other than C3HeB/FeJ ("Kramnik")
- Inventory identified a variety of relapse-based pre-clinical studies with corresponding clinical trial outcomes data

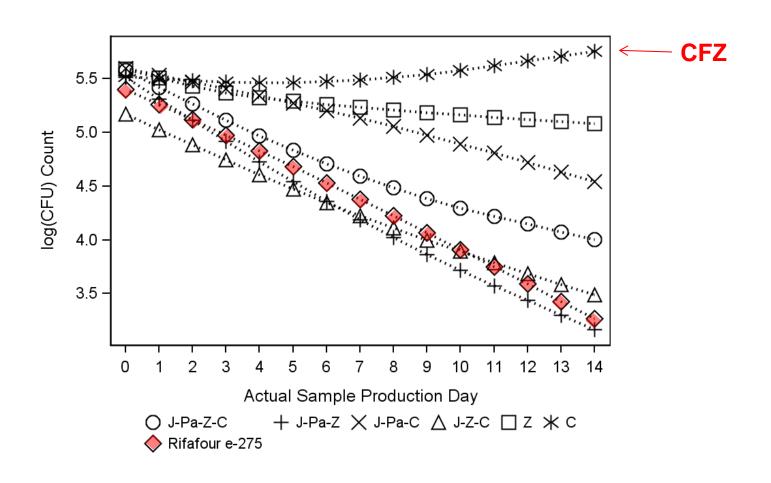
Test regimen intervention	Regimen comparison	# of expts
Combining INH+STR	HS vs. H or S monotherapy	1
Shortening duration of INH+STR	6HS <u>vs</u> . 18HS	1
Adding RIF to INH+STR or INH+EMB+PZA	HR (or HRS or HREZ) vs. HS (or HEZ)	4
Adding STR to INH+RIF	HRS <u>vs</u> . HR	1
Adding PZA to INH+RIF (±STR/EMB)	HRZ (or HRSZ or HREZ) vs. HR (or HRS or HRE)	4
Shortening duration of PZA	2HREZ/4RH vs. 6HREZ	1
Increasing dose of RIF	High-dose R plus HEZ vs. HREZ	2
Extending dosing interval of 1st-line Rx	HREZ (2/7) vs. HREZ (daily)	1
Replacing EMB with MXF	HRMZ <u>vs</u> . HRZ(E)	3
Replacing INH with MXF	MRZ(E) vs. HRZ(E)	10
Replacing RIF with RPT	HPZ(E) vs. HRZ(E)	7
Replacing RIF+EMB with RPT+MXF	H <mark>PM</mark> Z <u>vs</u> . HRZ	3
Replacing RIF with RPT and extending dosing interval (in continuation phase)	HP(1/7) cont phase <u>vs</u> . HR(2/7)	2
Comparing INH+RIF+PZA+EMB with PMD+MXF+PZA	PaMZ <u>vs</u> . HRZ(E)	8

#### **Summary points**

- An initial step to address the "translational gap" is to learn what data from what models analyzed in what way best inform key trial design decisions.
- Evidence-based validation of pre-clinical models is important:
  - to confidently place preclinical models on the critical development path,
  - to increase the efficiency of regulatory interactions,
  - to set a precedent for objective, data-driven processes to apply to other models (e.g., C3HeB/FeJ mouse, marmoset), and
  - to identify gaps in knowledge & in existing tools to drive future research.
- Evaluation of sterilizing mouse models is the appropriate first step for *in vivo* models, with other models to follow

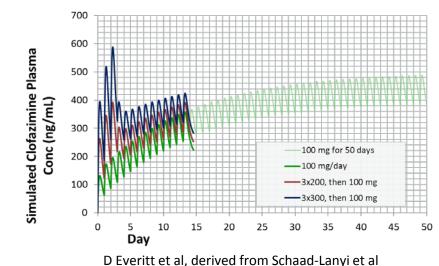
#### Clofazimine has no EBA in TB patients

Serial sputum colony counts over 1st 14 days of treatment

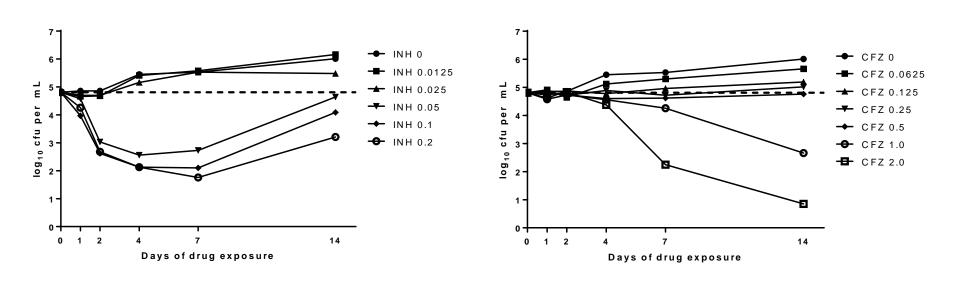


## Compartmentalized activity of CFZ and slow onset of effect – PK/distribution

- >1 month to reach steady state
- Slow, steady accumulation in adipose tissue & macrophages
  - pH-dependent ion trapping in lysosomes

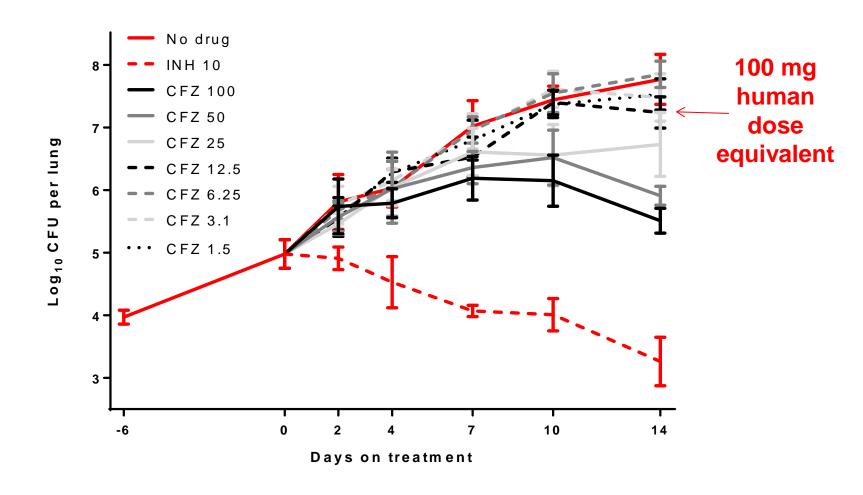


# In vitro EBA of INH and CFZ at similar multiples of their MICs

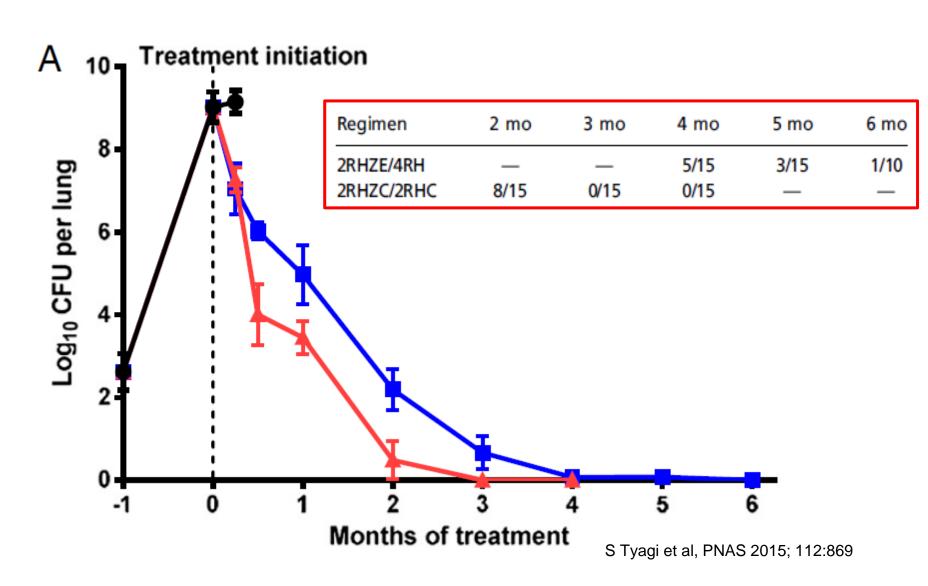


 $\circ$  MICs: INH = 0.05 μg/mL, CFZ = 0.25 μg/mL

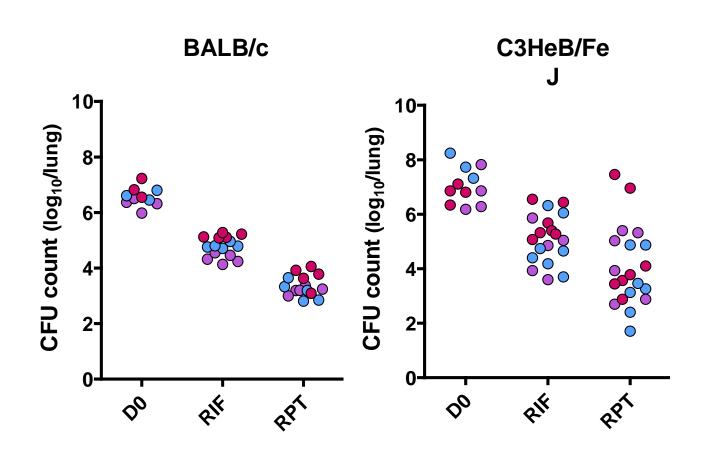
#### EBA of INH and CFZ in BALB/c mice



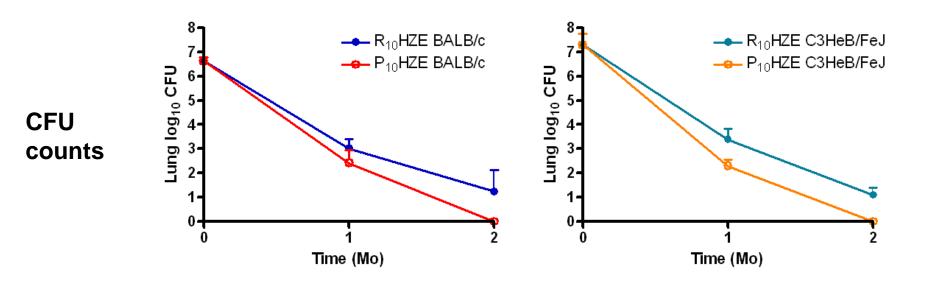
## Incorporation of CFZ into the 1<sup>st</sup>-line regimen in mice



## Comparative activity of RIF and RPT in BALB/c and C3HeB/FeJ mice over 4 wks of treatment



#### RHZE vs. PHZE in 2 mouse strains

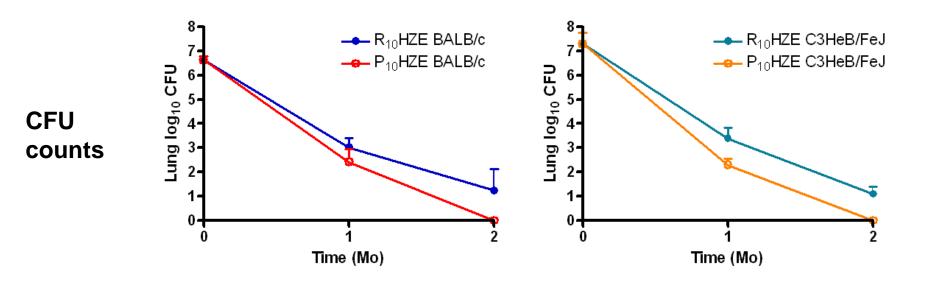


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	Mouse strain	% (proportion) of mice with relapsing after treatment for:				
Drug regimen		1 month	2 months	3 months	4 months	
R <sub>10</sub> HZE	BALB/c	ND	100% (15/15)	47% (7/15)	13% (2/15)	
	C3HeB/FeJ	ND	100% (15/15)	86% (12/14)	7% (1/15)	
P <sub>10</sub> HZE	BALB/c	100% (14/14)	7% (1/15)	0% (0/15)		
	C3HeB/FeJ	100% (13/13)	21% (3/14)	33% (5/15)		

2 month shortening with P in BALB/c

#### RHZE vs. PHZE in 2 mouse strains



#### Relapse

_		% (proportion) of mice with relapsing after treatment for:				
9	Mouse strain	1 month	2 months	3 months	4 months	
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< 1 month shortening with P in

Rosenthal et al, AAC 2012; 56:4331

#### PZA PK/PD in non-clinical models

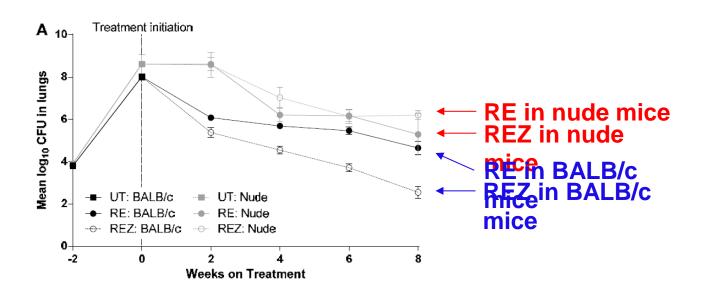
- AUC/MIC correlates best with activity<sup>1</sup>
- AUC associated with a -0.11 CFU/day reduction:
  - Hollow fiber system = 1500  $\mu$ g-h/ml<sup>1</sup>
  - BALB/c mouse =  $323 \mu g-h/ml^2$
- More potent effect of PZA in mice is likely due to lower pH (≤ 5) inside mature phagosomes of activated macrophages³ vs. that in the HFS-TB (5.8) which effectively reduces the PZA MIC by ~10x
- Increasing current dose by 2-4x increases kill rate<sup>1,2</sup>

<sup>&</sup>lt;sup>1</sup>Gumbo et al, AAC (2009); 53:3197

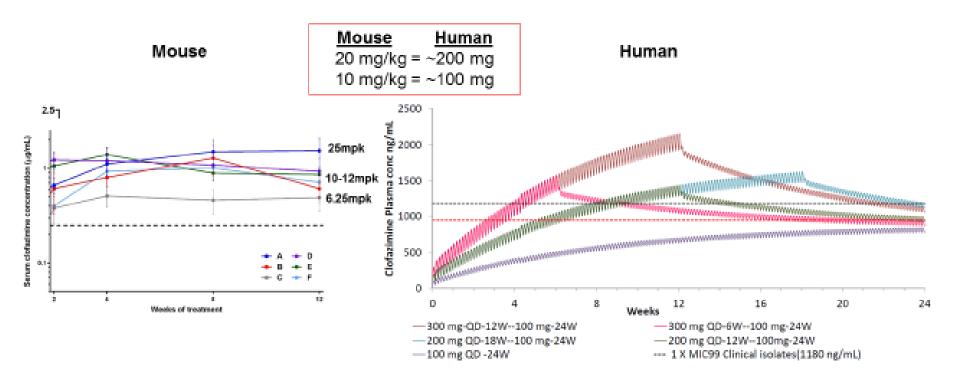
<sup>&</sup>lt;sup>2</sup> Lanoix et al, AAC (2016); 60:735

<sup>&</sup>lt;sup>3</sup> Vandal et al, Nat Med 2008; 14:849

# PZA (Z) is relatively ineffective in immunocompromised mice



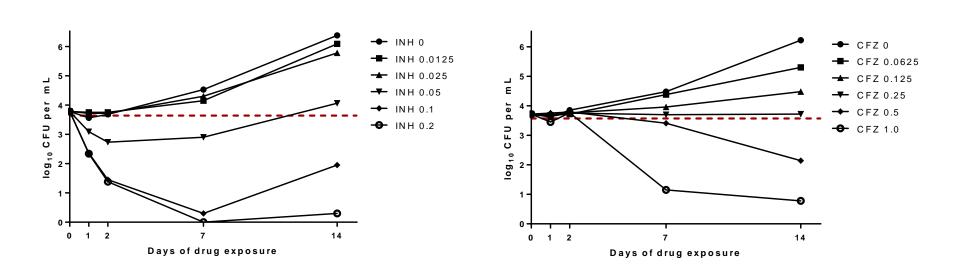
#### Clofazimine PK: mouse vs. human dose equivalence



Swanson et al, AAC 2015; 59:3042

From Ganesan S, Sunkara G, McNeeley D and Hughes D. Novartis. UNION Congress. Cape Town 2015.

# In vitro EBA of INH and CFZ at similar multiples of their MICs



 $\circ$  MICs: INH = 0.05 μg/mL, CFZ = 0.25 μg/mL