

# Overview of Analytical Validation of Donor Screening Tests

#### Krishna (Mohan) V. Ketha, Ph.D.

Division of Emerging and Transfusion Transmitted Diseases Office of Blood Research and Review CBER



#### **Presentation Outline**

- Overview of analytical validation
  - Why, When, and What
- General requirements for IVD analytical validation
  - Analytical sensitivity, specificity, precision, reproducibility and repeatability, interference, etc.
- Considerations and review issues for IVDs used for infectious disease screening
  - Study design, controls, standards

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#### Why:

To demonstrate that the manufactured product meets its prescribed requirements for safety and effectiveness

- Preclinical and analytical both are being discussed here
- Analytical performance is as critical as clinical performance

#### When:

Preferably before the IND Definitely before the final submission!



# "What" to Perform (1)

- Setting the blank
- Setting the cut-off
- Demonstration of the dynamic range
- Setting the calibration curve
- Setting the Positive and Negative Controls



# "What" to Perform (2)

- Rationale and demonstration of a re-test algorithm, where applicable
- Linearity (quantitative and semi-quantitative assays)
- Establishment of gray zone where applicable
  - To demonstrate true positives/negatives
- Sample and matrix suitability, including analyte stability



## **Regulations, Guidance, and Standards**

- **21 CFR 58** Good Laboratory Practice for Nonclinical Laboratory Studies.
- **Guidance for Industry and FDA Staff** Statistical Guidance on Reporting Results from Studies Evaluating Diagnostic Tests. March 2007.
- **CLSI EP05-A3** Evaluation of Precision of Quantitative Measurement Procedures; Approved Guideline. Third Edition. October 2014.
- **CLSI EP06-A** Evaluation of the Linearity of Quantitative Measurement Procedures: A Statistical Approach; Approved Guideline. April 2003.
- **CLSI EP07-A2** Interference Testing in Clinical Chemistry; Approved Guideline, Second Edition. November 2005.
- **CLSI EP09-A3** Measurement Procedure Comparison and Bias Estimation Using Patient Samples; Approved Guideline, Third Edition. August 2013.
- **CLSI EP12-A2** User Protocol for Evaluation of Qualitative Test Performance; Approved Guideline, Second Edition. January 2008.
- CLSI EP17-A2 Evaluation of Detection Capability for Clinical Laboratory Measurement Procedures; Approved Guideline, Second Edition. June 2012. (Replaced CLSI EP17-A2 Protocols for Determination of Limits of Detection and Limits of Quantitation; Approved Guideline. October 2004.)
- **CLSI EP25-A** Evaluation of Stability of *In Vitro* Diagnostic Reagents; Approved Guideline. September 2009.
- ICH Guideline: Validation of Analytical Procedures: Text and Methodology Q2(R1), November 2005.



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# Analytical Validation Establishes Device Performance

- Precision
- Reproducibility
- Analytical sensitivity & specificity
- Cross reactivity and interference
- Matrix comparison
- Measuring range, reference range
- Stability studies



## **Precision and Reproducibility**

- Evaluates how well the assay yields the same result on repeated determinations.
- Statistically valid approach to evaluate multiple aliquots, multiple lots at multiple sites, via multiple runs on multiple days, etc.
  - Intra- and intra-assay variability
  - Intra- and inter-lot variability
  - Inter-operator variability
  - Inter-instrument variability, if needed
- Other assay-critical, or system-critical variables (e.g., plate A/plate B when there is a specific order recommended)
- Total variability



## **Analytical Sensitivity**

Definition: "slope of the calibration curve"; capacity of a test method to differentiate between two very close concentrations of an analyte (CLSI EP17-A2)

#### Study design

- End-point dilutions
- Contrived specimens as needed
- > 3 concentrations of the analyte/panels
- Multiple replicates

Analytical Sensitivity **≠** Limit of Detection (LoD)

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# **Limit of Detection (LoD)**

**L(LoD)**: the lowest concentration of an analyte that can be consistently detected (typically in  $\geq$  95% of samples tested) (CLSI EP17-A2)

#### Study design

- Known-positive or standards/panels
- 5 dilutions/panel
- At least 20 replicates/panel (include non-reactives)
- Statistical analysis: > 95% reactivity

## Analytical Specificity/Interference Testing/Cross-Reactivity



Cause of significant difference in the test result due to the effect of another component or property of the sample (CLSI EP07-A2)

- Samples to test for cross-reactivity
  - Other species/serotypes/genetic variants
  - Other disease conditions (autoimmune, infections, etc.)
- Sample size variable for different conditions
- Interference testing
  - Endogenous (albumin, bilirubin, hemoglobulin, lipid, IgG)
  - Exogenous interferents (various drugs/supplements)

# Matrix Comparison

- Matrices claimed: whole blood, plasma, and serum
- Lysed/prepared specimen vs neat/diluted
- Different anticoagulants
- Cadaveric claims
  - Analytical sensitivity
  - Analytical specificity
  - Reproducibility

## **Stability**

- Samples
  - Room temperature, refrigerated, or frozen
  - Neat, pooled, prepared (lysed), on-board

#### • Kit

- Calibrators, and controls
- Real-time: basis for shelf-life claims
- Open kit on-board

#### Labeling claim

- Test one time point beyond the proposed claim for shelf-life
- Based on data, not extrapolated or interpolated points
- Based on real-time stability, not accelerated studies

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# **Complex Donor Screening IVDs**

- Diverse IVDs with different final analytes
  - Antigen, antibody, nucleic acid
- Technology
  - EIA, chemiluminescence, PCR, transcription-mediated amplification (TMA)
- Different limit of detection (LoD) parameters
  - g/mL, IU/mL, copies/mL, iRBC/mL
- Different clinical samples/sample size
  - Genetic variants, prevalence/risk of transfusion-transmission (TT)



## **Precision and Reproducibility**

- Panel of 6-10 well-characterized specimens, representing a clinically relevant range:
  - Minimum of one positive and negative sample near assay cut-off
  - Assay controls and calibrators
- Different group/panel of specimens
  - Each type of specimen matrix
  - Each analyte
  - Genotype (or variant)
- Tested using at least three kit lots

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# **Analytical Sensitivity (1)**

- Each specimen group, genotype or strain
- Each sample matrix (e.g., serum, EDTA-plasma)
- Approaches
  - End-point dilution
  - Earliest time of reactivity in serially-collected specimens
  - Comparison to reference standards
  - Comparison to an independent method
  - Quantitative biochemical characterization
- Direct comparison to a FDA-licensed, approved, or cleared test
- Controls targeted to clinical decision points
  - Low positive between 1-3 S/CO or 1-3 x LoD
- Validation of assay's gray zone(s)



# **Analytical Sensitivity (2)**

- Appropriate standards or CBER reference panels e.g., HIV, HCV, HBsAg, *Babesia*
- Seroconversion panels, when available

   e.g., multiple specimens from at least 10 subjects undergoing
   seroconversion)
- Low-titer panels for each strain/analyte/matrix, if applicable e.g., 6-10 specimens per panel
- Dilution series

e.g., at least 10 specimens from 10 subjects for each strain/matrix

- Known-positives from relevant populations e.g., samples from an HIV-1 high risk group
- NAT: confirm sequence identity for strains/genotypes claimed

# **Analytical Sensitivity (3)**

- FDA prepares and provides panels of samples
  - Different panels for all analytes
  - Analytes at various levels
- Not to be confused with lot release panels
- Number of samples correctly detected is evaluated



#### **Seroconversion Panels**

- Panels collected from plasmapheresis donors who are in the process of seroconverting
- Real clinical samples from blood donors with values near the cut-off are rare
- Seroconversion panels are real samples with analytes at relevant clinical concentrations



# **Analytical Specificity**

- Samples to include
  - Other strains/variants confirm identity
  - Other disease/medical conditions (autoimmune, infections)
  - Potentially interfering substances
  - Endogenous (albumin, bilirubin, hemoglobulin, IgG, etc.)
  - Exogenous interferents (various drugs/supplements)
  - Different anticoagulants/collection tubes
- Sample size variable for different conditions
- Labeling claim: include interference/cross-reactivity



## **Device Performance - What to Submit**

- Summaries of study designs
  - Materials, procedures, analysis, and oversight
  - Sample collection, selection criteria, handling, and storage
  - Statistical and clinical considerations
  - Documentation that all testing performed at an approved facility using Good Laboratory Practice (GLP)
- Summaries of results and line data for all studies
  - Data for each specimen
  - Each assay run performed, including failed runs
  - Documentation and justification of excluded data
  - Documentation and justification of deviations, outliers, etc.

#### **Common Review Issues**



- Results don't meet pre-specified acceptance criteria
- Inconsistent definition of LoD
- Validation not performed on final device (including algorithm and cut-off)
- Insufficient samples around the cut-off
- Intended Use too broad (such as "for infectious diseases testing")
- No definition of guard bands
- Gray zone included in final device
- Not all claims are validated

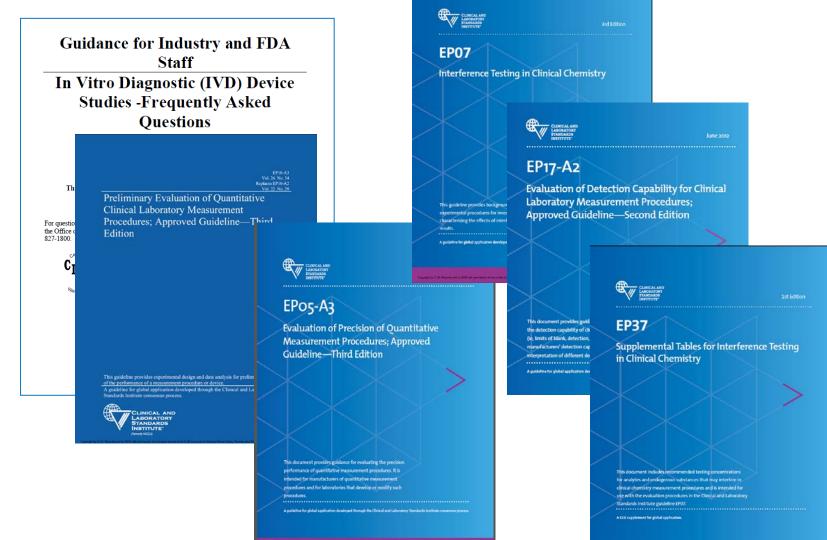


#### **Summary**

- Analytical studies = Foundational studies
- Device performance final results that are precise with high sensitivity and specificity, reproducible across variables, demonstrating no effect of interferents



#### References





#### Thanks!

## Krishna (Mohan) Ketha krishna.ketha@fda.hhs.gov