# EMERGENCY USE AUTHORIZATION (EUA) SUMMARY CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay Clinical Research Sequencing Platform (CRSP), LLC at the Broad Institute of MIT and Harvard

For *In vitro* Diagnostic Use
Rx Only
For use under Emergency Use Authorization (EUA) only
Updated June 10, 2021

The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay will be performed at the Clinical Research Sequencing Platform (CRSP), LLC at the Broad Institute of MIT and Harvard located at 320 Charles Street, Cambridge, Massachusetts 02141, which is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. §263a, as described in the laboratory procedure that was reviewed by the FDA under this EUA.

#### **INTENDED USE**

The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1) is intended for the qualitative detection of nucleic acid from SARS-CoV-2 in nasopharyngeal, oropharyngeal, anterior nasal (nasal), and mid-turbinate swabs, nasopharyngeal wash/aspirate and nasal aspirate specimens, and bronchoalveolar lavage specimens from individuals suspected of COVID-19 by their health care provider.

Testing is limited to the Clinical Research Sequencing Platform (CRSP), LLC at the Broad Institute of MIT and Harvard, located at 320 Charles Street, Cambridge, MA 02141 which is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. §263a, and meets requirements to perform high complexity tests.

Results are for the detection and identification of SARS-CoV-2 RNA. The SARS-CoV-2 RNA is generally detectable in respiratory specimens during the acute phase of infection. Positive results are indicative of the presence of SARS-CoV-2 RNA; clinical correlation with patient history and other diagnostic information is necessary to determine patient infection status. Positive results do not rule out bacterial infection or co-infection with other viruses. The agent detected may not be the definite cause of disease.

Laboratories within the United States and its territories are required to report all results to the appropriate public health authorities.

Negative results do not preclude SARS-CoV-2 infection and should not be used as the sole basis for patient management decisions. Negative results must be combined with clinical observations, patient history, and epidemiological information.

The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay is intended for use by qualified clinical laboratory personnel specifically instructed and trained in the techniques of real-time PCR and in vitro diagnostic procedures. The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay is only for use under the Food and Drug Administration's Emergency Use Authorization.

#### DEVICE DESCRIPTION AND TEST PRINCIPLE

The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay is a reverse transcriptase real-time polymerase chain reaction (rRT-PCR) assay for the qualitative detection of SARS-CoV-2 specific RNA. Version 1 of this test uses primer/probe sets developed by the CDC that target two viral gene targets in the Nucleocapsid gene of SARS-CoV-2, N1 and N2, and an internal control gene, RNase P (RP).

The test consists of four processes in a single assay: 1) nucleic acid extraction, 2) reverse transcription of target RNA to cDNA, 3) PCR amplification of target and internal control DNA, and 4) simultaneous detection of PCR amplicons by fluorescent dye labelled probes.

<u>Assay Version 1</u> uses CDC developed SARS-CoV-2 nucleocapsid N1, N2, and human RP primers and probes. The respiratory specimen types are transported in VTM or sterile saline. Assay Version 1 was validated for use with the Applied Biosystems Viia7 thermocycler or the Applied Biosystems QuantStudio 7 Flex each with QuantStudio software version 1.3.

**Table 1.** Attributes of the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay Version 1.

rissay version i.	V1
Transport medium	VTM or 0.9% sterile saline
Extraction method	ThermoFisher MagMAX Viral RNA Isolation Kit
Extraction format	96-well
Liquid handler for extraction	Agilent Bravo
Extraction sample input volume	50ul into extraction
Extraction output volume	50μ1
Liquid handler for RI-PCR plate set-up	Formulatrix Tempest and Agilent Bravo
RT-PCR template volume	5μl
RT PCR total reaction volume	15μ1
RT-PCR plate format	384-well
Thermocycler	Viia7 or QuantStudio 7 Flex
Thermocycler software version	QuantStudio software version 1.3

#### INSTRUMENTS USED WITH TEST

The CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay is to be used with the following instrumentation:

- Specimen Lysis/RNA Extraction/Realtime PCR Reagent Preparation: Agilent Bravo Liquid Handling Platform (running VWorks (Build 11.4.0.1233)) for automated extraction and/or RT-PCR plate set-up, and Formulatrix Tempest and Agilent Bravo Liquid Handling Platform also for RT-PCR plate set-up.
- RT-PCR Platforms: Applied Biosystems QuantStudio 7 Flex Real-Time PCR System (QuantStudio Software V1.3), Applied Biosystems ViiA7 (QuantStudio Software V1.3).

**Table 2. REAGENTS AND MATERIALS** 

Item	Vendor	Vendor Catalog Number
4X TaqPath, 1-Step RT-qPCR Master Mix, GC (2,000 rxns/kit)	Thermo Fisher Scientific	A15300
2019-nCoV CDC EUA Primer/Probe Kit (500 rxns/box)	IDT	10006606
Tip, 1000ul Fltr, Conductive(480/PK)	Hamilton Robotics, Inc.	235905
Tip, 70ul, St. Fltr, Cond, Bravo (3840/PK)	AGILENT TECHNOLOGIES INC	19133-142
Plate, 384W, DW, V-Bottom(60/CA)	VWR International, LLC	82051-320
Plate, 384w Twintec Clear 40ul(25/BX)	VWR International, LLC	12000-658
Plate, 96w Twintec Clear (25/BX)	VWR International, LLC	47744-116
Plate, 384w Clear, FB, NS (100/CA)	VWR International LLC	82051-298
AB-3720 Thermo foil pierce-able seal	Thermo Fisher Scientific	AB-3720
Ethanol, 100%, (24pints/CA)	VWR International, LLC	89125-170
Thermo Viral RNA Isolation Kit	Thermo Fisher Scientific	AMB18365
Isopropanol, 99% (4x4L/case)	VWR International, LLC	TXMK303216BRI
Wipe, RNase Zap Ambion (100/PK)	Thermo Fisher Scientific	AM9786
DNA ZAP!, Degradation Solution(250ml/BT)	Life Technologies, Inc.	AM9890
Disinfectant, Germicidal (625ml/BT)	WW GRAINGER CO	3VDL4
Ethanol, 70% USP 140Proof (32oz/BT)	VWR International, LLC	76212-358
Kit,2019-nCoV N_Positive Control(1/bx)	Integrated DNA Technologies, Inc.	10006625
Tube, 50ml orange cap (25/PK)	VWR International, LLC	21008-775
Super rags (250/case)	BLUE THUNDER TECHNOLOGIES, INC.	WI-1318Q.250
Bags, Heavy weight, 8"x12" (100/pack)	VWR International, LLC	11215-280
Tip, 2ml St. Fltr LTS(480/CA)	Mettler-Toledo Rainin LLC	17002923
Pipette, 25ml Serological, St(200/CA)	VWR International, LLC	53392-198
Tip, pipette1000ul LTS Rainin (768/Ca)	Mettler-Toledo Rainin LLC	30389212
AB-3720 Thermo foil pierceable seal	Thermo Fisher Scientific	AB-3720
Kit, 2019-nCoV CDC EUA(500Rxn/BX)	Integrated DNA Technologies, Inc.	10006606
TaqPath,1Step RTqPCR MtrMx,CG(2000rx/KT)	Life Technologies, Inc.	A15300
Water, Sterile, Nuclease-Free (1000ml/BT)	VWR International, LLC	10220-384
DNA ZAP!, Degradation Solution(250ml/BT)	Life Technologies, Inc.	AM9890

Item	Vendor	Vendor Catalog Number
Wipe, RNase Zap Ambion (100/PK)	Life Technologies, Inc.	AM9786

# CONTROLS TO BE USED WITH THE CRSP SARS-C<sub>0</sub>V-2 REAL-TIME REVERSE TRANSCRIPTASE (RT)-PCR DIAGNOSTIC ASSAY

- A "no template" (negative) control (NTC) is used for every run and is needed to confirm that there is no contamination for the assay.
- A positive template control (COVID-19\_N\_Postive, IDT, #10006625) targeting the SARS-CoV-2 N-gene (N1 and N2) is used for every run and is needed to confirm that the assay is completed by the intended design.
- An internal control primer/probe set, targeting the human RNase P gene, is used for every
  patient sample to confirm appropriate specimen collection and to monitor the integrity of
  nucleic acid extraction and RT-PCR reactions.
- A human specimen (HSC) extraction control is included in each run to test for failure in lysis and extraction and potential contamination during extraction.

#### INTERPRETATION OF RESULTS

All test controls should be examined prior to interpretation of patient results. If the controls are not valid, the patient results cannot be interpreted (Refer to Table 3 for a summary of expected control results).

# 1. COVID-19 RT-PCR Test controls - Positive, Negative, Extraction, and Internal:

Controls should produce the results outlined in Table 3, below.

**Table 3**. Expected Control Results for the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1)\*

Control Location	Ni	N2	RP	Result Interpretation	Action
NTC well			-	Plate passes NTC QC	Plate sent for review and reporting
NTC well	A	Any target positive	e	Plate Fails NTC QC	Plate reworked from RNA extraction
HSC well		-	+	Plate passes Extraction QC	Plate sent for review and reporting
HSC well	Any targo	et positive	+/-	Plate fails Extraction QC	Plate reworked from RNA extraction
HSC well	-	-	-	Plate fails Extraction QC	Plate reworked from RNA extraction
nCoVPC	+	+	-	Plate passes Assay QC	Plate sent for review and reporting

Control Location	N1	N2	RP	Result Interpretation	Action
nCoVPC	If ≤1 target is positive		-	Plate fails Assay QC	Plate reworked from RNA extraction

<sup>\*</sup>Note: The results of the assay are reported according to the following categories where a + indicates a Ct of<40 and a - indicates a Ct of>40 or undetermined.

# 2. Examination and Interpretation of Patient Specimen Results:

Assessment of clinical specimen test results should be performed after the positive and negative controls have been examined and determined to be valid and acceptable. If the controls are not valid, the patient results cannot be interpreted. Please see **Table 4** for guidance on patient specimen result interpretation and reporting of results.

Table 4. CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR

Diagnostic Assav (Version 1)\*

Diagnostic rissay (	v Cibion i j		
2019 nCoV_N1	2019 nCoV_N2	RP	Result Interpretation
+	+	H	SARS-CeV-2 detected
If at least one tar	get is positive	H	Inconclusive
-	-	+	SARS-CoV-2 not detected
-		_	Invalid Result

<sup>\*</sup>Note: The results of the assay are reported according to the following categories where a + indicates a Ct of<40 and a - indicates a Ct of>40 or undetermined.

# PERFORMANCE E VALUATION

# Analytical Sensitivity - Limit of Detection (LoD):

The LoD for Assay Version I SARS-CoV-2 detection was determined using dilutions of patient nasopharyngeal samples previously determined to be positive by the Massachusetts State Public Health Laboratory (MSPHL) CDC EUA assay. The SARS CoV-2 copy number in each patient sample was estimated using a relative standard curve generated by diluting SARS-CoV-2 synthetic RNA of a known concentration from Twist Biosciences (Cat no. MN908947.3, SKU: 102024). Each patient sample was diluted in VTM, independently extracted, and analyzed on the Applied Biosystems Viia7 thermocycler. Twenty of twenty replicates were detected at 4.0 x 10<sup>3</sup> copies/mL dilution (**Table 5**).

**Table 5.** CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1) LoD Confirmation Study.

Concentration	# Positive Replicates/# Total Replicates
$4.0 \times 10^3$ copies/mL	20/20

### **Analytical Sensitivity – Inclusivity:**

The sequences for the N1 and N2 primers/probes used in this assay are identical to the primer/probe sequences used in the FDA emergency use authorized CDC 2019-Novel Coronavirus (2019-nGoV) Diagnostic Panel. CDC has provided a right of reference to their Inclusivity Study data, which is available at <a href="https://www.fda.gov/media/134922/download">https://www.fda.gov/media/134922/download</a>.

#### <u>Analytical Specificity – Cross-Reactivity:</u>

## In-silico Cross-Reactivity Assessment

The sequences for the N1 and N2 primers/probes used in this assay are identical to the primer/probe sequences used in the FDA emergency use authorized CDC 2019-Novel Coronavirus (2019-nCoV) Diagnostic Panel. CDC has provided a right of reference to their Cross-Reactivity Study data, which is available at <a href="https://www.fda.gov/media/134922/download">https://www.fda.gov/media/134922/download</a>.

## **Clinical Evaluation:**

Performance of the CRSP SARS-CoV 2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1) was evaluated using the Applied Biosystems Viia7 thermocycler and residual specimens collected from individual patients. Thirteen positive oropharyngeal (OP) swabs, 10 positive nasopharyngeal (NP) swabs 10 positive NP/OP swabs, and 40 negative NP swabs were all previously tested by Massachusetts State Public Health Laboratory (MSPHL) using the CDC EUA authorized SARS-CoV-2 test. The results from the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1) had 100% agreement with the expected results for the 73 samples compared to the EUA authorized SARS-CoV-2 comparator (**Table 6**).

**Table 6**. Summary of Clinical Performance of the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1) Compared to MSPHL CDC EUA Assay.

NP, OP, and NP/OP Specimens		EUA Comparator Assay			
		Positive	Negative	Total	
CRSP SARS-	Positive	33	0	33	
CoV-2 Assay VI	Negative	0	40	40	
CUV-2 Assay W1	Total	33	40	73	
Positive Agree	ement	100% (33/33), 95%	6 CI: (89.6, 100%)		
Negative Agre	ement	100% (40/40), 95% CI: (91.2, 100%)			

#### **LIMITATIONS**

- The use of this assay as an *in vitro* diagnostic under the FDA Emergency Use Authorization (EUA) is limited to Clinical Research Sequencing Platform, LLC at the Broad Institute of MIT and Harvard, Cambridge, MA which is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA), 42 U.S.C. § 263a, and meets requirements to perform high complexity tests. Use of this assay is limited to personnel who are trained in the procedure. Failure to follow these instructions may result in erroneous results.
- The performance of the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay Version 1 was established using nasopharyngeal (NP), oropharyngeal (OP), and combined nasopharyngeal/oropharyngeal (NP/OP) swabs in viral transport media (VTM) or 0.9% sterile saline nasal swabs, mid-turbinate nasal swabs, nasopharyngeal wash/aspirates, nasal aspirates, and bronchoalveolar lavage specimens are also considered acceptable specimen types for use with the CRSP SARS-CoV-2 Real-time Reverse Transcriptase (RT)-PCR Diagnostic Assay (Version 1), but the performance has not been established with these specimens.
- Samples must be collected, transported, and stored using appropriate procedures and conditions. Improper collection, transport, or storage of specimens may hinder the ability of the assay to detect the target sequences.
- The clinical performance has not been established in all circulating variants but is anticipated to be reflective of the prevalent variants in circulation at the time and location of the clinical evaluation. Performance at the time of testing may vary depending on the variants circulating, including newly emerging strains of SARSCoV-2 and their prevalence, which change over time.

#### **WARNINGS**:

- This product has not been FDA cleared or approved, but has been authorized for emergency use by FDA under an EUA for use by the authorized laboratory;
- This product has been authorized only for the detection of nucleic acid from SARS-CoV-2, not for any other viruses or pathogens, and
- The emergency use of this product is only authorized for the duration of the declaration that circumstances exist justifying the authorization of emergency use of *in vitro* diagnostics for detection and/or diagnostics of COVID-19 under Section 564(b)(1) of the Federal Food, Drug, and Cosmetic Act, 21 U.S.C. § 360bbb-3(b)(1), unless the declaration is terminated or authorization is revoked soorier.