

Focused On Results. CERTIFICATE OF ANALYSIS

Job Name: Task 3 - Analysis of Official Samples
Job Location: 1st Group - 8 Samples
Job Number: CLIN 1 - Task 3 (8 Samples)
PO Number: HHSF223201810337P

Client: US Food & Drug Adminitration
Address: Office of Cosmetics & Colors
4300 River Road
College Park, MD 20740

Chain of Custody: 300396

Attention: John Gasper

Date Submitted: 3/14/2019

Date Analyzed: 3/29/2019 - 4/18/2019

Report Date: 4/25/2019

Date Sampled: Not Provided

Person Submitting: Steve Wolfgang Revised: 4/30/2019 (1st Revision)

SUMMARY OF ASBESTOS IN TALC ANALYSIS

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Comments																								
L	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9
% e Other	88.8%	92.4%	89.8%	80.89	68.5%	%9.79	70.6%	72.1%	72.0%	74.7%	72.5%	73.0%	72.5%	72.8%	73.4%	74.1%	75.8%	75.8%	8.96	97.5%	97.4%	33.7%	36.4%	29.3%
% Acid Soluable	2.0%	1.4%	4.0%	3.3%	2.1%	3.3%	4.7%	4.5%	3.9%	12.4%	13.8%	15.5%	3.5%	3.0%	2.5%	9.7%	5.3%	%0.9	3.1%	2.4%	2.5%	11.0%	8.3%	15.4%
% Organics	6.2%	6.2%	6.2%	28.7%	29.4%	29.1%	24.7%	23.4%	24.0%	12.8%	13.7%	12.5%	24.0%	24.1%	24.1%	19.2%	18.9%	18.3%	%0.0	0.1%	%0.0	55.3%	55.3%	55.3%
% Asbestos by PLM	ND	QN	ND	ND	QN	ND	QN	ND	ND															
% Total Asbestos a by TEM Using ASTM D5756 Mass Calculation	ND	< 0.00080%	0.00030%	0.00574%	< 0.00013%	0.00371%	ND																	
% Chrysotile by TEM Using ASTM D5756 Mass Calculation	ND	< 0.00080%	0.00030%	0.00503%	< 0.00013%	0.00005%	ND																	
% Tremolite by TEM Using ASTM D5756 Mass Calculation	ND	0.00071%	< 0.00013%	0.00367%	ND																			
TEM LOQ Using ASTM D5756 Mass Calculation	0.00000872%	0.00000648%	0.00000574%	0.00000769%	0.00000819%	0.00000773%	0.00001016%	0.00080274%	0.00001479%	0.00000536%	0.00012905%	0.00000671%	0.00000751%	0.00000454%	0.00000599%	0.00000599%	0.00000714%	0.00000629%	0.00000536%	0.00000694%	0.00000539%	0.00000524%	0.00000721%	0.00000540%
TEM LOD Using ASTM D5756 Mass Calculation	0.00000218%	0.00000162%	0.00000144%	0.00000192%	0.00000205%	0.00000193%	0.00000254%	0.00000285%	0.00000370%	0.00000134%	0.00000188%	0.00000168%	0.00000188%	0.00000114%	0.00000150%	0.00000150%	0.00000178%	0.00000157%	0.00000134%	0.00000173%	0.00000135%	0.00000131%	0.00000180%	0.00000135%
Client Sample ID	D-32	D-32	D-32	D-33	D-33	D-33	D-34	D-34	D-34	D-35	D-35	D-35	D-36	D-36	D-36	D-37	D-37	D-37	D-38	D-38	D-38	D-39	D-39	D-39
AMA Sample ID S	300396-1	300396-1A	300396-1B	300396-2	300396-2A	300396-2B	300396-3	300396-3A	300396-3B	300396-4	300396-4A	300396-4B	300396-5	300396-5A	300396-5B	300396-6	300396-6A	300396-6B	300396-7	300396-7A	300396-7B	300396-8	300396-8A	300396-8B

Analytical Method(s): PLM by Modified NY ELAP 198.6

LOD = Limit of Detection

TEM by Modified NY ELAP 198.4/ASTM D5756

(b) (6)

Analyst(s): PLM

Technical Director: Andreas Saldivar

TEM = Transmission Electron Microscopy

PLM = Polarized Light Microscopy

ND = Not Detected

LOQ = Limit of Quantification

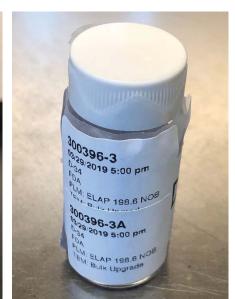
All results are to be considered preliminary and subject to change unless signed by the Technical Director or Deputy

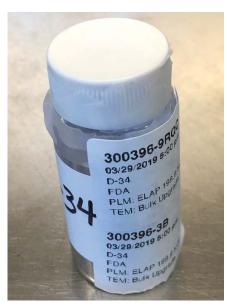
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300396-3, 3A, 3B/D-34





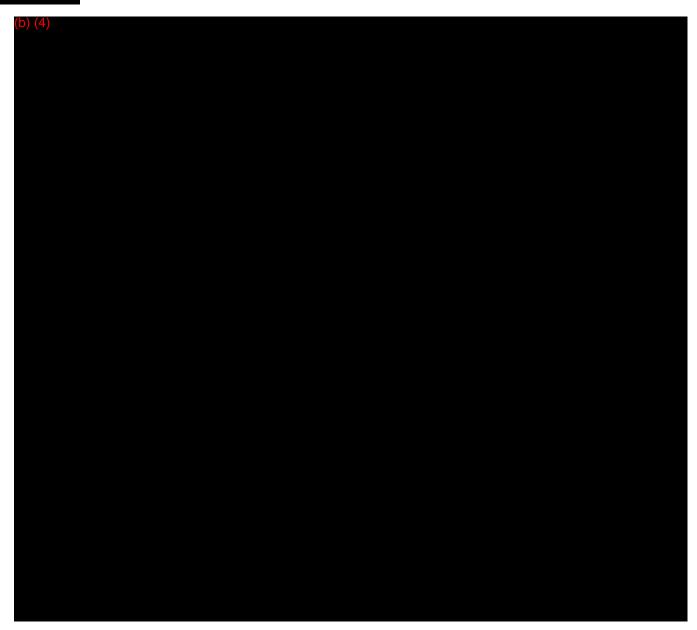








(b) (4)



Sample Preparation

Samples were prepared for PLM and TEM bulk analysis by 6 on March 15, 2019 through March 29, 2019. Sample preparation consisted of the following steps:

- 1) Label and weigh two 8mL glass vials for each sample in the set one vial for the PLM preparation and one vial for the TEM preparation.
- 2) Weigh out 0.1 to 0.8 grams of material and place in corresponding 8mL glass vial. Record weight.
- 3) Burn samples at 480° C for at least 12 hours.
- 4) Record Post-Ash Weight.
- 5) Treat ashed sample with concentrated hydrochloric acid.
- 6) Filter acid reduced material onto a pre-weighed 47mm 0.4um PolyCarbonate filter.
- 7) Place filter into drying oven for 30 minutes and then record Post-Acid Reduced weight.
- 8) Make four PLM slide preparations from the PLM residual ash for each sample in 1.550 dispersion oil. Make additional preparations in 1.605, 1.625, 1.680 and 1.700 dispersion oil as necessary for particle identification.

- 9) Weigh a portion of the residue from the TEM residual ash and place it into the corresponding pre-weighed 100ml jar.
- 10) Fill the 100ml jar with deionized water
- 11) Sonicate the jars for approximate 5-minutes.
- 12) Filter 0.2ml to 1ml of the solution onto a 47mm 0.22um MCE filter.
- 13) Dry the filter for 10 minutes then collapse, carbon coat, and place on a 3 TEM grids.

PLM Analysis

Analysis was performed in accordance with NY ELAP 198.6 protocols. The analysis was conducted using an Olympus BH-2 polarized light microscope (PLM) equipped with a dispersion staining objective. All four slide preparations for each aliquot were examined. 400-point count was performed for those samples on which asbestos was observed. If no asbestos was detected on any of the slides, the percentage of fibrous components was determined by visual estimation. The results of this analysis are detailed below in the *Discussion and Interpretation of Analytical Findings* section for each individual sample.

TEM Analysis

Analysis was performed in accordance with modified NY ELAP Method 198.4 protocols. The analysis was performed using a JEOL JEM-100CX II transmission electron microscope (TEM), equipped with a Thermo Fisher Quest Energy Dispersive X-Ray Analyzer (EDXA), at magnifications of 19,000x. Two grids for each aliquot were examined. Twenty (20) grid openings were examined per sample.

Modifications to the NY ELAP 198.4 Method were:

- 1) The residue was not placed in alcohol and prepared using the quick drop method. To obtain a more uniform preparation, the residue was placed in a jar and filled with 100ml of deionized water. The jar was sonicated, and a portion of the solution was filtered onto a 47mm 0.22um MCE filter.
- 2) The tremolite and chrysotile were not visually estimated. The length and width of the observed particles were measured, and the mass of each amphibole particle was calculated using the ASTM D5756 method.

The results of this analysis are detailed below in the *Discussion and Interpretation of Analytical Findings* section for each individual sample.

Calculations

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ASTM \ D5756 \ Mass \\ M = \pi/4 \ L^* \ W^2 * D * 10^{-12} \\ M = mass \\ L = length \\ W = width \\ D = density \\ Percent \ Calculation \\ EFA(mm^2) * 100ml * MA(g) * RW(g) \\ VF(ml) * IW(g) * AA(mm^2) * RJ(g) \\ The \ calculated \ value \ is then \ multiplied \ by 100 \ to \ convert \ it \ to \ percent. \\ EFA - Effective \ filter \ area
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MA – Mass of asbestos
RW – Weight of residue
VF – Volume filtered
IW – Initial weight of the sample
AA – Area analyzed
RJ – Weight of residue placed into the jar



Limit of Detection and Quantification

We used the mass of a 0.5 x 0.04-micron tremolite or chrysotile fiber, depending on what was found in each sample, as the basis for our calculations. Limit of detection was defined as 1 fiber and limit of quantification was defined as 4 fibers.

Some aliquots of samples D34 (b) (4) contained very small amounts of asbestos that were either at or below our 4-fiber limit of quantification. For these samples we defined our limit of quantification as follows:

300396-3A: mass of the single observed chrysotile fiber plus the mass of three tremolite fibers measuring

0.5 x 0.04 microns

300396-4A: mass of the two observed chrysotile fibers, the single observed tremolite structure plus the mass of

one 0.5 x 0.04 microns tremolite fiber.

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300396-3, 3A, 3B, Client Sample D-34

PLM

All three aliquots of sample D-34 were analyzed by (b) (6) on March 29, 2019. No asbestos or non-asbestos amphibole variants were detected the samples. The results were calculated using the equations detailed in the calculations section.

300396-3	NAD
300396-3A	NAD
300396-3B	NAD

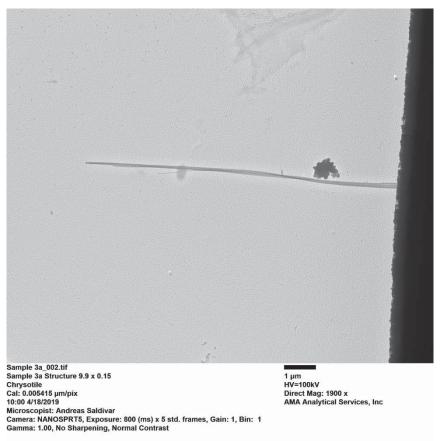
TEM

Sample 3 was analyzed by (b) (6) on April 4, 2019. Andreas Saldivar analyzed sample 3A on April 15, 2019 and sample 3B on April 16, 2019. All three samples contained mica, talc, and titanium particles. One 9.9 x 0.15 micron chrysotile bundle was counted on sample 3A. Five (5) chrysotile structures were counted on sample 3B. The results were calculated using the equations detailed in the calculations section.

300396-3	NAD
300396-3A	< 0.00080%
300396-3B	0.00030%

Below are pictures, diffraction patterns, and chemistry of the counted Chrysotile particles. The mica, talc, and titanium particles are similar to those pictured in samples 1 and 2. The unidentified peaks in chemistry spectra are copper, zinc, and carbon. Those peaks are from the TEM specimen holder and specimen grid.

Sample 300396-3A Chrysotile structure



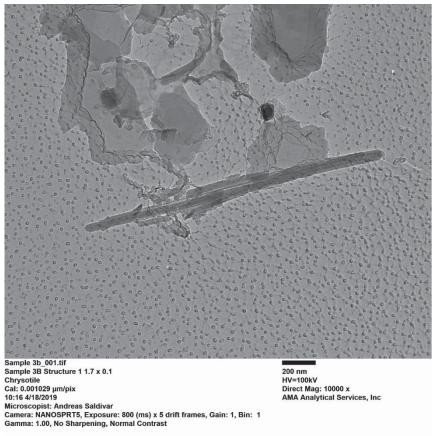
Sample 300396-3A Diffraction pattern from the chrysotile structure pictured above.



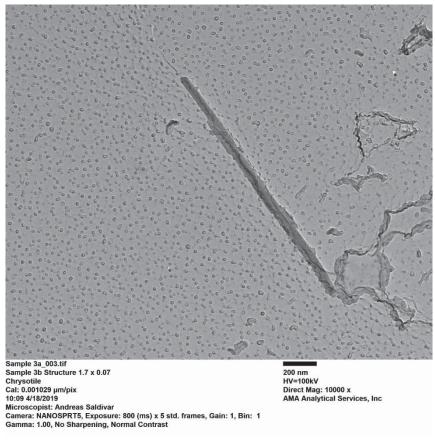
Sample 3a_001.tif
Sample 3a Structure 1
Chrysotile Diffraction
09:58 4/18/2019
Microscopist: Andreas

Microscopist: Andreas Saldivar Camera: NANOSPRT5, Exposure: 800 (ms) x 5 std. frames, Gain: 1, Bin: 1 Gamma: 1.00, No Sharpening, Normal Contrast 100 (1/Å) HV=100kV Cam Len: 0.2200 m AMA Analytical Services, Inc

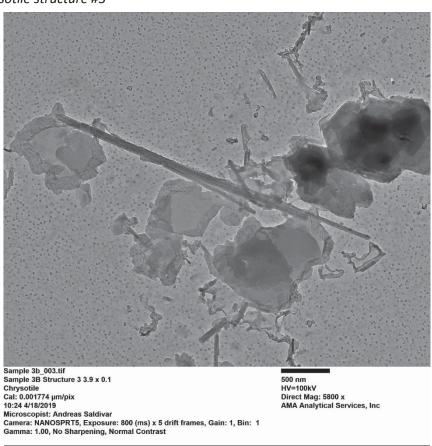
Sample 300396-3B Chrysotile structure #1



Sample 300396-3B Chrysotile structure #2



Sample 300396-3B Chrysotile structure #3



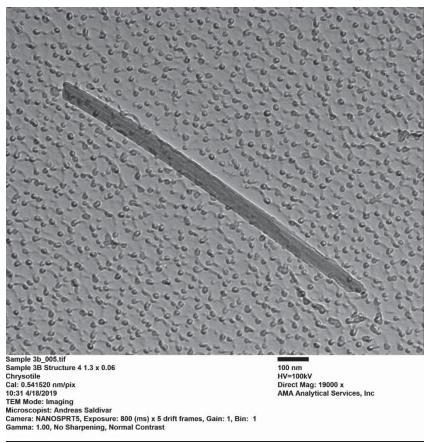
Sample 300396-3B Chrysotile diffraction pattern from structure #3



Sample 3b_002.tif Sample 3B Structure 3 3.9 x 0.1 Chrysotile Diffraction 10:21 4/18/2019 Microscopist: Andreas Saldivar

Microscopist: Andreas Saldivar Camera: NANOSPRT5, Exposure: 800 (ms) x 5 drift frames, Gain: 1, Bin: 1 Gamma: 1.00, No Sharpening, Normal Contrast 100 (1/Å) HV=100kV Cam Len: 0.2200 m AMA Analytical Services, Inc

Sample 300396-3B Chrysotile structure #4



Sample 300396-3B Chrysotile structure #5

