

# The new post-genomic field of science

- Proteins are the primary target for most therapeutics
- New biomarkers for disease, toxicity and treatment
- Determine product potency, purity and consistency
- Endpoints and release specifications for drug products
- Molecular targeted therapy and patient tailored therapy -  
*Recent examples: GLEEVEC, HERCEPTIN. These drugs target activated and disregulated proteins, not genes.*

# Regulatory Impact

- Vaccine assessment/potency
- Surrogate endpoints - efficacy/toxicity
- Quality control/quality assurance for product production
- New bioassays
- Biomarkers for early detection
- Toxicity detection and prediction

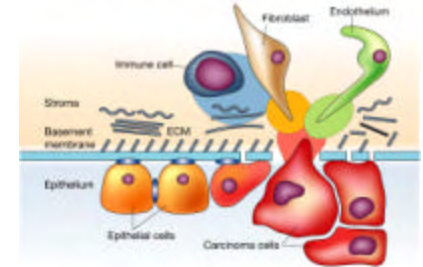
## Regulatory Impact (cont.)

- Discovery of new therapeutic targets
- Risk of disease recurrence
- Patient-tailored therapy. Prospective selection
- New paradigm in disease classification/characterization
- Proteomic-based epidemiology

# FDA views on proteomics:

- Critical component of safe and effective drug development
- Basis for new drug discovery, biomarkers and surrogate endpoints for toxicity and efficacy monitoring
- Means to detect and assess chemical and biological terrorist agents

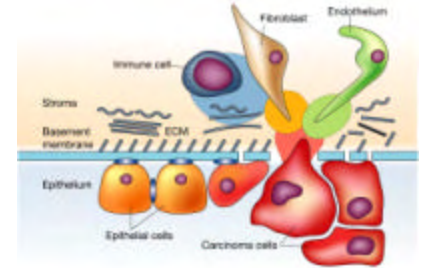
# TISSUE MICROENVIRONMENT



NATURE 2001

- Proteomic networks exist **within** the cell and **outside** the cell at the tumor-host interface
- Cancer is a proteomic disease at the functional level.
- The state of protein networks is dictated by the tissue **context** of the cell, and the local cell-cell or cell-matrix interactions
- Cell culture models may not accurately represent the **fluctuating** protein expression pattern and the **state** of protein interactions in the native tissue microenvironment

# PROTEOMIC INFORMATION



JAMA 2002

- **State of protein circuits within the cell and outside the cell at the tumor-host interface: Pathogenic role of dominant or deranged signal pathways**
- **Proteomic information content of circulating blood: Patterns of LMW proteins and peptides reflect organ pathologic states**

## **Keys to Cancer:**

- **Early Detection-**

**Development of new artificial intelligence-based bioinformatics tools for diagnostic proteomic pattern discovery**

- **Molecular Diagnostics**

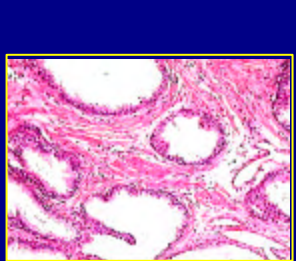
**New Target Discovery (2D-PAGE)**

**Signal Pathway Profiling (Protein Arrays)**

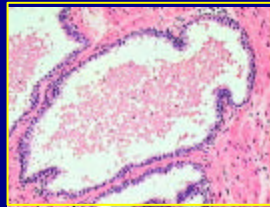
**Phosphoproteomics (Protein Arrays/ 2D-PAGE)**

- **Molecular Targeted Therapeutics**

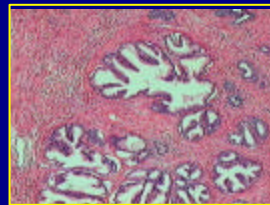
**Implementation of proteomics to ongoing NCI-based clinical trials**



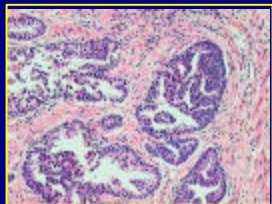
**Low-grade PIN**



**Normal**

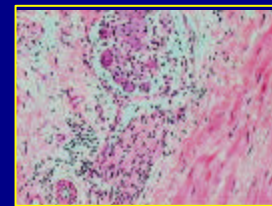


**Hyperplasia**

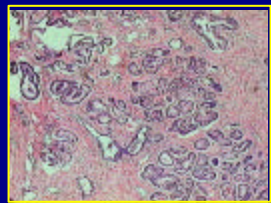


**High-grade PIN**

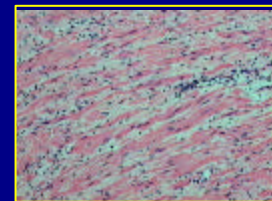
**PROTEOMIC ANALYSIS  
IN THE CONTEXT OF THE  
TISSUE MICROENVIRONMENT**



**Nerve**



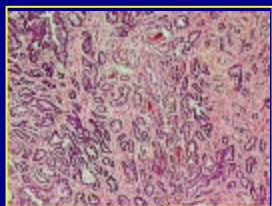
**Well-differentiated carcinoma**



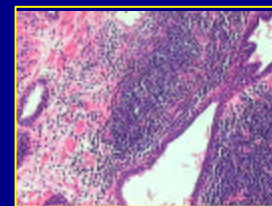
**Stroma**



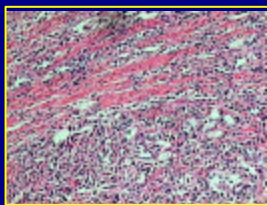
**Human Prostate**



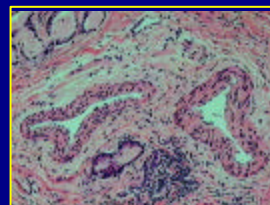
**Moderately-differentiated carcinoma**



**Inflammation**



**Poorly-differentiated carcinoma**



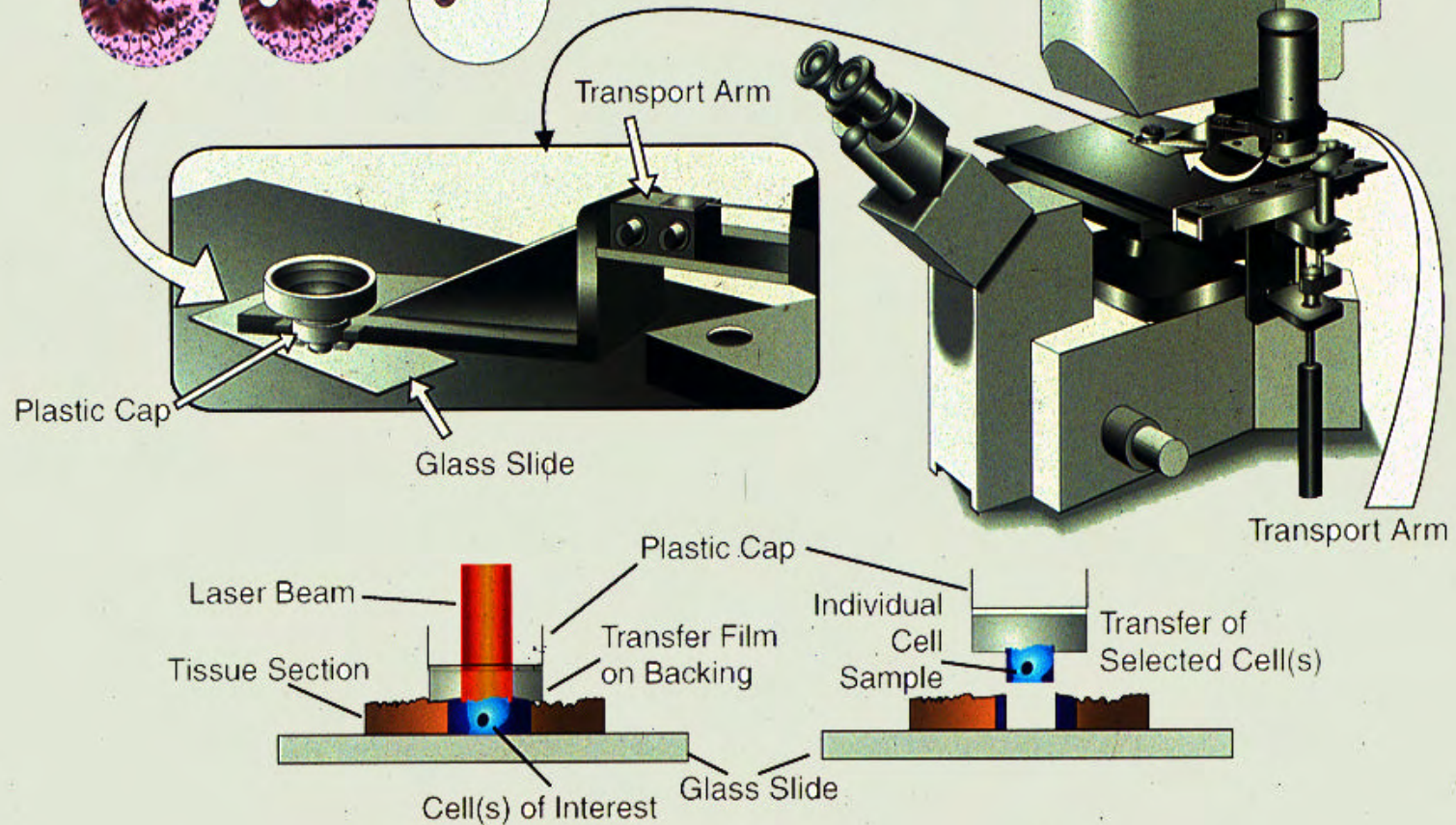
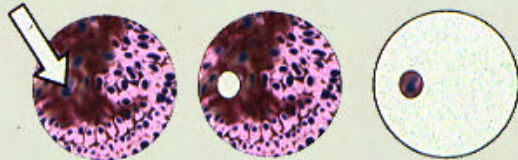
**Neovessels**



# NIH Laser Capture Microdissection

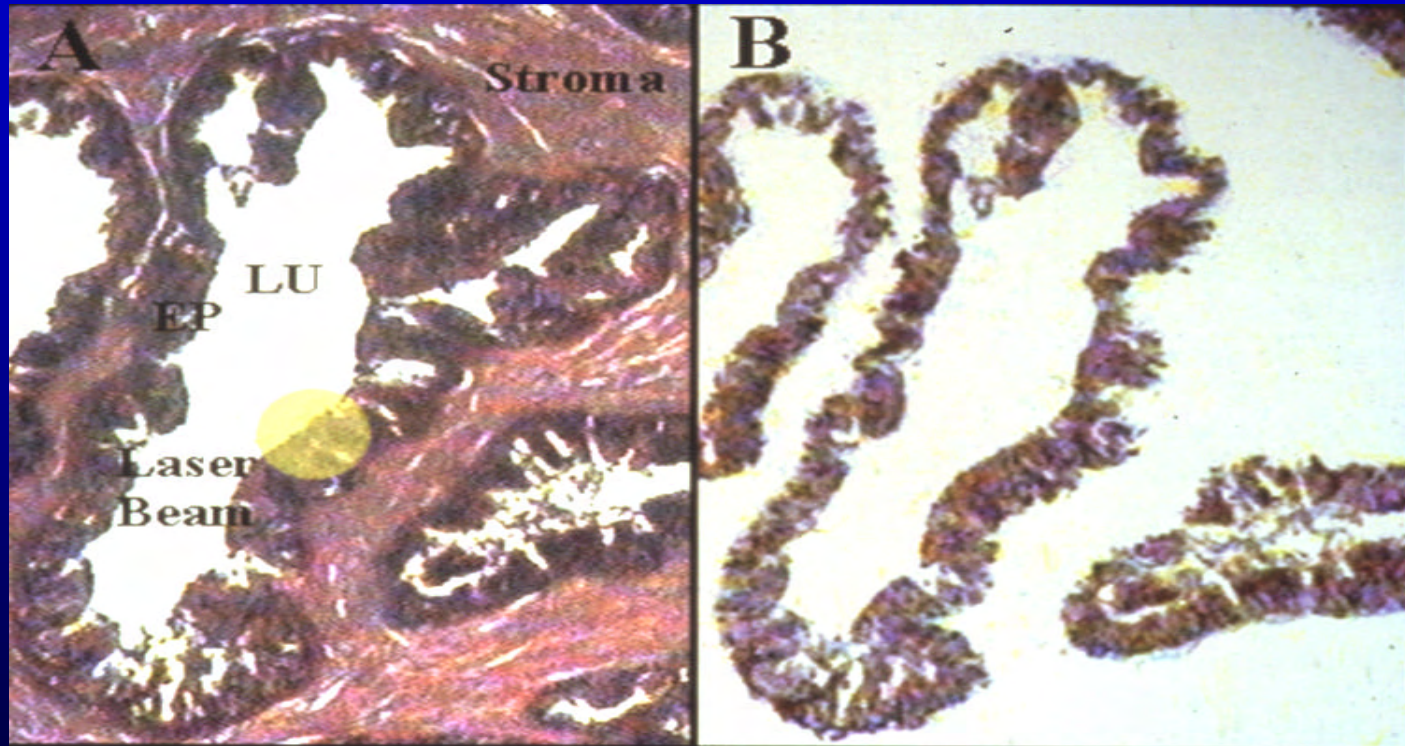
*Science*' 96, 97, 98

Cancerous Cell



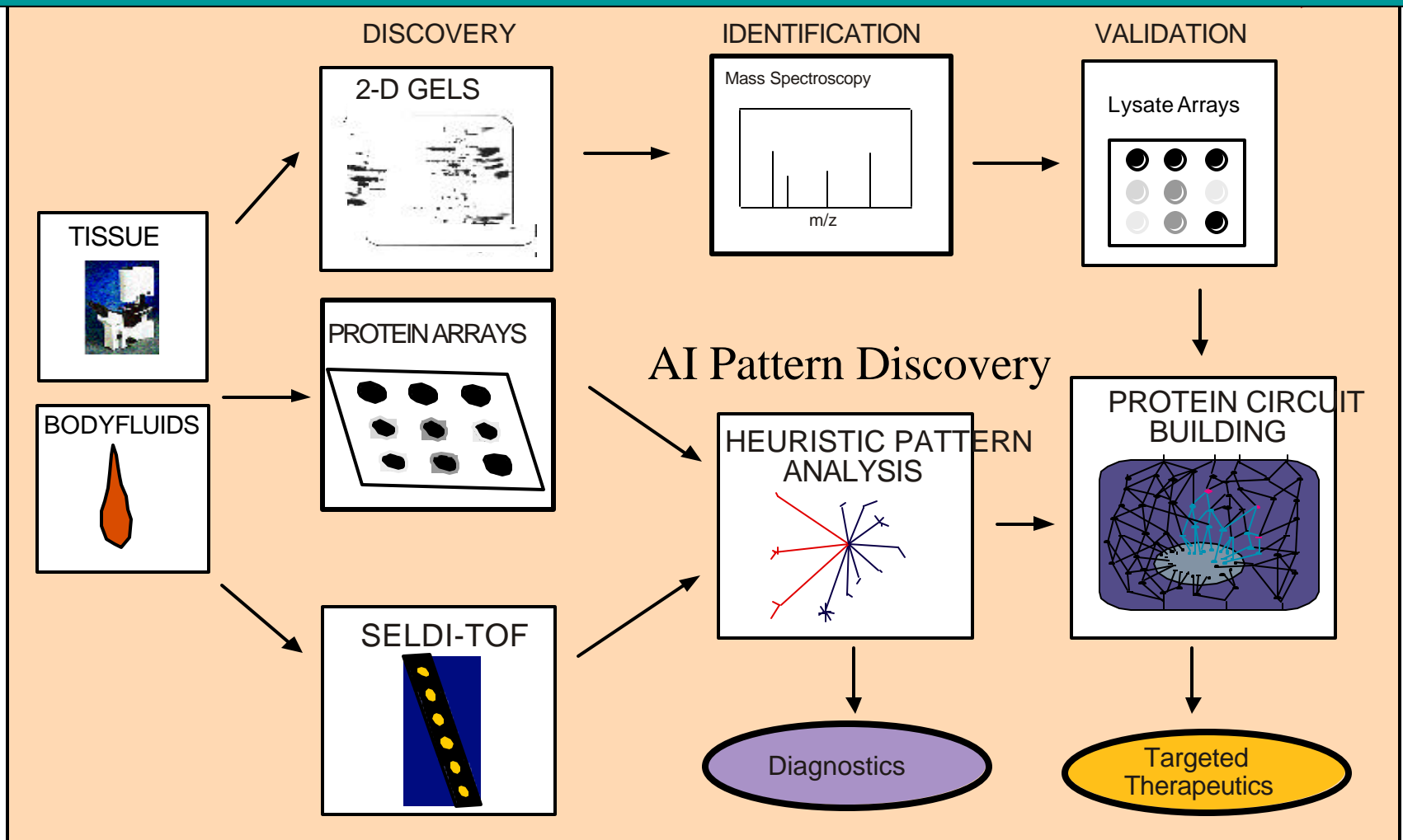
Before LCM

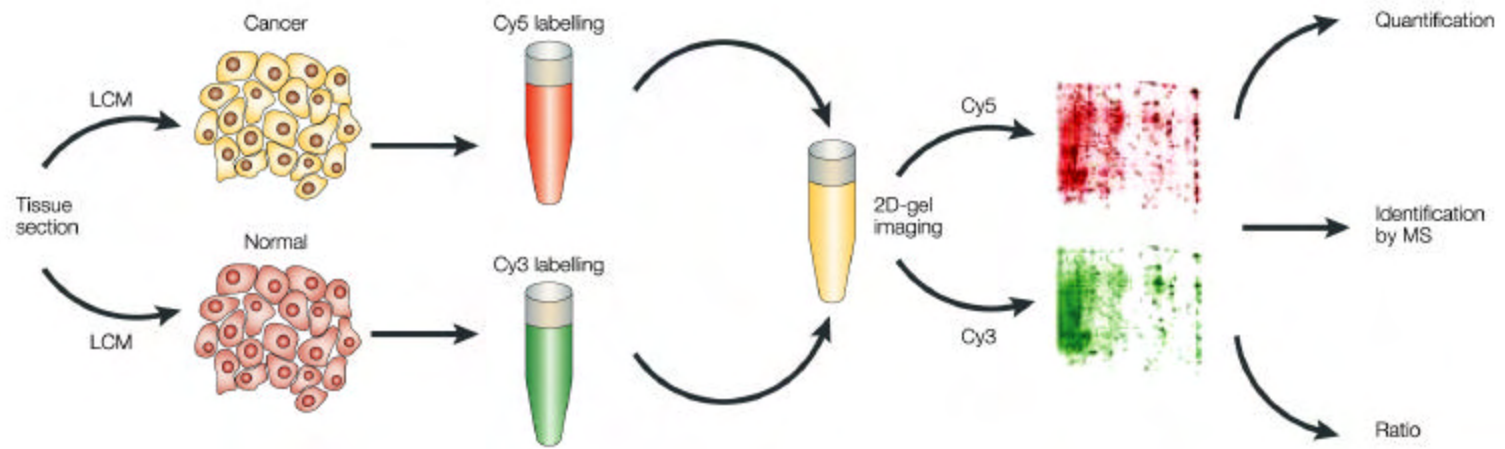
After LCM



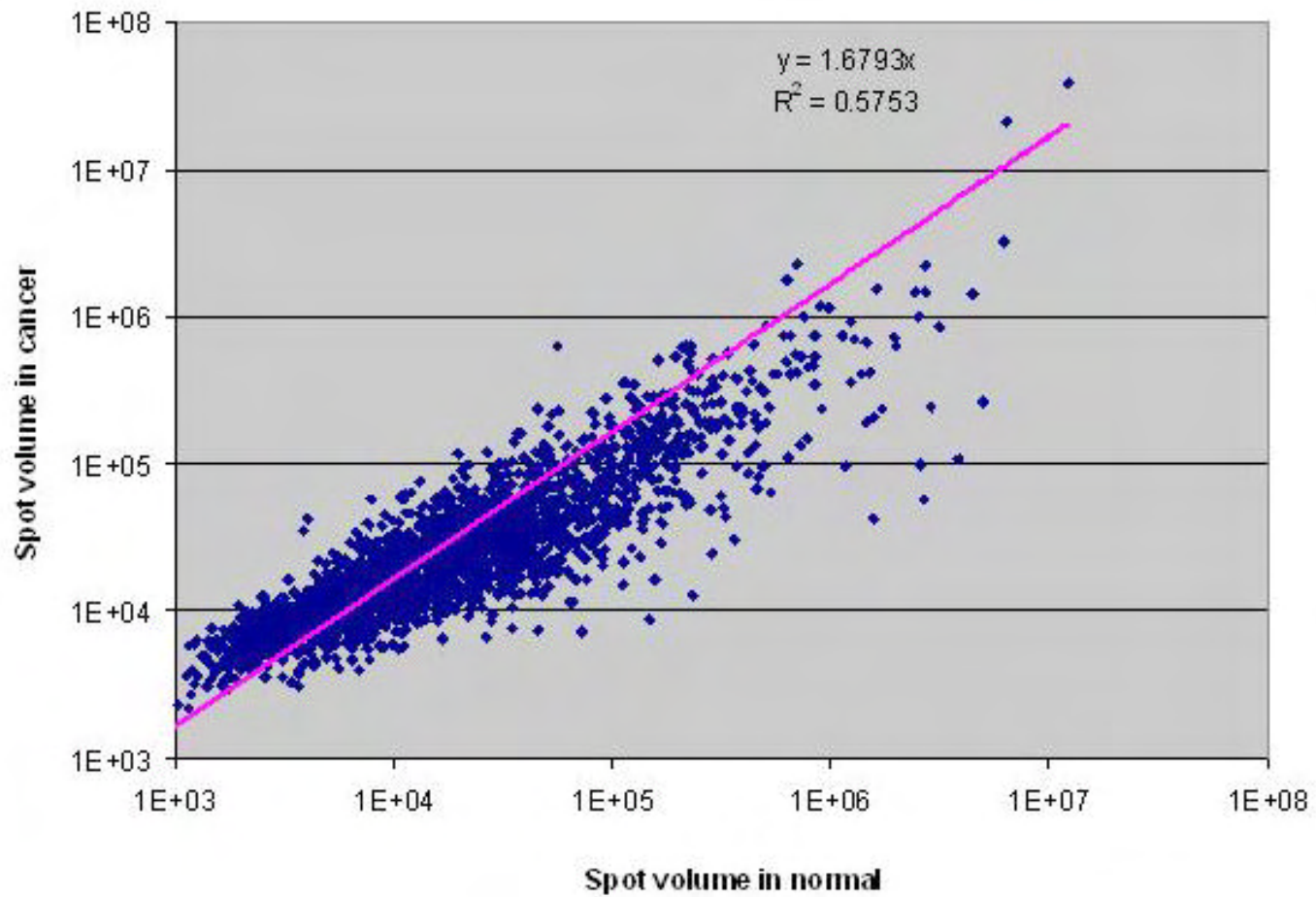
Case study: Prostate normal epithelium (human)

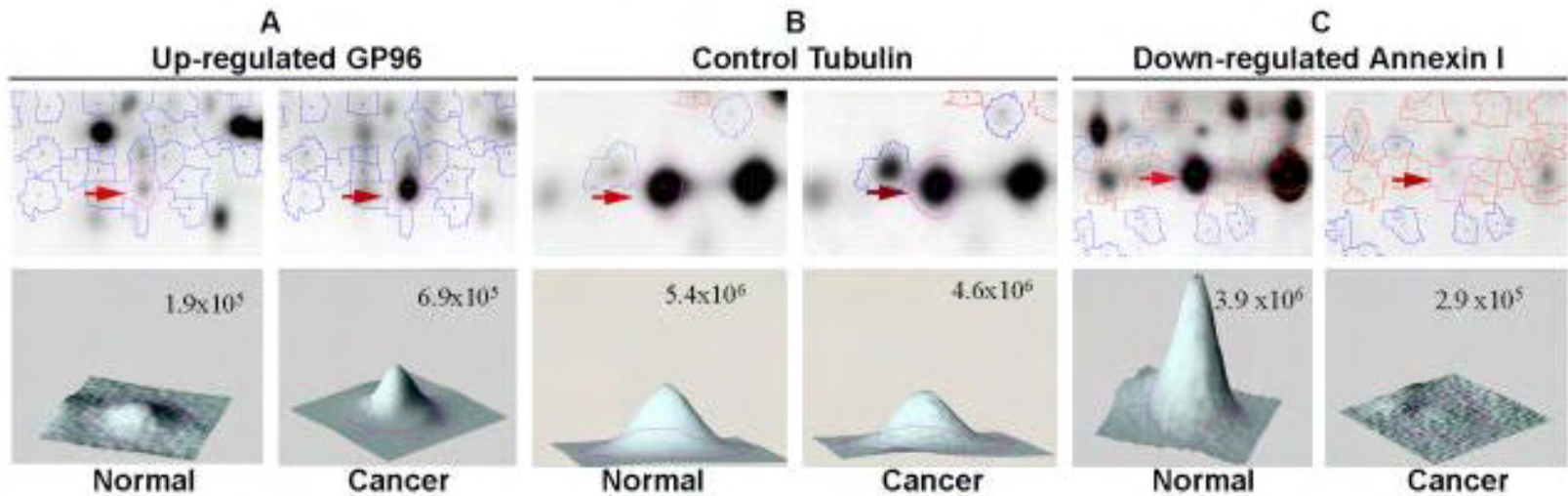
# NCI - CBER/FDA CLINICAL PROTEOMICS PROGRAM- Began 2000



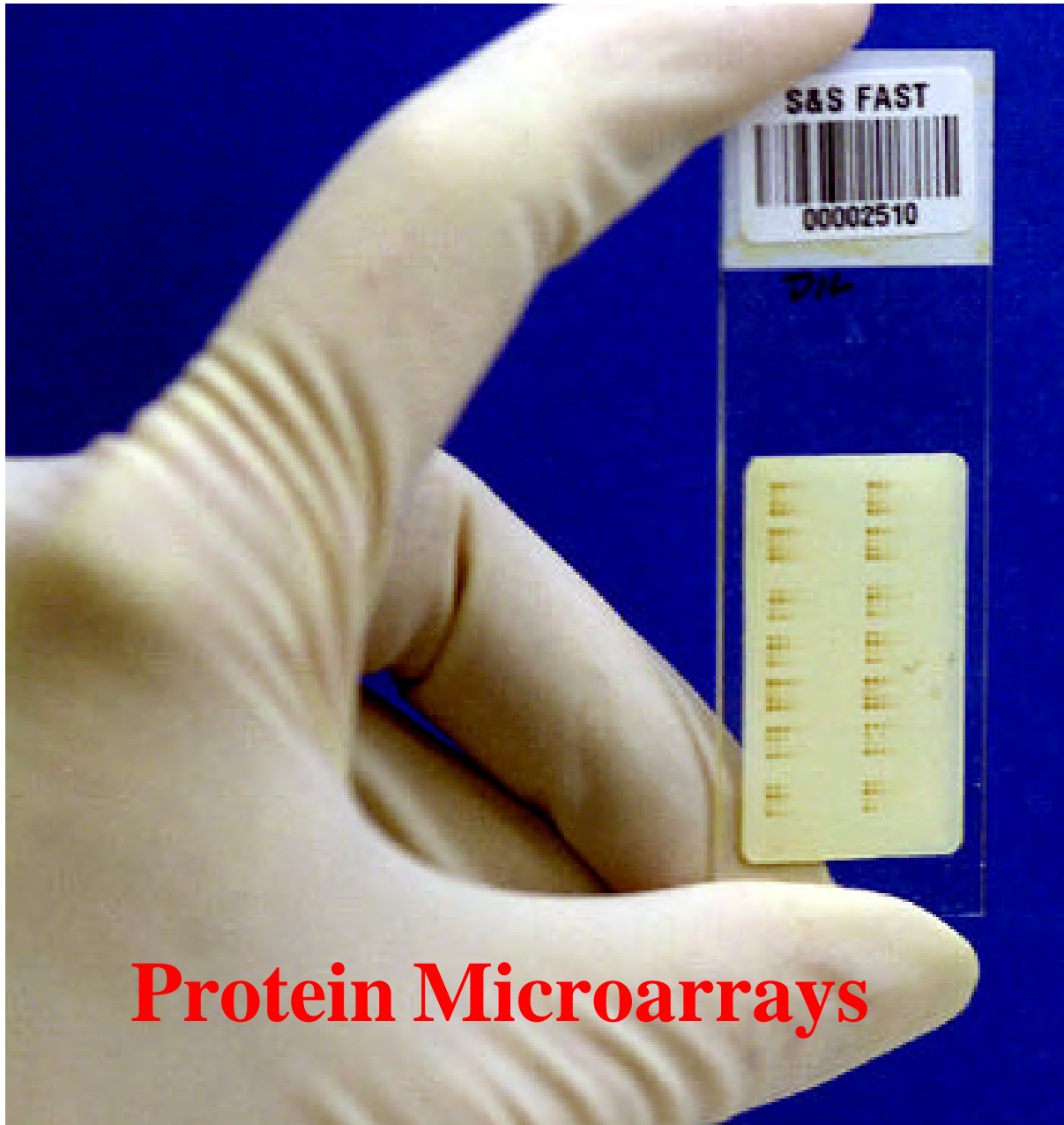




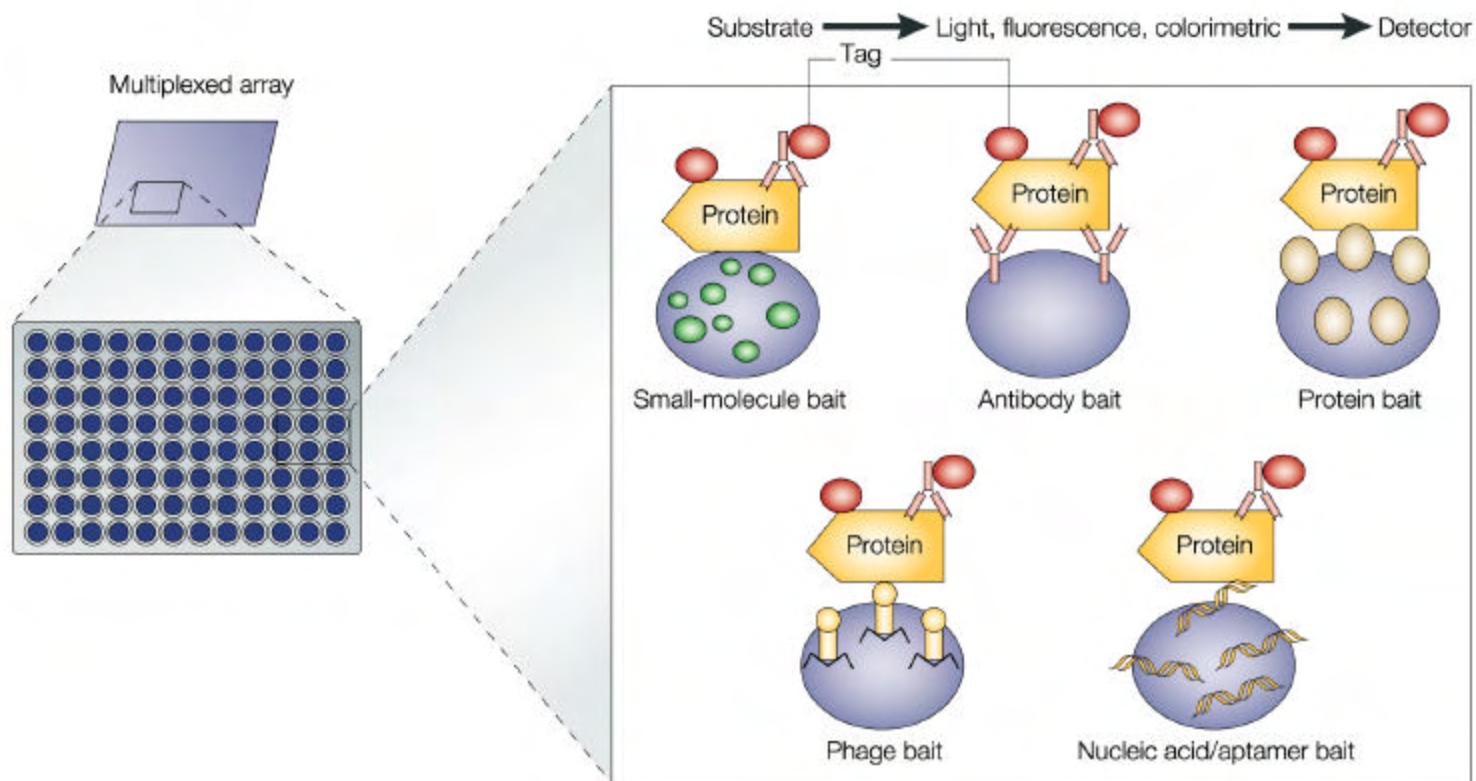




**Differentially expressed proteins identified to date: >400  
Breast, Prostate, Ovary, and Esophageal Cancer**



# Protein Microarrays



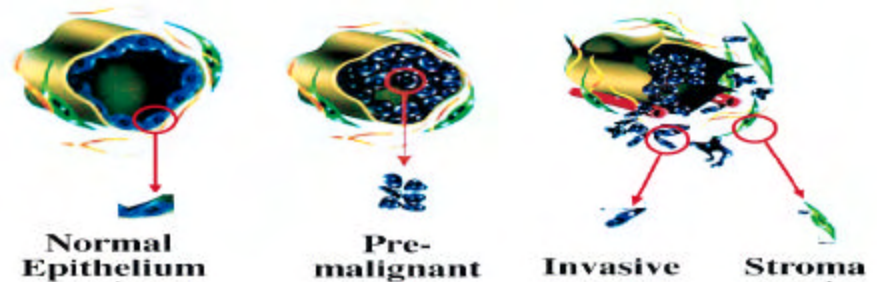


# A New Protein Array Technology: Reverse Phase Protein Array

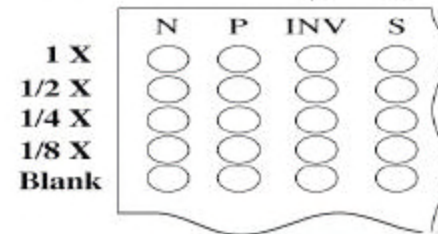
Oncogene 2001

Coupling Laser Capture Microdissection  
With High Throughput Protein Arrays

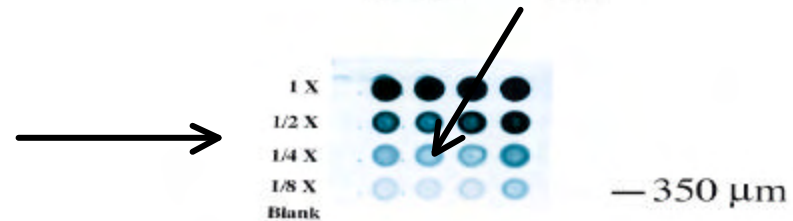
**Patient biopsy tissue cells are microdissected:**  
  
2000 cells = 100 arrays



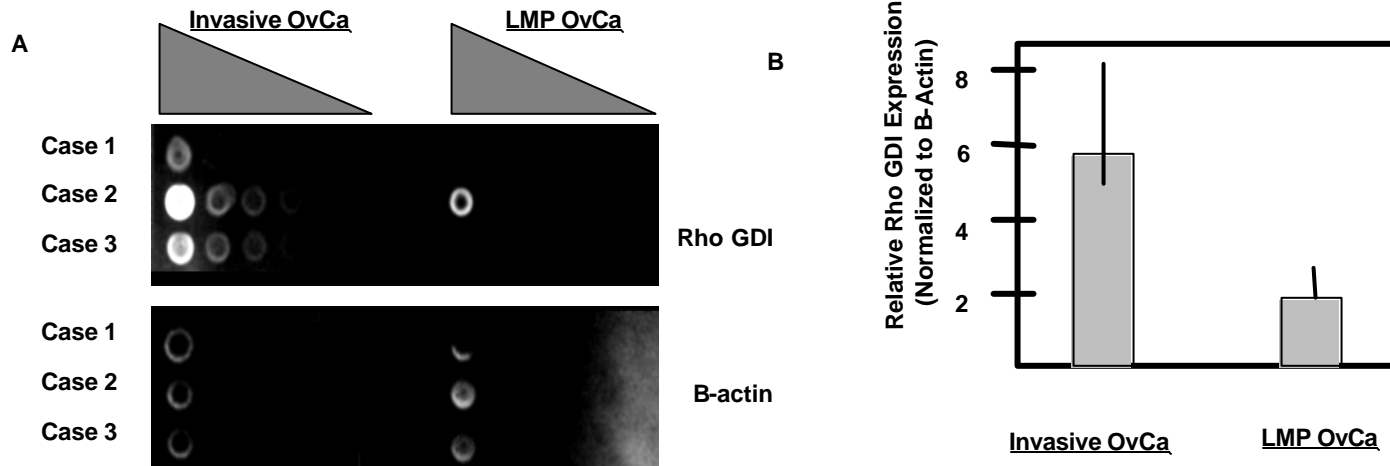
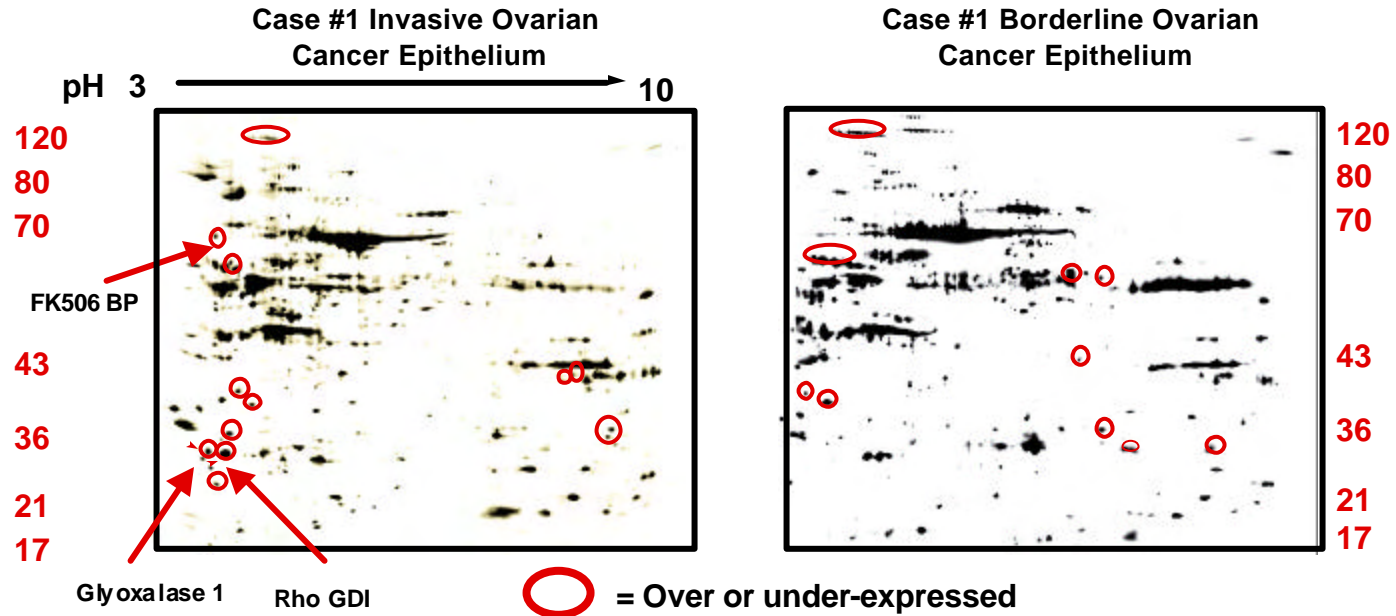
**Each patient sample is arrayed in a miniature dilution curve:**  
Always in linear dynamic range of any antibody/ analyte pair

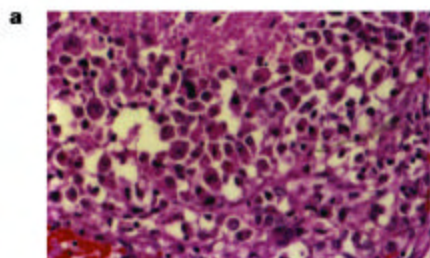


**Arrays probed with labeled amplified antibody:**  
  
e.g. prostate cancer progression  
From one patient probed with Phospho-ERK antibody

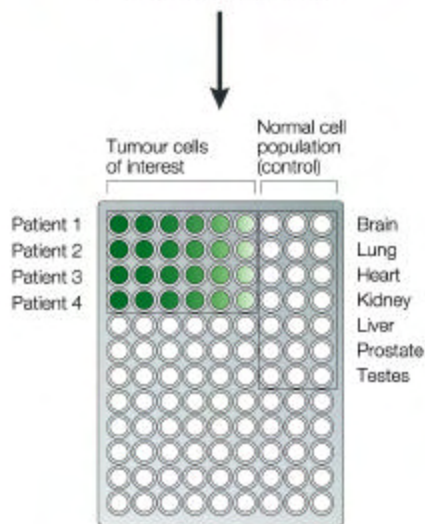


# Protein Microarrays for High-throughput Target Validation

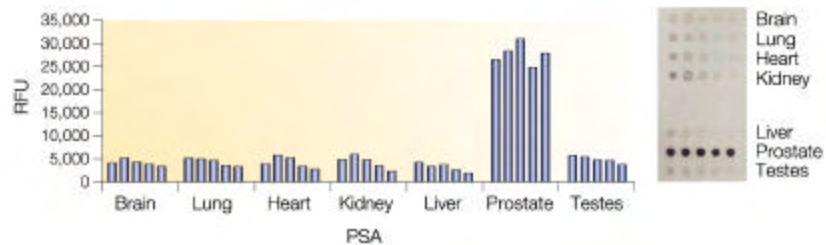
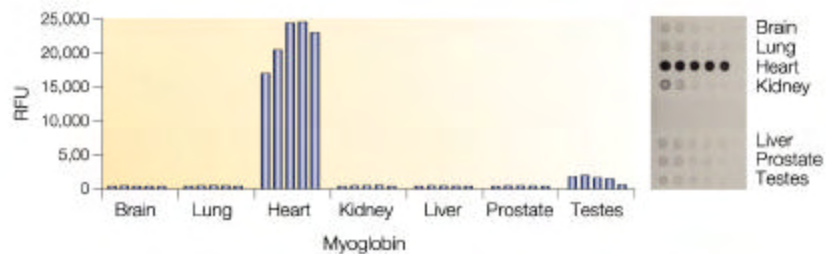
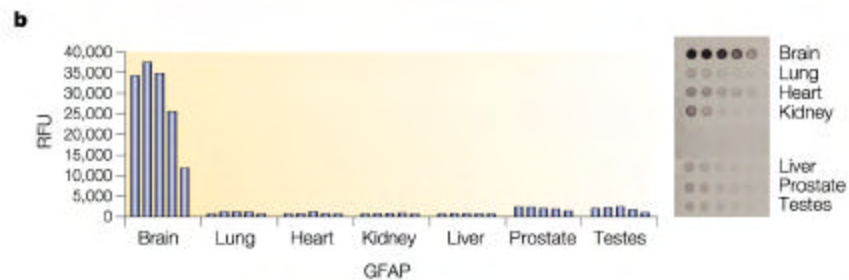




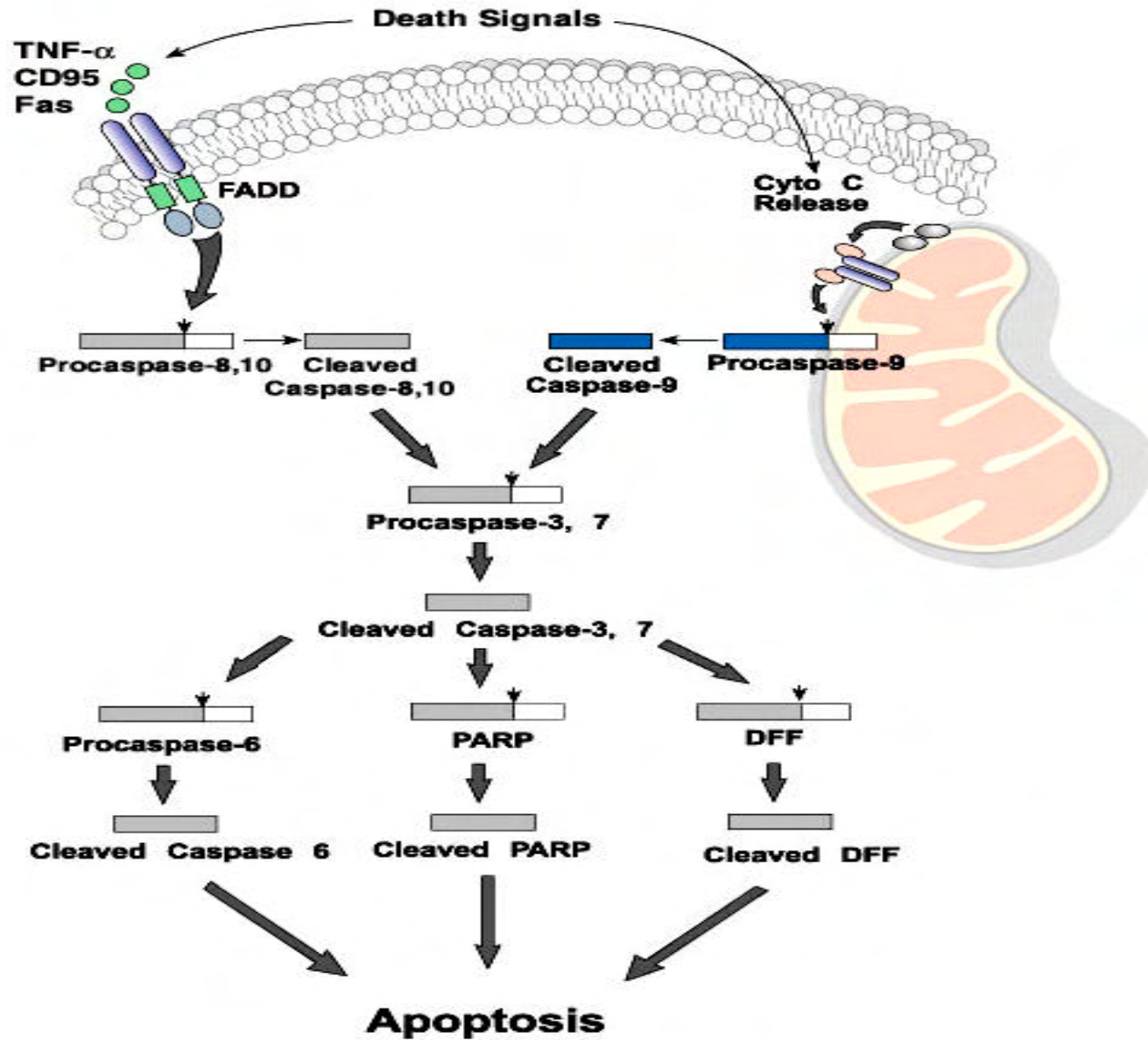
Microdissection of tissue cells



Analysis of vaccine candidates



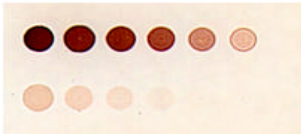
# Cell Survival Pathways



# Protein Microarrays

## Validation of Apoptosis Pathway

Cleaved-Caspase 3



Cytochrome-C treated Jurkat cells

Untreated Jurkat cells

Cleaved-Caspase 7



Cytochrome-C treated Jurkat cells

Untreated Jurkat cells

Caspase 7



Cleaved-Caspase 9



Cytochrome-C treated Jurkat cells

Untreated Jurkat cells

Caspase 9



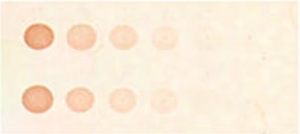
Cleaved-PARP



Cytochrome-C treated Jurkat cells

Untreated Jurkat cells

PARP



Negative Control



Cytochrome-C treated Jurkat cells

Untreated Jurkat cells

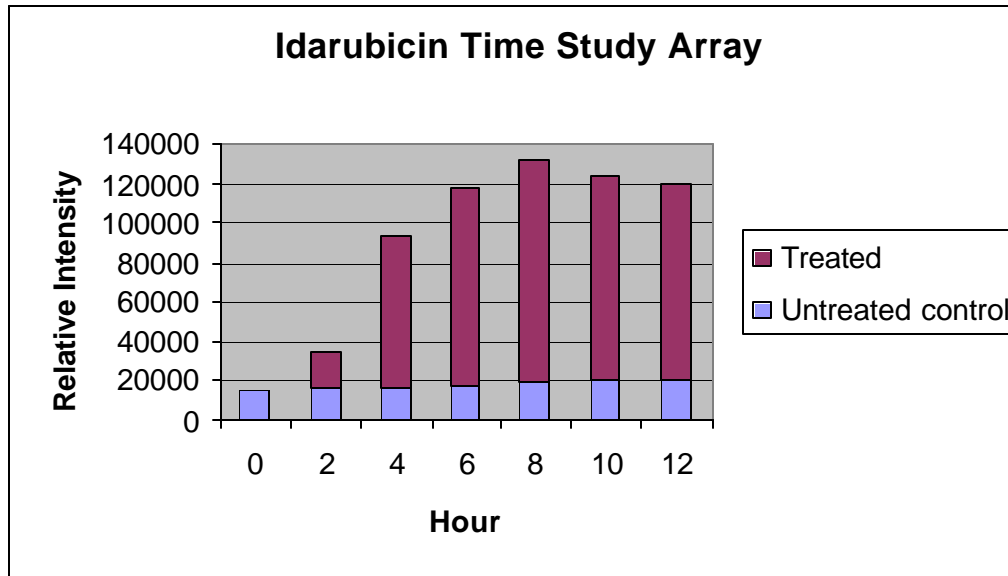
**Human B Lymphoma  
Apoptosis Pathway  
Protein Microarrays**

**Cleaved Caspase 3**

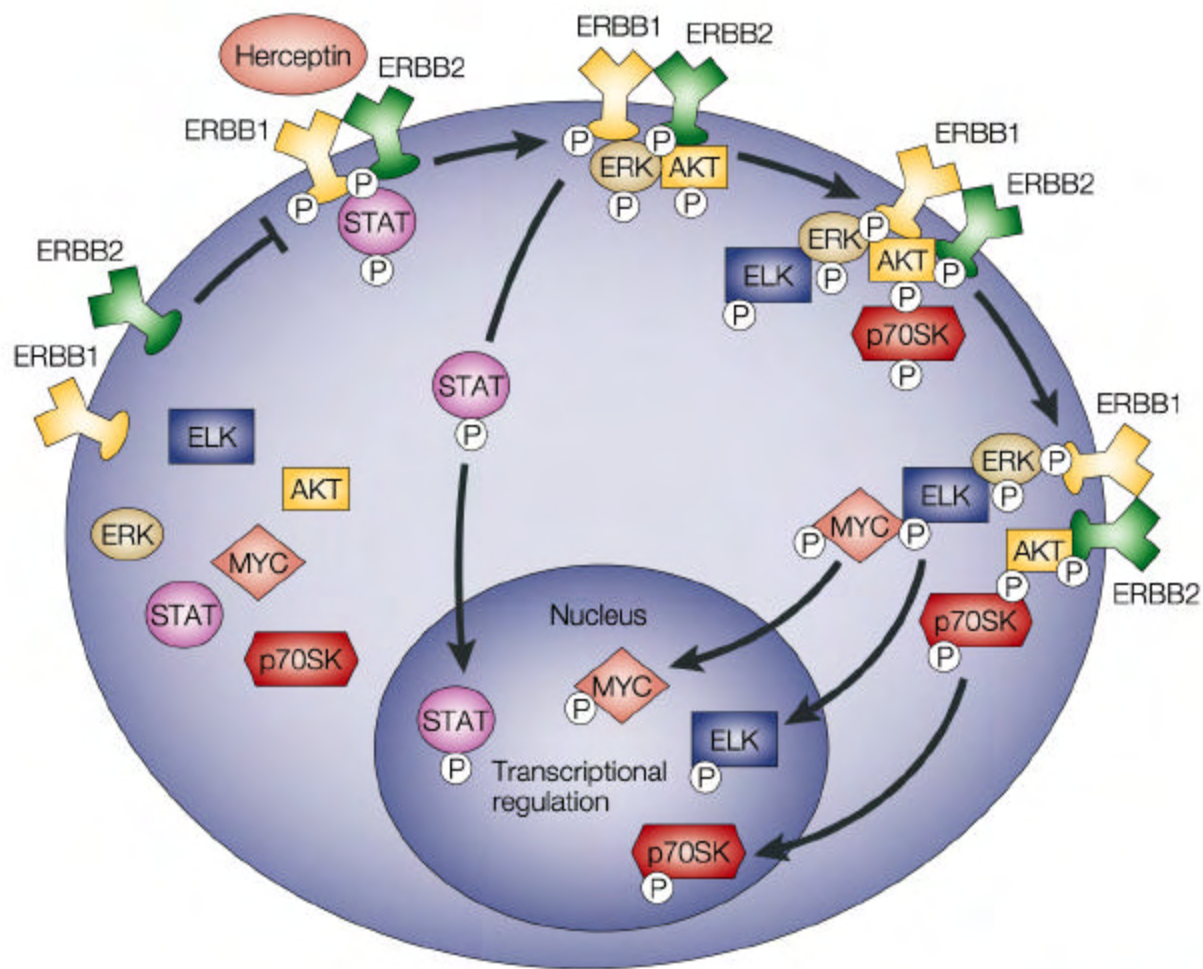


Untreated Treated

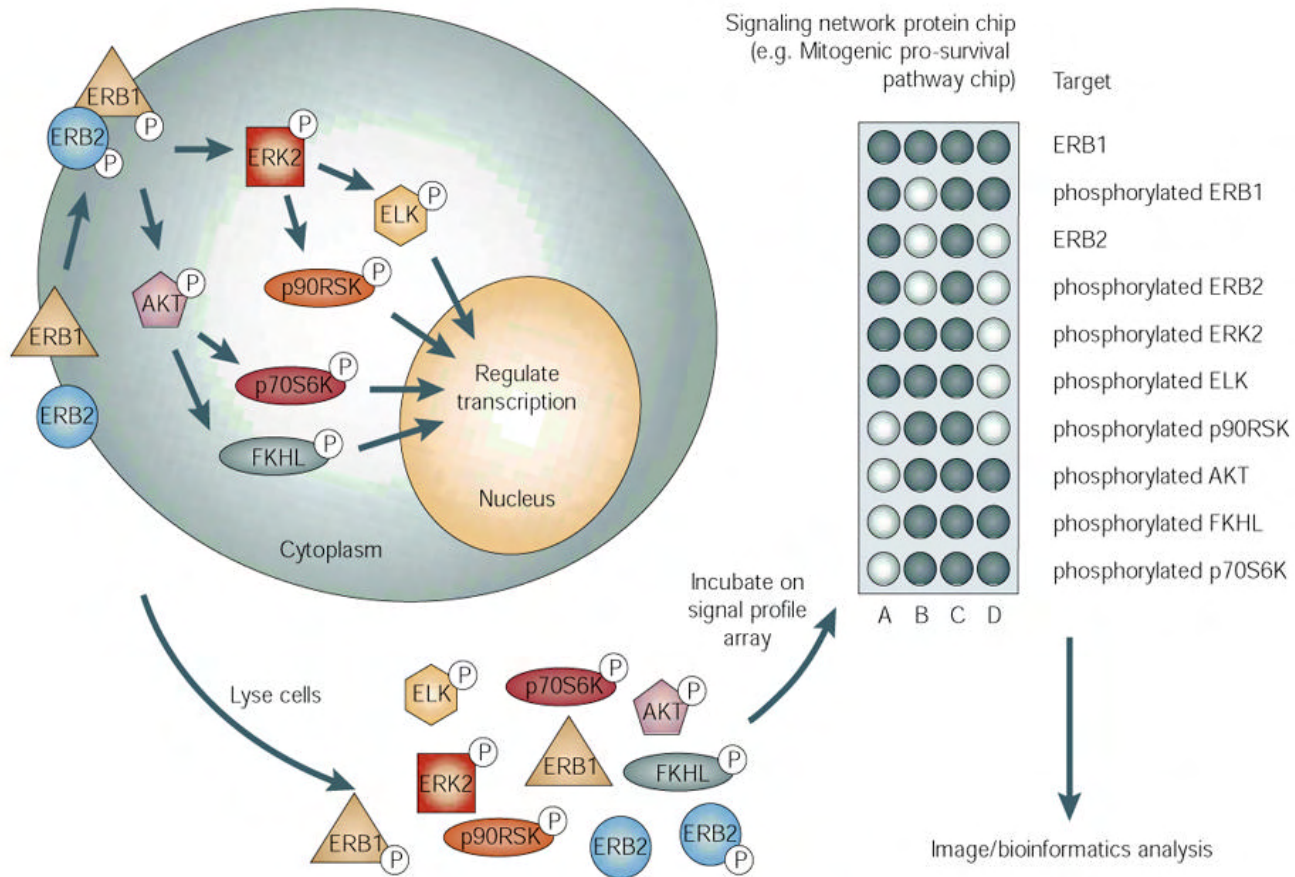
Time  
Course







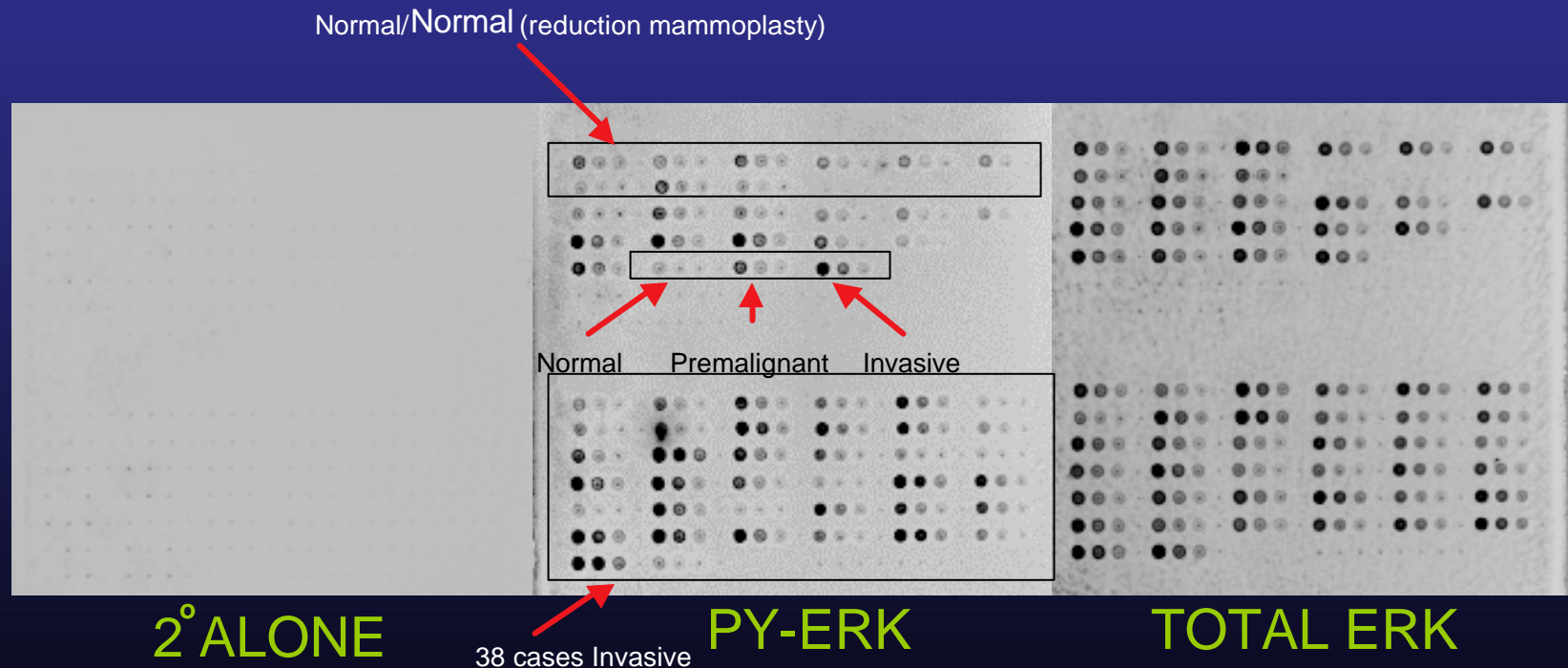
# Signal Transduction Pathway Profiling





# Use of Novel Protein Array Technology: Signal Pathway Profiling in Human Breast Cancer Biopsy Specimens

Coupling Laser Capture Microdissection With True Signal Pathway Profiling

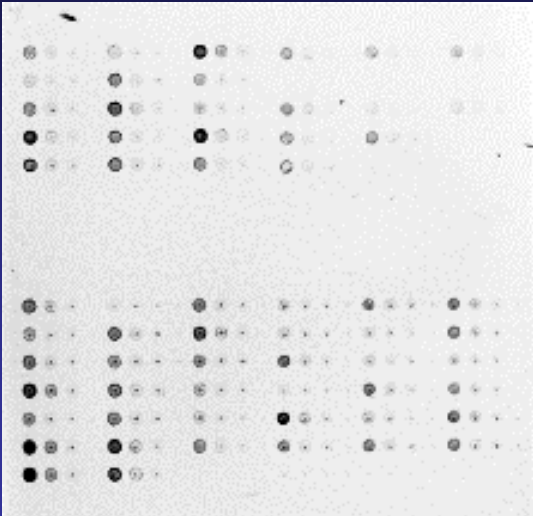
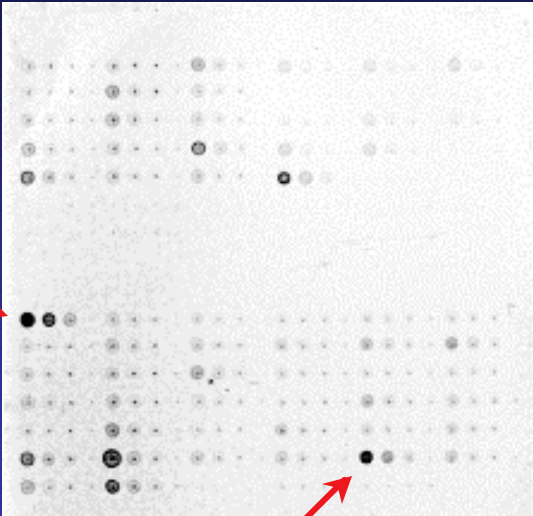


Ongoing work: cluster analysis with 135 phospho-specific endpoints, all normalized to the self protein for true signal pathway profiling

**Phospho-AKT (ser 473)**

**TOTAL AKT**

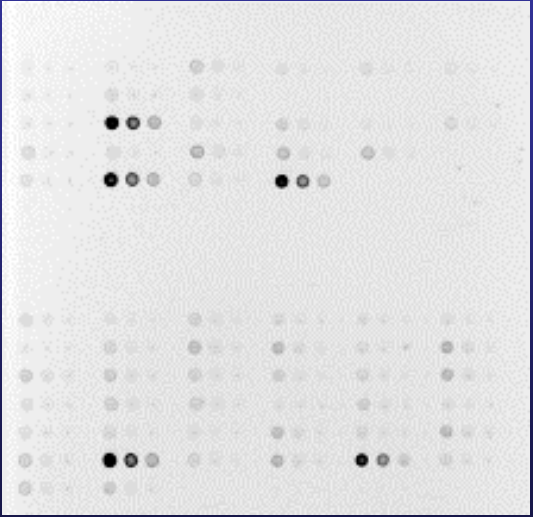
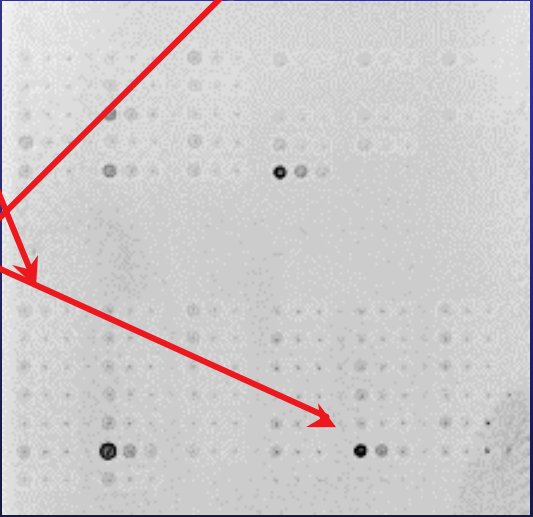
Discordant activation



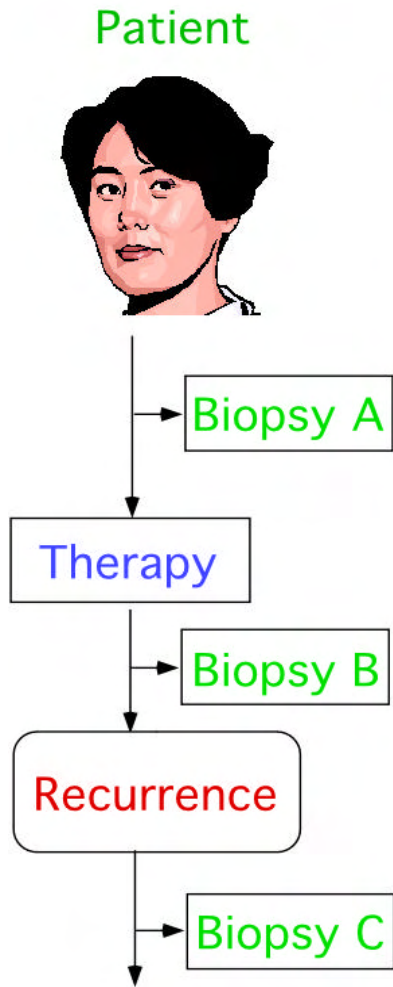
**Phospho-ERB2**

**Total ERB2**

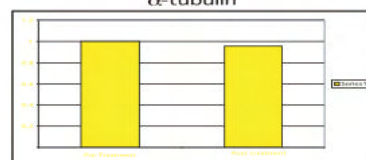
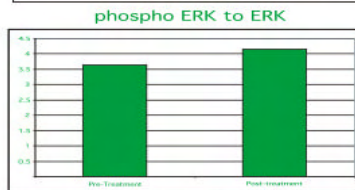
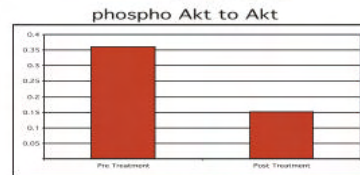
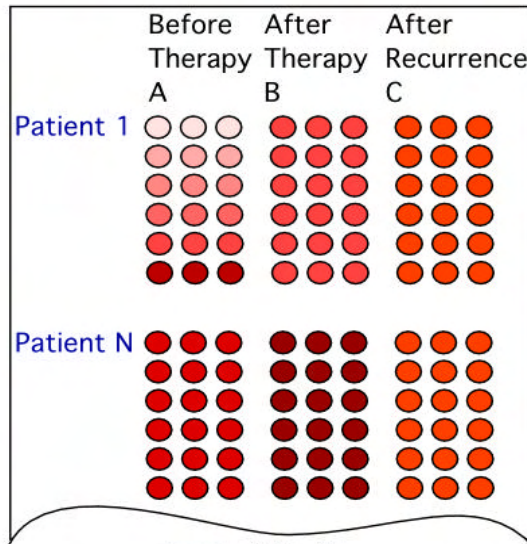
Concordant activation



# Clinical Trial Molecular Target Analysis



## Reverse Phase Protein Arrays



## Clinical Trial:

- Herceptin followed by Taxol
- Metastatic Breast and Ovarian Ca
- Findings to date

## HERCEPTIN REDUCES P-Akt PROSURVIVAL PATHWAY

## HYPOTHESIS:

- Increased Sensitivity to apoptosis inducing therapy (e.g. Taxol)
- Suppression of growth through de-repression of p21(Cip1/WAF)

# Proteomic Endpoints from Clinical Trial Biopsies: Use of Protein Arrays

Pre and Post HERCEPTIN (1 Month)

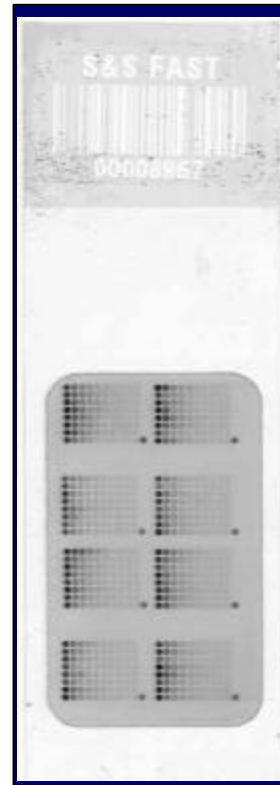
- Phospho-AKT Endpoint
- 500 microdissected cells
- Pre and Post Treatment Studies

**Responders: 1,3,4**  
**Non-Responder: 2**

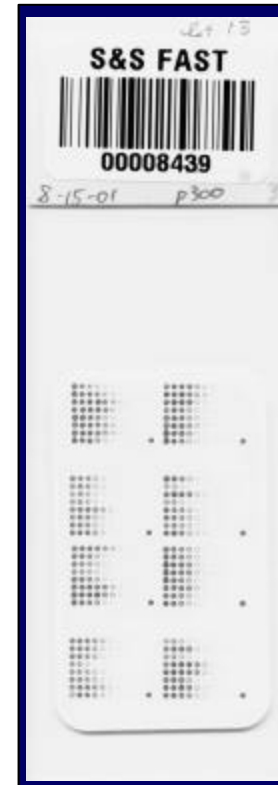


# Protein array specification

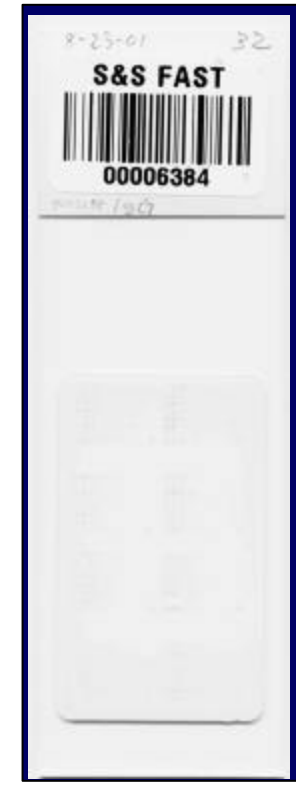
- Schleicher & Schuell ([www.s-and-s.com](http://www.s-and-s.com)) FAST slide (glass slide embedded 21 x 35 mm nitrocellulose membrane) was used.
- Total number of spots is 648.
- Spot all NCI60 cell lysates and 4 pools on a single slide.
- Each cell line has 10 different concentration spots.
- Achieved more than 1000-fold dynamic range.
- Requires total protein and negative control stains for a protein expression measurement.
- Takes 5 hours for making 20 full arrays.



Total protein (stained by SYPRO RUBY)



Protein of interest (p300)



Negative control (mouse IgG)

# Raw pixel data generation by P-SCAN

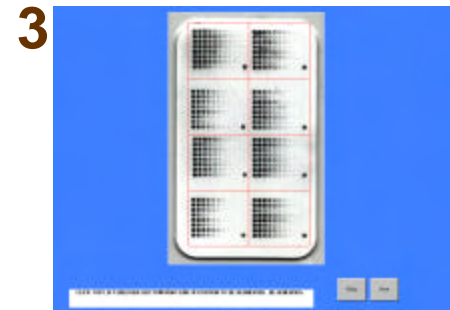
P-SCAN (Peak quantification using Statistical Comparative ANalysis) is available at <http://abs.cit.nih.gov/pscan/>



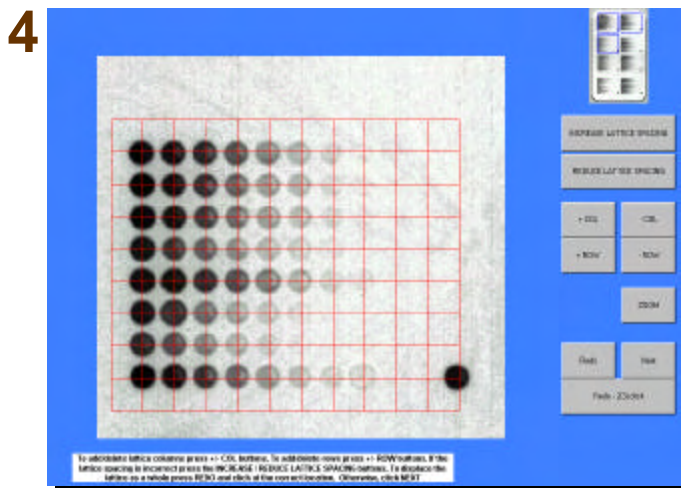
Apply an image (TIFF) and select the area of array.



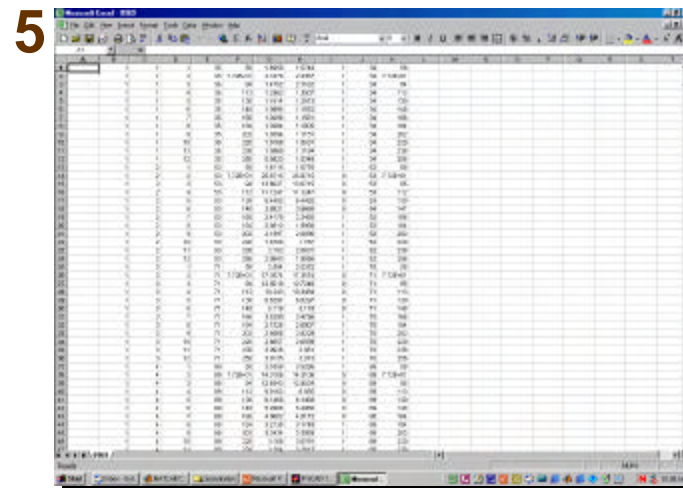
The array has been selected.



Set up the field. Intensity reading will be carried out by each field. There are  $2 \times 4 = 8$  fields above.



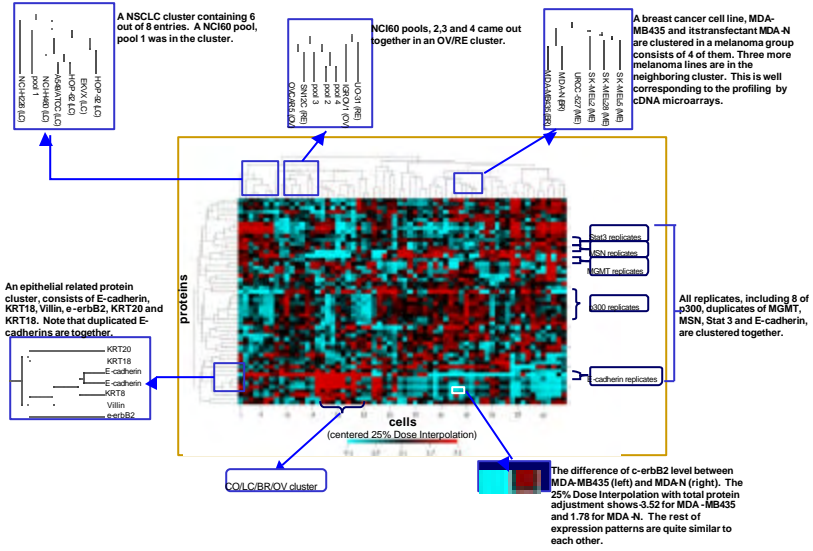
Align the lattice. A total of 120 intersections will set in a field and generates intensity number per spot. The right bottom dark spot is for control the alignment.



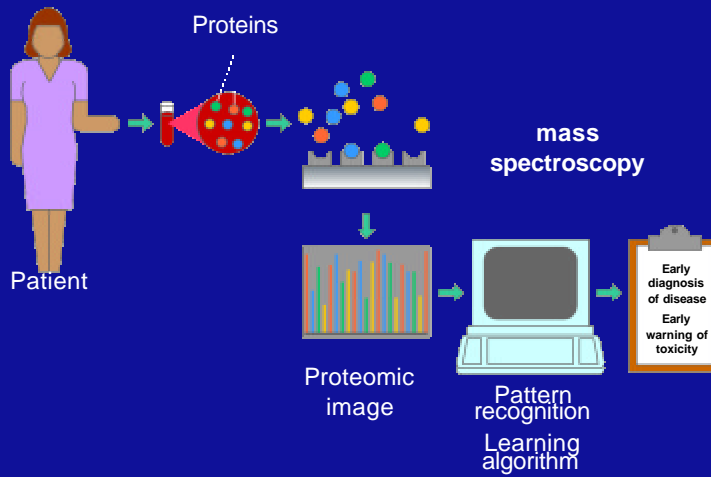
Raw pixel intensity data is exported onto an Excel worksheet along with its address on the array.



# Cluster analysis of protein expression (1)

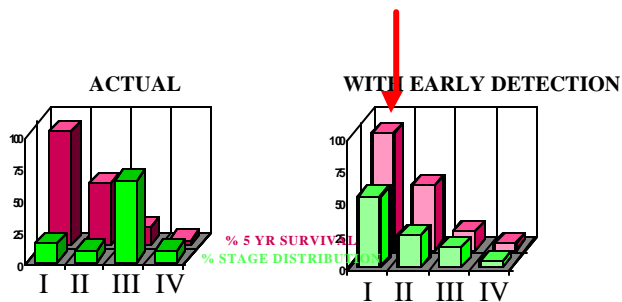


# Serum Protein Pattern Diagnostics



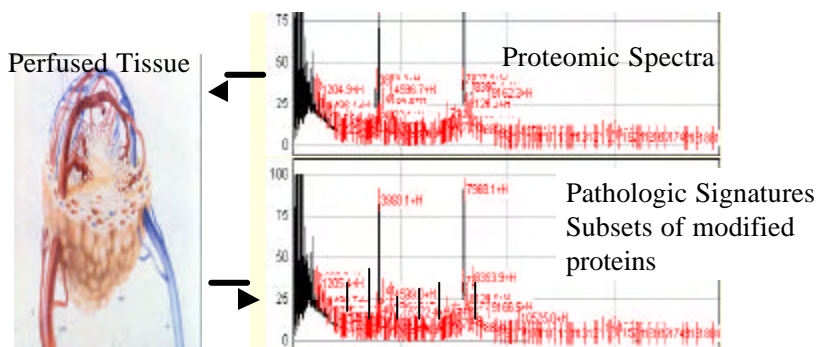
## PROTEOME SNAPSHOT PATTERN ANALY APPLICATION TO EARLY DETECTION OF OVA

CAN PROTEIN PROFILING IDENTIFY A PROTEIN EXPRESSION  
DIAGNOSTIC OF INVASIVE EPITHELIAL OVARIAN CA



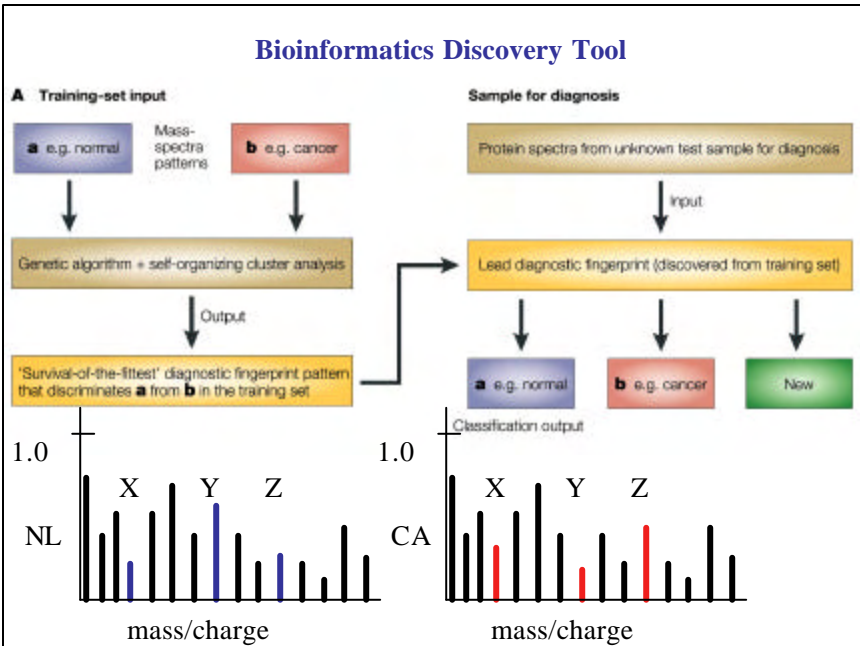
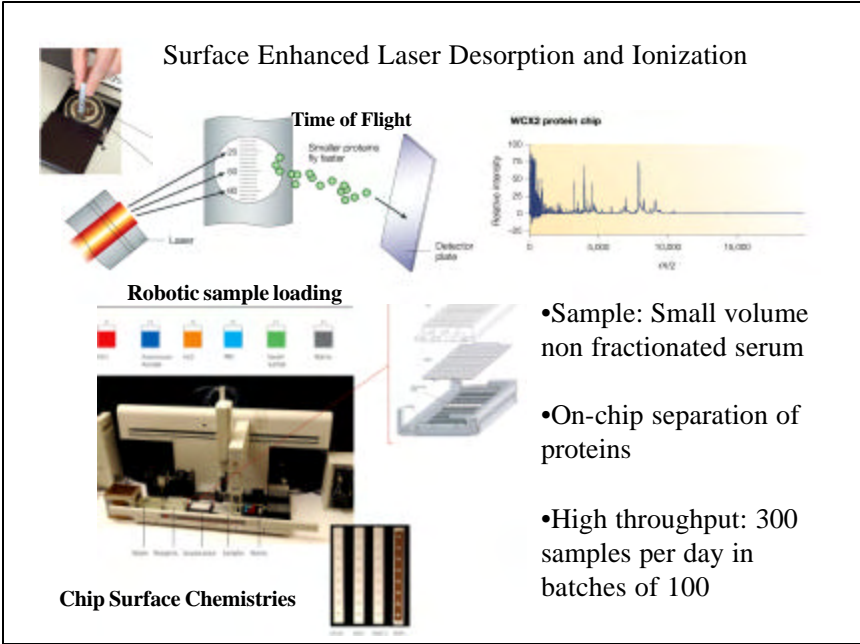
A SHIFT IN NUMBER OF EARLY STAGE PATIENTS WILL EFF  
AND OVERALL SURVIVAL IN ABSENCE OF TREATMENT CO

## PATTERNS OF PROTEOMIC INFORMATION IN SERUM



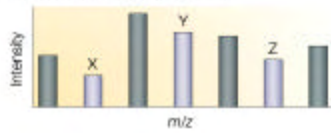
**•How can we discover diagnostic proteomic patterns even  
without knowing the identity of the proteins ahead of time?**



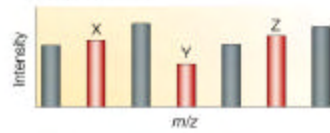


## Proteomic Pattern Clustering in n-Space

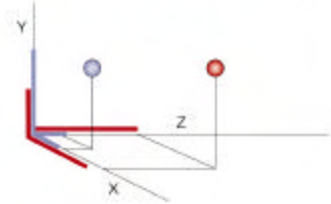
**B** Benign/unaffected spectra



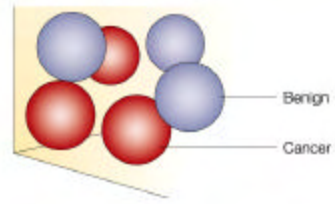
Cancer spectra



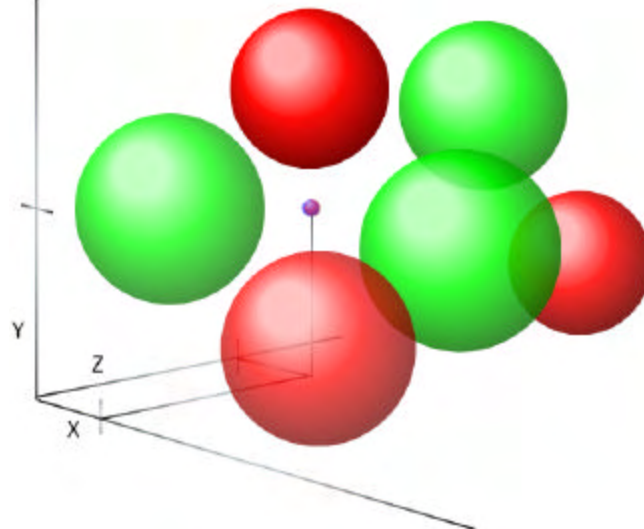
Plot of each pattern as a point in n-space



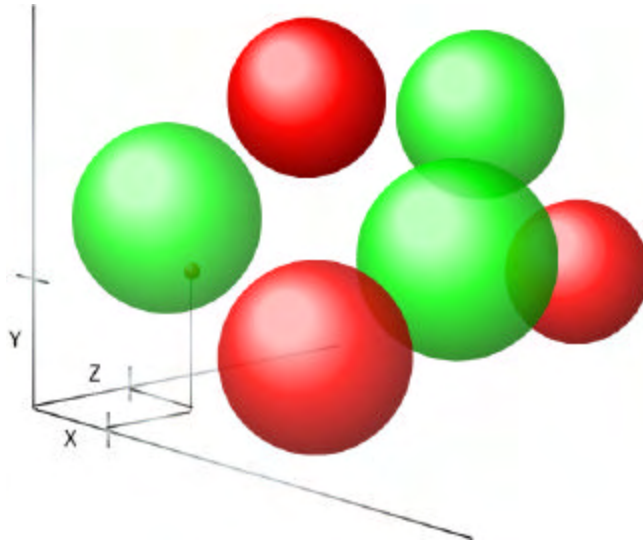
Clusters of points in n-space



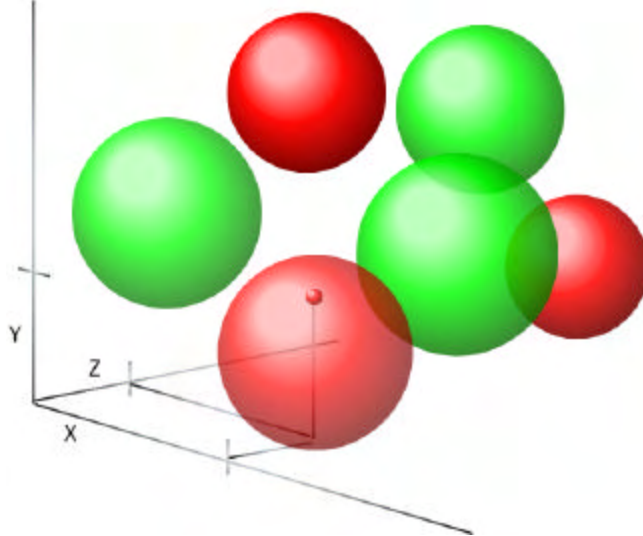
## Classification of Unknown: NO MATCH



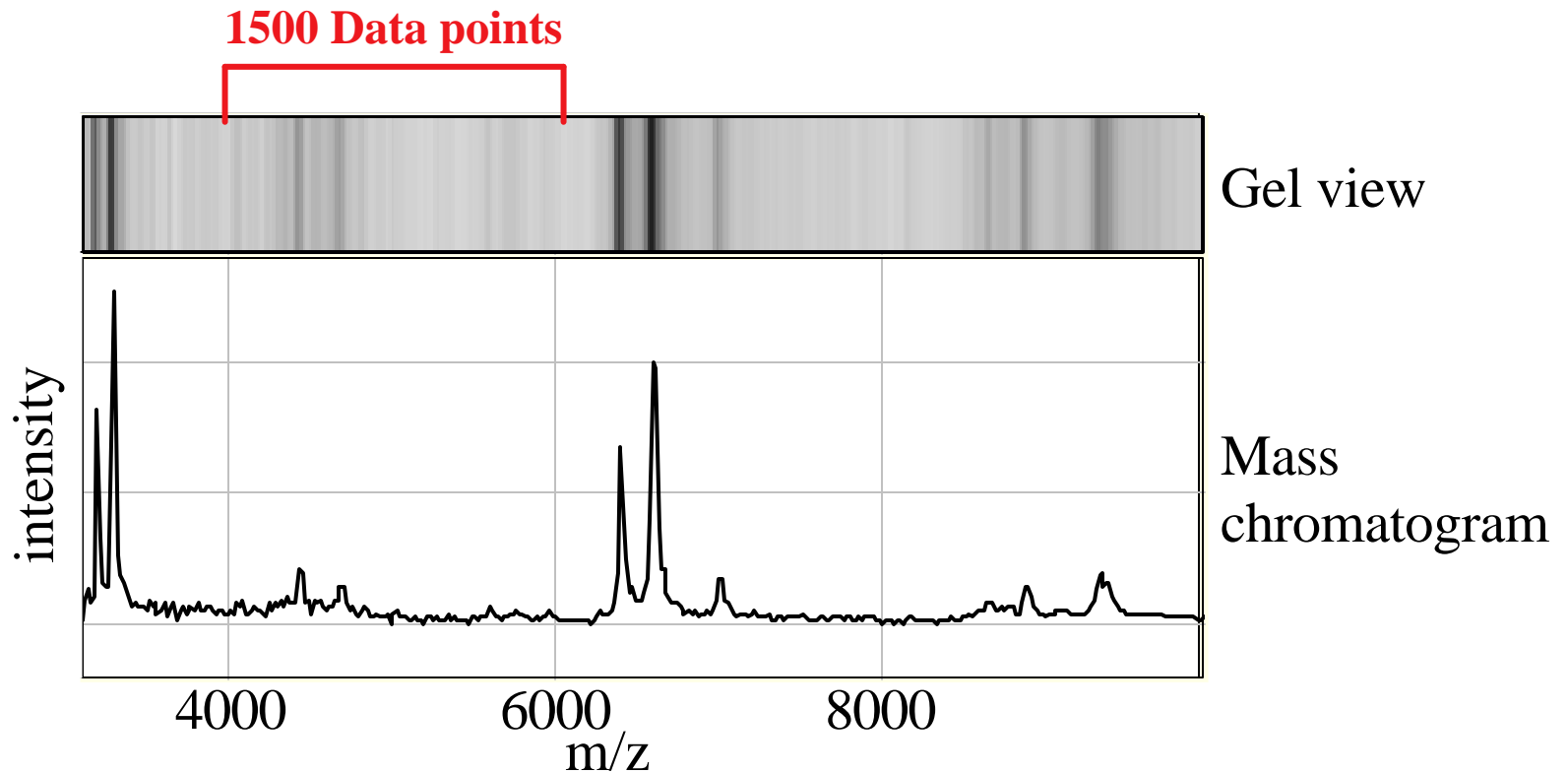
Classification of Unknown: BENIGN MATCH



Classification of Unknown: CANCER MATCH



# Typical serum profile from SELDI analysis



Data analysis window: 0-20,000 Daltons = 15,500 data points

<b>OVARY UNAFFECTED TRAINING SET <sup>a</sup></b>	
<b>DISEASE STATUS</b>	<b>N</b>
<b>No Evidence of Ovarian Cysts</b>	37
<b>Benign Ovarian Cysts &lt; 2.5 cm</b>	11
<b>Benign Ovarian Cysts &gt; 2.5 cm</b>	2
<b>TOTAL</b>	<b>50</b>

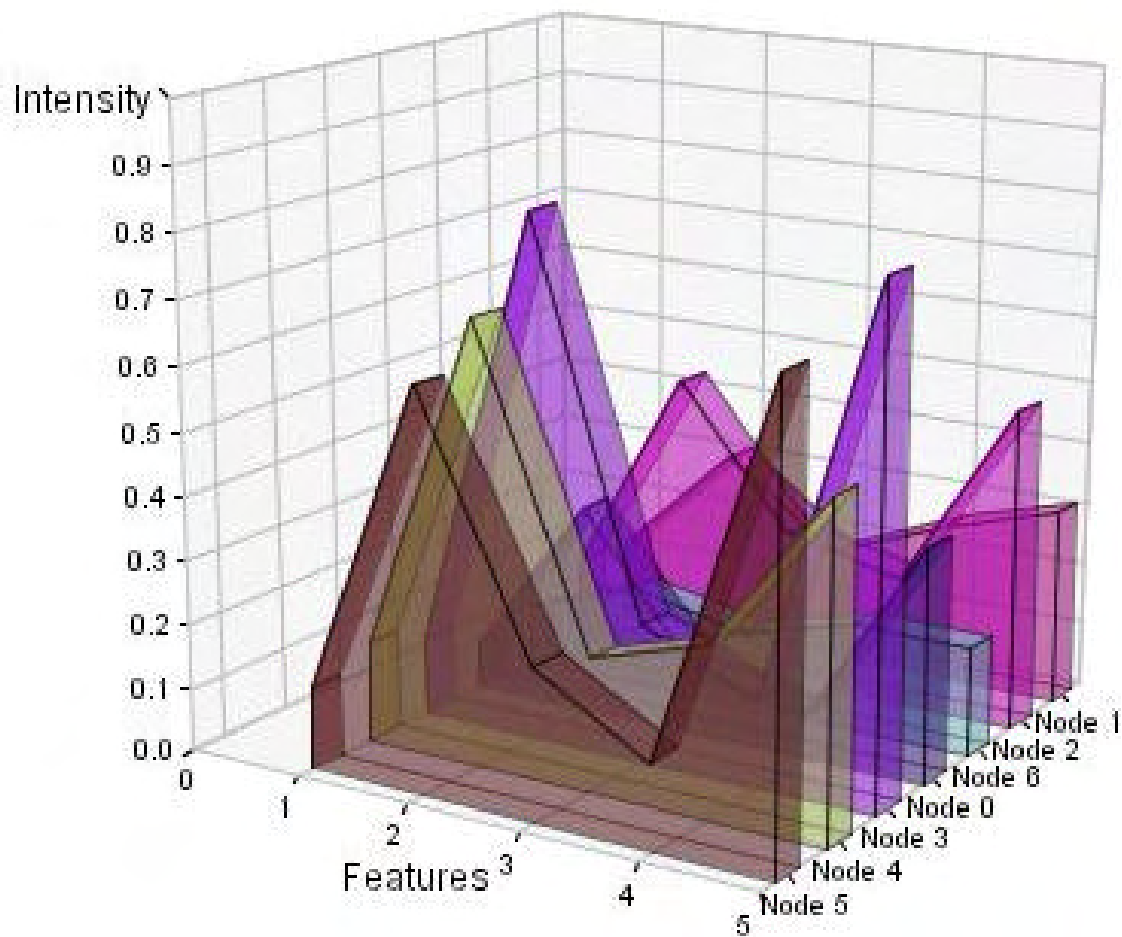
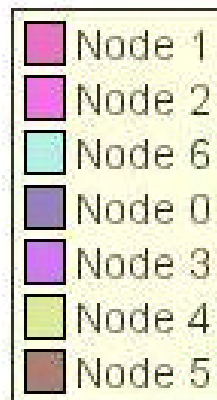
<b>OVARY CANCER TRAINING SET <sup>b</sup></b>	
<b>DISEASE STATUS</b>	<b>N</b>
<b>Surgically Staged Ovarian Cancer Stage II, III, IV</b>	44
<b>Surgically Staged Ovarian Cancer Stage I</b>	6
<b>TOTAL</b>	<b>50</b>

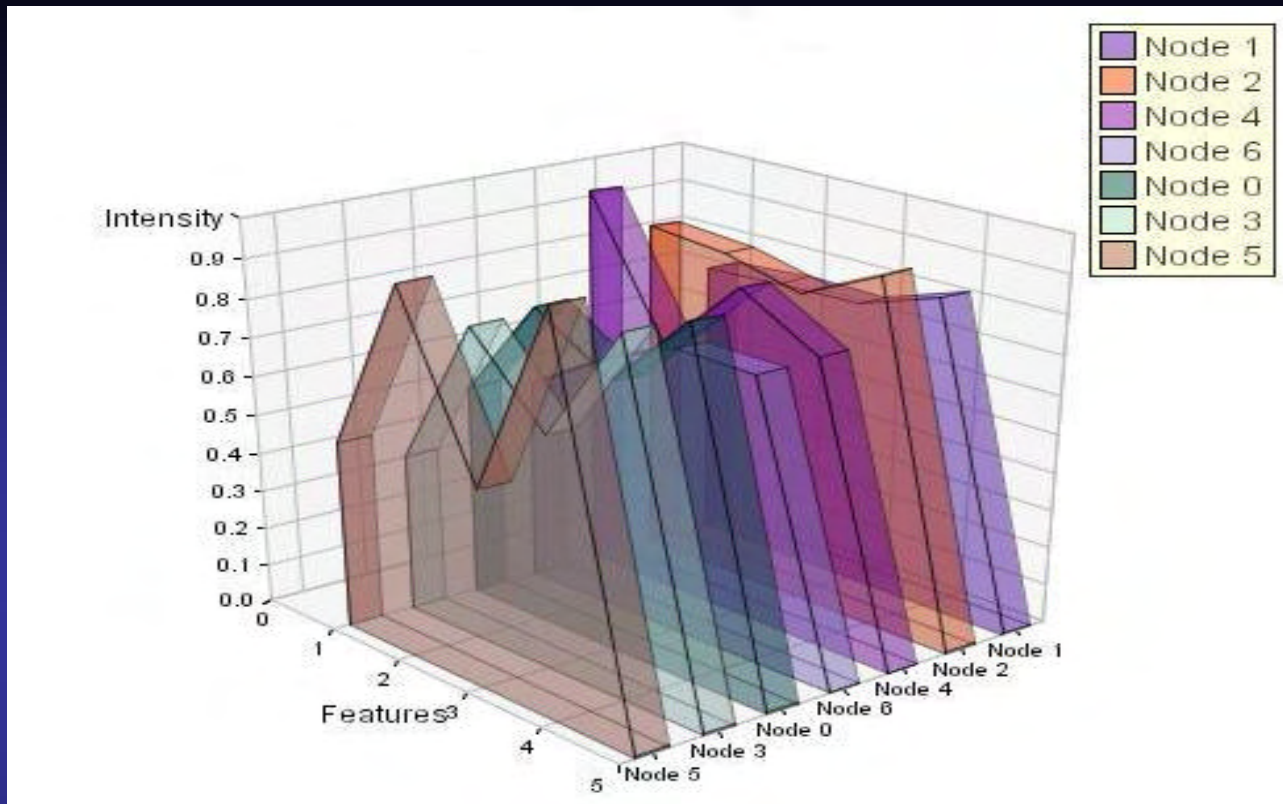
a = Sera donated before 3-dimensional color Doppler flow ultrasound confirmation of benign cysts.  
All patients followed 5 years after sera collection.

b = Sera donated before surgical staging and diagnosis.

100% Sensitivity  
95 % Specificity

### 3D Model Chart





**NEW MODEL:**

**BLINDED TEST RESULTS:**

**100% Specificity and Sensitivity**

**50/50 Cancers, 63/63 Unaffected or Benign**

# Artificial Intelligence-Based Proteomic Pattern Diagnostics Ovarian Cancer Results

BLINDED TEST DATA ( N= 116 PATIENTS)

NED (5 Year follow up) **22/24 (92%)**

Benign gynecologic and non-gyn inflammatory  
(cysts, fibroids) (RA, colitis, sinusitis) **41/42 (98%)**

**TOTAL SPECIFICITY: 63/66 = 95%**

Ovarian cancer Stage II, III, IV **32/32 (100%)**

Ovarian cancer Stage I **18/18 (100%)**

**TOTAL SENSITIVITY: 50/50 = 100%**

**POSITIVE PREDICTIVE VALUE: 95% VS. 20% FOR CA125**



# SERUM PROTEOMIC PATTERN DIAGNOSTICS

## BLINDED TEST RESULTS: PROSTATE CANCER

N= 266

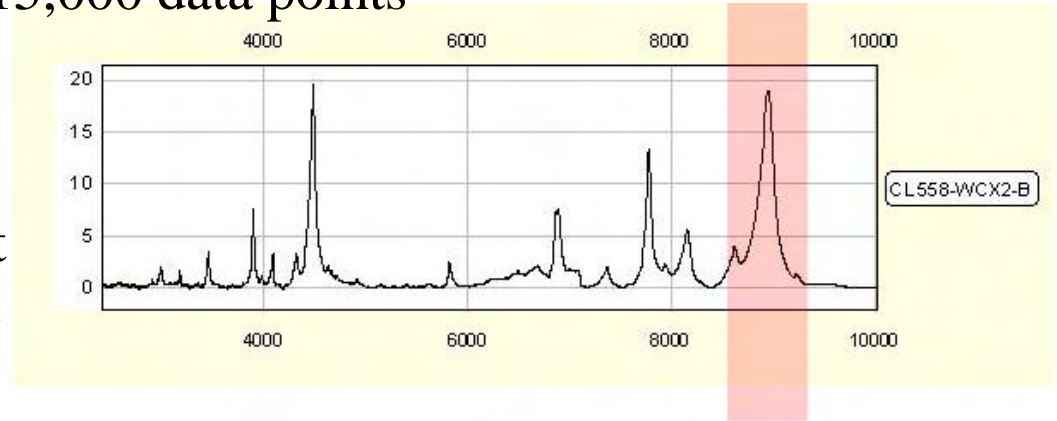
Predicted Diagnosis by  
Proteomic Pattern Analysis

Actual Diagnosis (serum PSA ng/ml)	N	CANCER (% total)	BENIGN (% total)
Prostate Cancer (> 4.0)	38	36 <b>(95%)</b>	2 <b>(5%)</b>
Benign (< 4.0)	75	5 <b>(7%)</b>	70 <b>(93%)</b>
Benign (>4.0)	153	46 <b>(30%)</b>	117 <b>(70%)</b>

# Ciphergen

- Low Resolution
- No protein peak ID
- MALDI-TOF ions do not necessarily reflect relative abundance in sera

15,000 data points

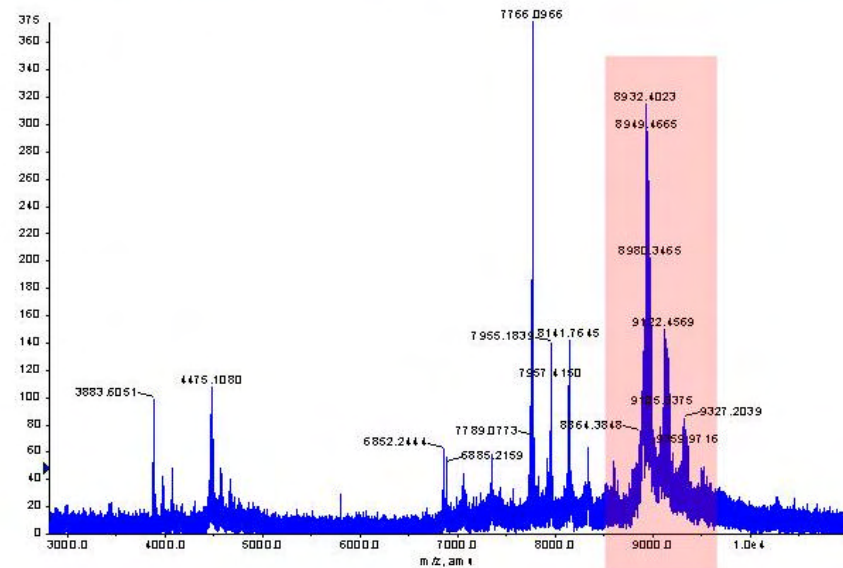


**The ABI QSTAR® Pulsar Hybrid LC/MS/MS System is a high performance hybrid quadrupole time-of-flight mass spectrometer**

+TOF MS: 35 MCA scans from Sample 2 (P) of CL558\_WCX2.W  
z=3.56000 4753643967006-004, ID=7.43988569479405680+001

Max. 3271.0 counts

ABI QSTAR  
900,000 data points



# ABI QqToF Qstar Ovarian Results:

## 100% Sensitive and Specific

- **Q-STAR VALIDATION** : Direct comparison to Ciphergen PBS II using the same WCX chips
- Independent analysis by Dr. Tim Veenstra and Dr. Thomas Conrads (Director, NCI Biomedical Proteomics Program)
- Serum sample: National Ovarian Cancer Early Detection Program (NOCEDP), Northwestern University; Director: Dr. David Fishman
- Total Number of samples:  
Unaffected = 95, Ovarian Cancer = 153

Samples were divided into 3 groups:

- A. 84 training samples (28 Unaffected and 56 Ovarian Cancer)
- B. 87 testing samples (30 Unaffected and 57 Cancer)
- C. 87 blind validation samples (37 Unaffected and 40 Cancer)

### Results:

- Two different models generated 100% sensitivity and specificity.
- These same model correctly classified 100% of the blinded samples.

Key Ions (m/z) values from one model:

1276.8612

2374.2444

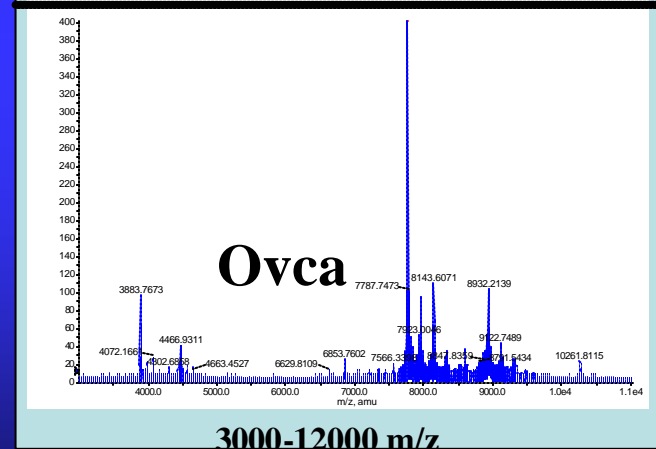
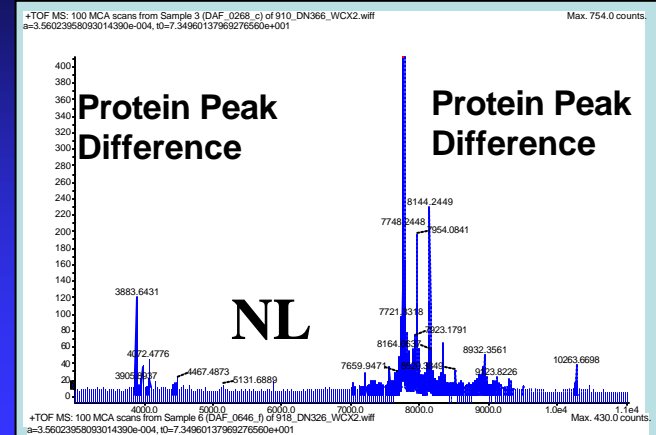
4292.900

7060.1210

8605.678

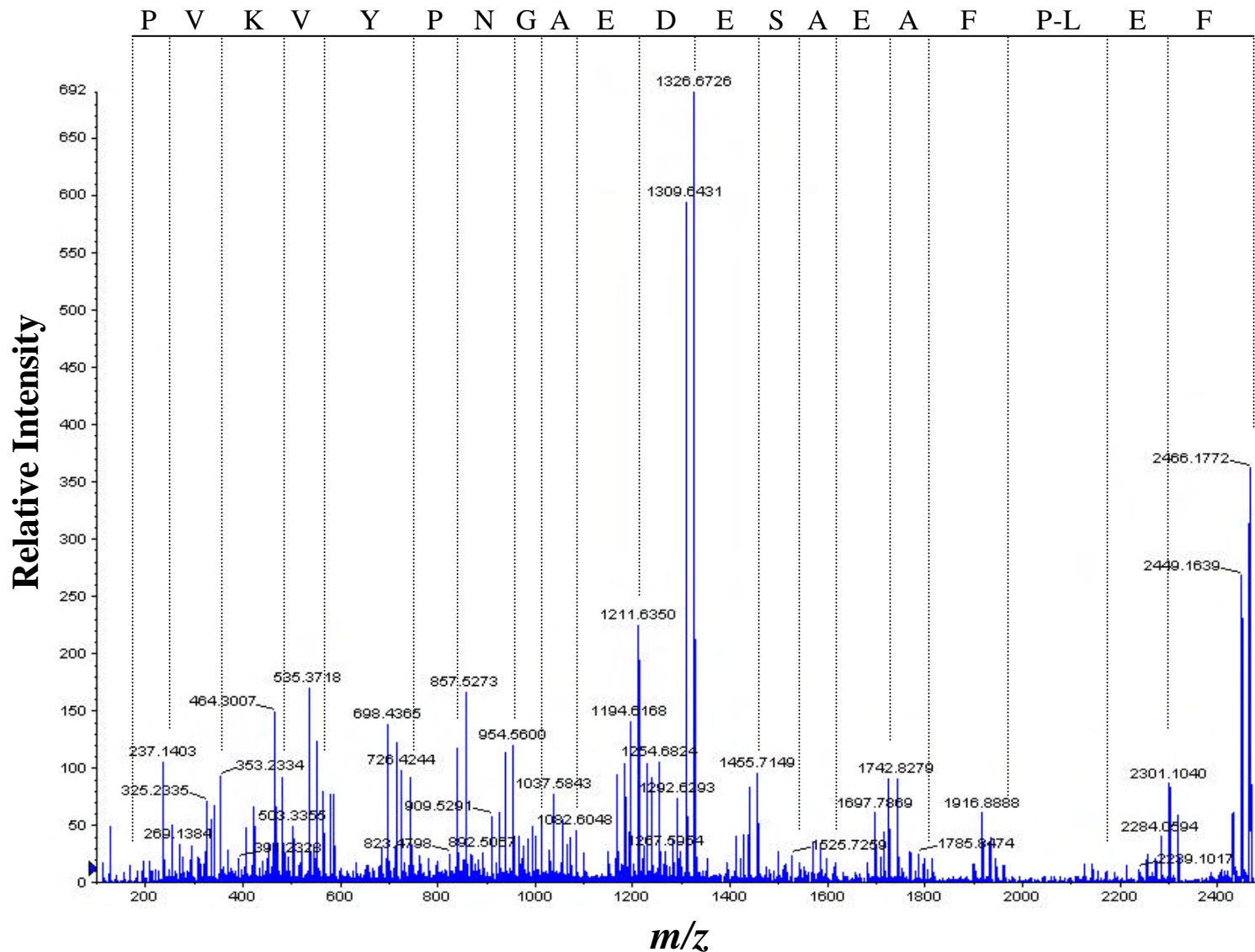
8706.065

9870.9375



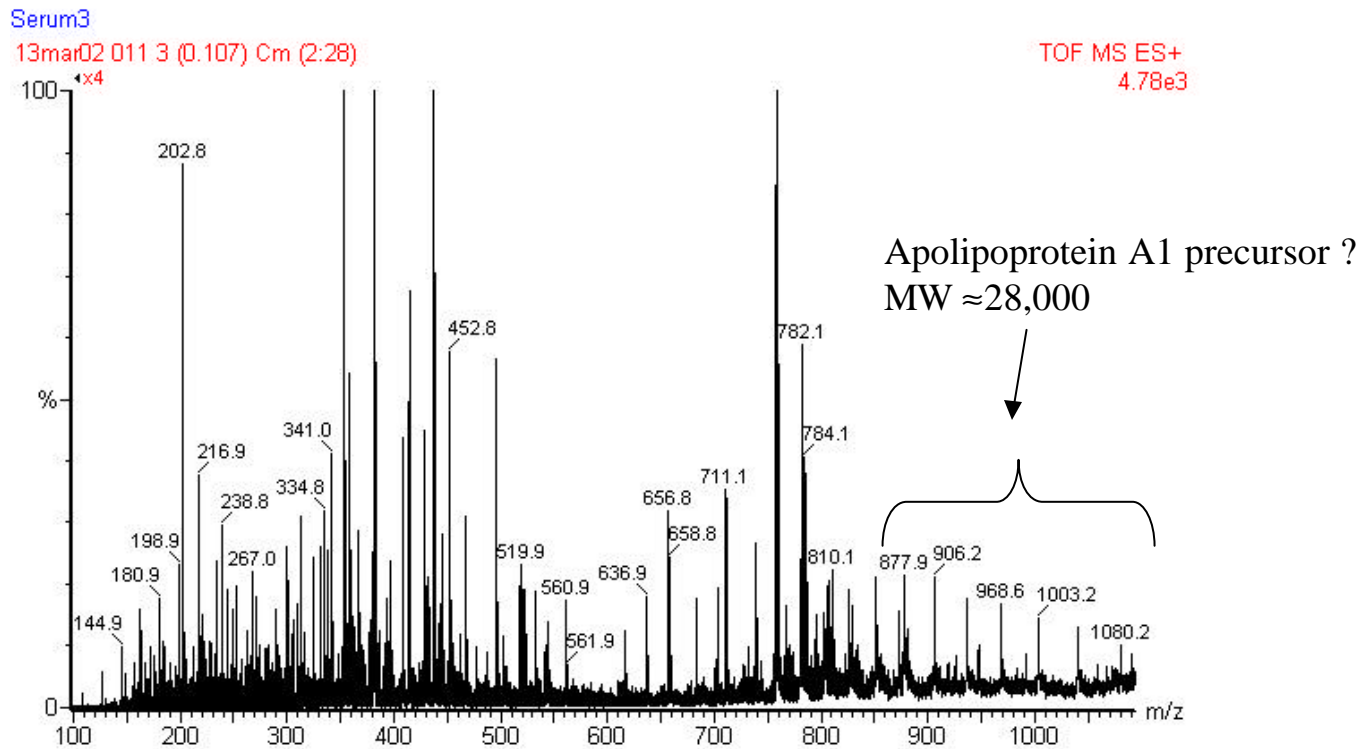
# Protein Peptide ID directly from complex Q-Star Spectra

## Adrenocorticotrophic hormone 2 clip 18-39 tandem MS spectrum

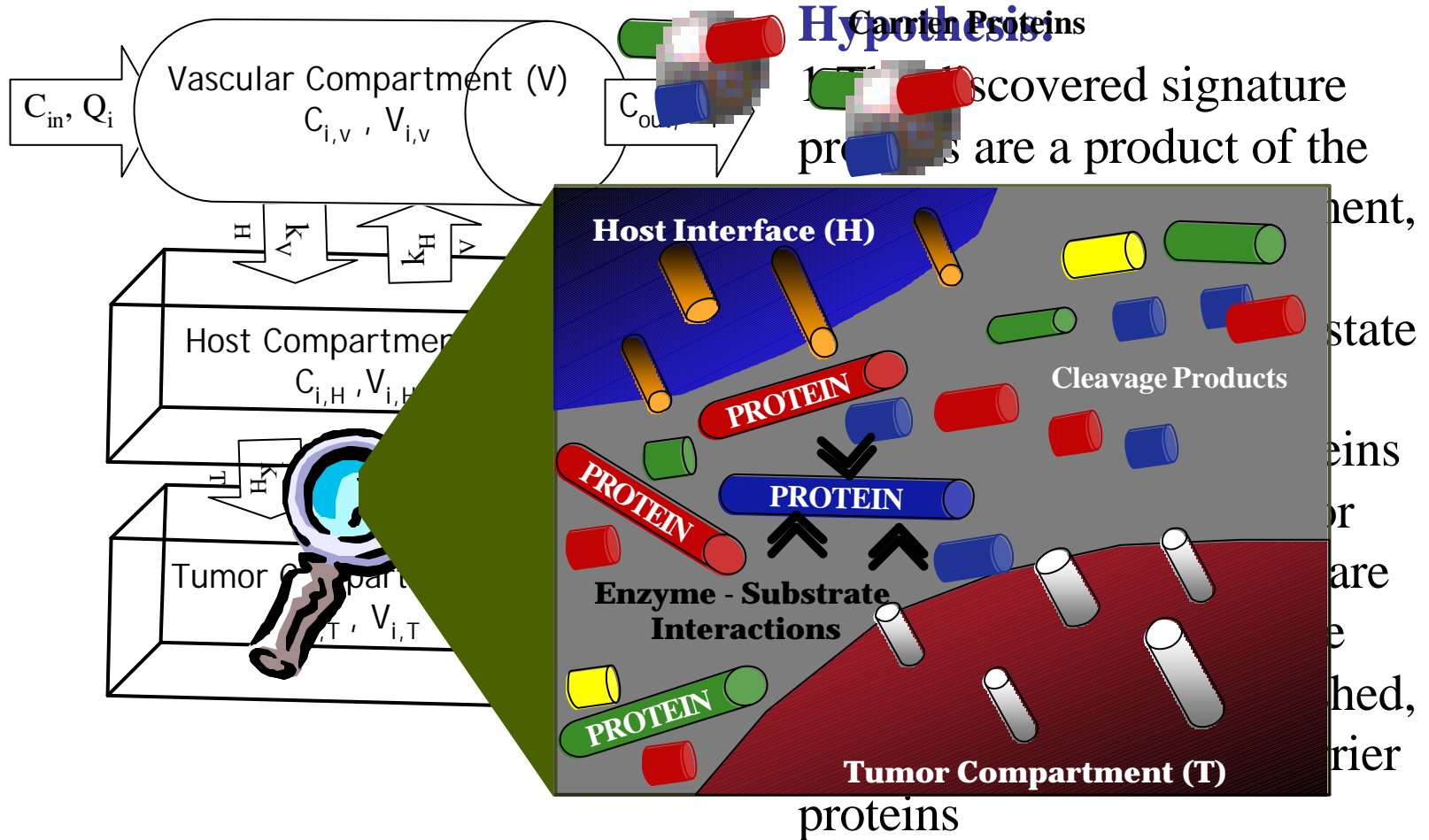


# Human serum - Micromass LCT and ESI-Chip

500 x diluted in 1:1 ACN:H<sub>2</sub>O, 0.2% formic acid



# Source of Ions Comprising Diagnostic Signature Subset



# **Nth Dimensional Proteomic Pattern Analysis:**

## **A New Paradigm for Diagnostics**

- **Requires less than 1 microliter of raw unfractionated serum**
- **MALDI-TOF profiles obtained in less than 5 minutes**
- **High throughput: Capacity to analyze more than 400 samples/day**
- **Nth-Dimensional analysis virtually instantaneous once training sets are defined**
- **Validated sera added into an ever-expanding training population: models get better and better as more data is analyzed**
- **Analysis compatible with web-centric platforms**

# Sistare Vasculitis Results 8-5-02

Vasculitis Samples

Negative = 21 samples

Positive = 16 samples

37 Total samples

19 Testing set (10 Negative and 9 Positive)

18 Training set (11 Negative and 7 Positive)

100% specificity and sensitivity

Validation with Renal Positive and Negative

Classified 15/15 (100%) Renal Negative as negative

Classified 8/11 (72.7%) Renal Positive as negative

Model:



Node	Count	State	StateSum	Error	475.495	875.625	1067.490	980.226	9551.137	2527.856	11683.457	6266.976	17988.113	4037.875
0	11	0	2	2	0.997	0.806	0.756	0.843	0.024	0.241	0.016	0.102	0.000	0.156
1	6	1	5	1	1.000	0.719	0.656	0.710	0.018	0.192	0.013	0.083	0.000	0.117
2	1	0	0	0	0.840	0.652	0.675	1.000	0.017	0.191	0.012	0.072	0.000	0.119



# Sistare Renal Results 8-5-02

Renal Samples

Negative = 15 samples

Renal = 11 samples

26 Total samples

10 Testing set (6 Negative and 4 Renal)

16 Training set (7 Negative and 7 Renal)

100% specificity and sensitivity

Validation with Vasculitis Positive and Negative

Classified 21/21 (100%) Vasculitis Negative as negative

Classified 16/16 (100%) Vasculitis Positive as positive



Model:

Node	Count	State	StateSum	Error	4183.515	12346.710	5396.381	10426.889	3605.287	14183.555	10286.063	3772.441
0	6	0	0	0	0.896	0.039	0.512	0.077	1.000	0.000	0.152	0.915
1	7	1	7	0	0.938	0.001	0.648	0.028	0.994	0.008	0.045	0.988
2	3	0	0	0	1.000	0.000	0.338	0.010	0.602	0.018	0.021	0.672

# Sistare Dox Results 8-5-02

Dox Samples

Negative = 79 samples

Positive = 19 samples

98 Total samples

50 Testing set (39 Negative and 11 Positive )

48 Training set (40 Negative and 8 Positive)

100% specificity and sensitivity

Validation with 46 unknown samples

Predicted 18/46 Positive

Predicted 28/46 Negative



Model:

Node	Count	State	StateSum	Error	14003.968	5136.478	2971.389	3631.956	8408.127	2639.668	2700.797	3995.735	3919.051	3996.903
0	1	Dox Positive	1	0	0.000	0.250	0.386	0.349	0.345	0.486	0.468	1.000	0.415	0.957
1	4	Dox Positive	3	1	0.000	0.283	0.543	0.446	1.000	0.695	0.666	0.568	0.443	0.561
4	1	Dox Positive	1	0	0.000	0.286	0.548	0.457	0.274	0.628	0.613	1.000	0.912	0.973
7	1	Dox Positive	1	0	0.000	0.368	0.682	0.540	0.310	0.827	0.789	1.000	0.783	0.986
2	1	Negative	0	0	0.000	0.175	0.644	0.461	0.886	0.885	0.847	1.000	0.491	0.946
3	13	Negative	1	1	0.000	0.418	0.831	0.636	0.241	1.000	0.966	0.668	0.608	0.662
5	8	Negative	0	0	0.000	0.420	0.776	0.638	0.919	0.959	0.925	0.959	0.644	0.938
6	18	Negative	1	1	0.000	0.432	0.812	0.648	0.485	0.994	0.951	0.923	0.657	0.901
8	1	Negative	0	0	0.000	0.378	0.764	0.609	0.980	1.000	0.966	0.702	0.589	0.703

# Sistare Cardiotoxicity Results 8-5-02

Cardiotoxicity Samples

Negative = 54 samples

Positive = 70 samples

124 Total samples

62 Testing set (29 Negative and 33 Positive )

62 Training set (25 Negative and 37 Positive)

91.6% specificity 81.5% sensitivity

Validation with 23 unknown samples

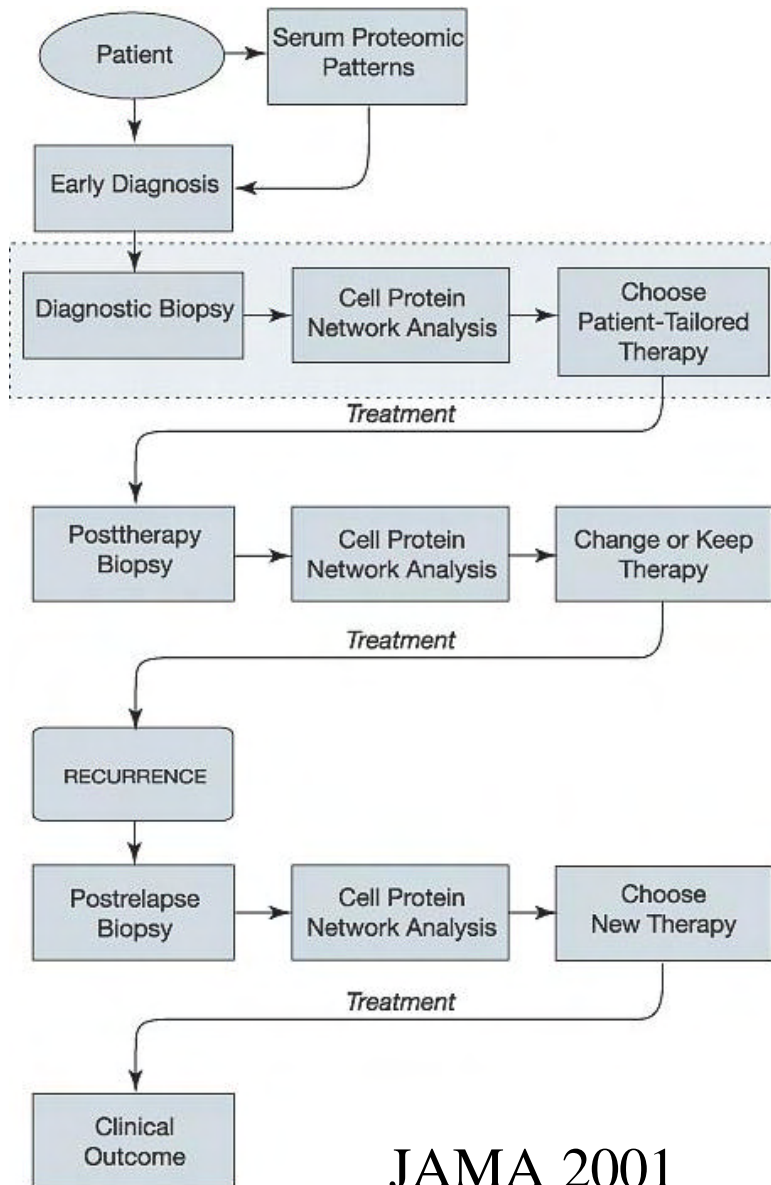
Predicted 7/23 Negative

Predicted 16/23 Positive

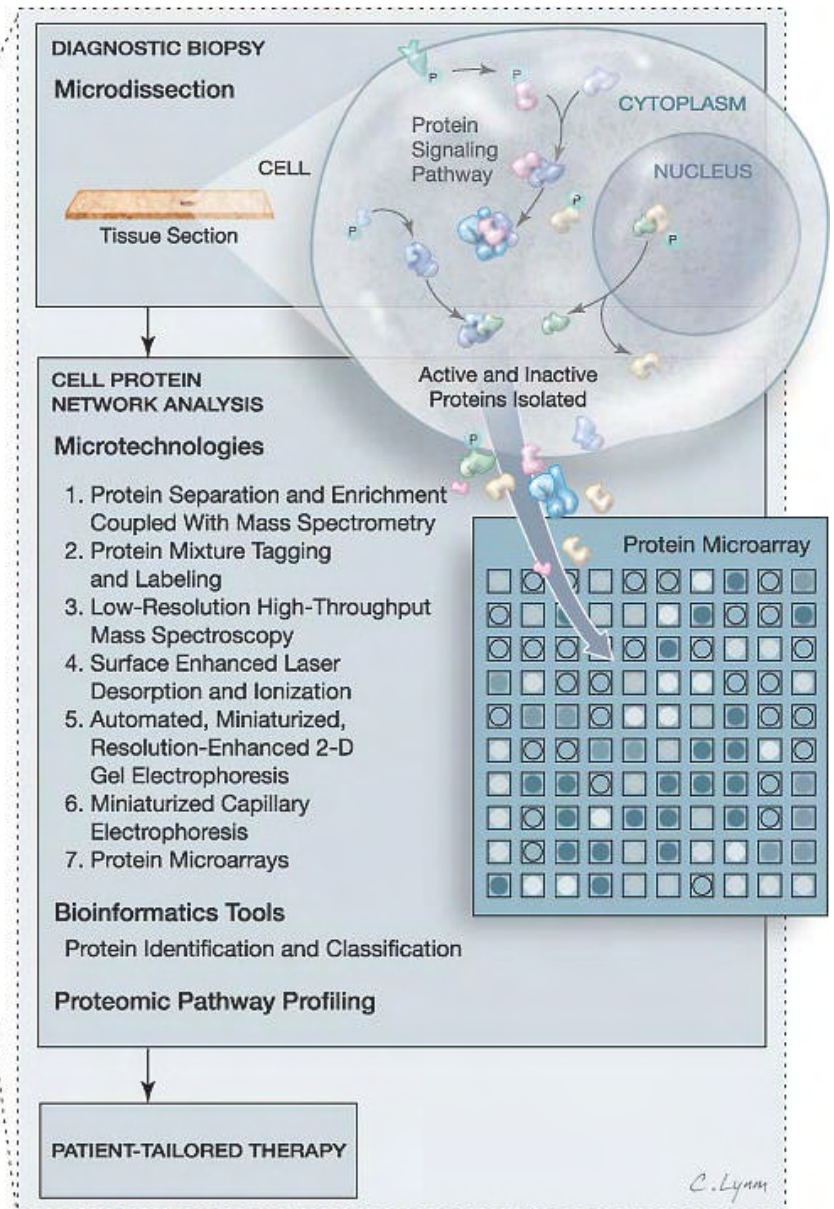
Model:



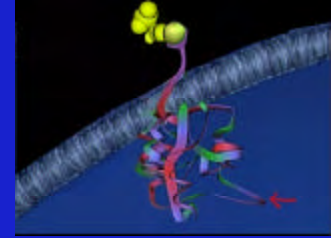
Node	Count	State	Error	493.592	984.298	6310.890	8526.993	577.758	577.758	7977.216	881.130	8865.985	16758.039
0	35	Cardiotox	10	0.976	0.735	0.096	0.041	0.980	0.980	0.048	0.886	0.045	0.000
1	30	Negative	6	1.000	0.574	0.101	0.059	0.753	0.753	0.083	0.661	0.087	0.000
2	45	Cardiotox	15	1.000	0.666	0.188	0.172	0.782	0.782	0.390	0.720	0.244	0.000
3	10	Cardiotox	2	1.000	0.456	0.133	0.141	0.566	0.566	0.331	0.501	0.185	0.000
4	4	Negative	1	1.000	0.807	0.251	0.274	0.897	0.897	0.667	0.858	0.351	0.000



JAMA 2001



# Serum Proteomic Pattern Diagnostics



- TISSUE PATHOLOGIC STATES ARE REFLECTED IN HIDDEN SERUM PROTEOMIC PATTERNS UNCOVERED USING AN ARTIFICIAL BIOINFORMATICS TOOL THAT LEARNS THE MOST FIT SOLUTION
- WE HYPOTHESIZE THAT SERUM PROTEOMIC PATTERNS ARE PRODUCT OF THE UNIQUE TUMOR-HOST MICROENVIRONMENT AND REFLECT TUMOR AND HOST INTERACTION

## CURRENT STRATEGY:

### TWO INDEPENDENT TRACKS:

1. SCIENTIFIC INVESTIGATION INTO SPECIFIC SOURCE AND IDENTITY OF THE CLASSIFIERS
2. NCI-BASED NATIONAL CLINICAL TRIAL ON SERUM PROTEOMIC PATTERN DIAGNOSTICS WHERE IDENTITY IS NOT NEEDED

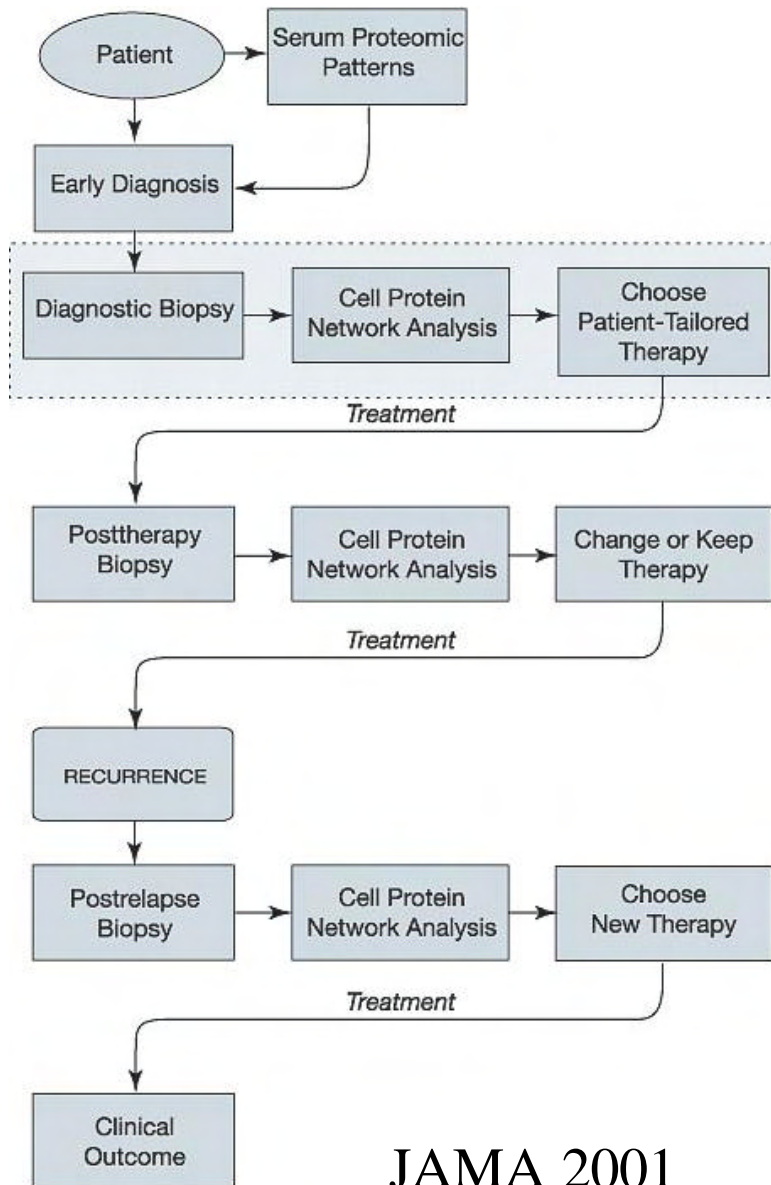
# CCR NCI Clinical Proteomics Reference Laboratory

- Phase One: Quality assurance, sensitivity, specificity, reproducibility, and exclusion of degraded samples, validation of initial results on larger retrospective study sets
- Phase Two:
  - A. Prospective longitudinal testing for ovarian cancer recurrence.
  - B. Classification of benign from malignant GYN disease in Women who have been diagnosed with a pelvic mass
  - C. High Risk Screening
- Phase Three: Extension to breast, prostate, lung, colon, and pancreatic cancer

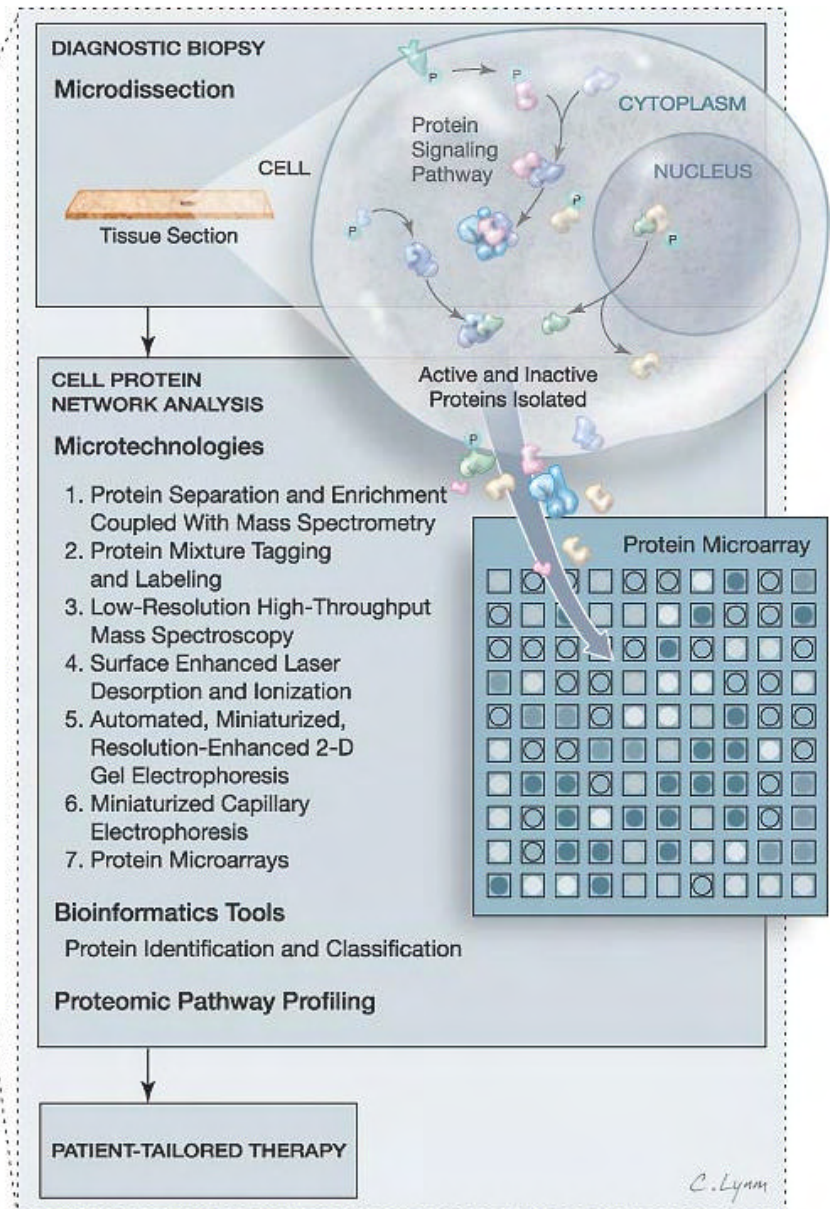
## Gynecologic Extramural Collaborators

- Northwestern Ovarian Cancer Early Detection Program, David Fishman
- Ovarian Cancer EDRN Consortium, David Fishman PI
- Pacific Ovarian Cancer Research Consortium, Marsha Rivken Center; Univ. Washington ovarian cancer SPORE, Nicole Urban, PI, Saul Rivken, Director
- University of Pennsylvania, George Coukos
- Memorial Sloan Kettering Cancer Center High Risk Screening Program, Ken Offit, PI
- Gynecologic Oncology Group clinical trial link newly diagnosed women, Elise Kohn PI
- InterSPORE collaboration for screening study (ovarian cancer SPORIS: U Wash, Urban confirmed; U Alabama, Partridge confirmed; Fox Chase Cancer Center, Hamilton/Daly) link for pelvic mass trial (protocol and formal SPORE collaboration mechanism in development)
- St. Bartholomew's Hospital, Ovarian Cancer Screening Program, Ian Jacobs, PI, Steve Skates, statistician
- Cancer Gene Network High Risk Women Pilot Program, through Steven Skates and CGN Steering Committee
- University of Alabama SPORE, support reference laboratory development, W. Grizzle
- Stony Brook and Long Island Jewish Hospital Consortium





JAMA 2001





**Ben Hitt**  
Chief Scientific Officer  
Correlogic Systems, Inc.

**FDA-NCI CLINICAL PROTEOMICS PROGRAM**  
**SCIENTIFIC TEAM AND COLLABORATORS**

Kristin Anderson  
Myron Gross

Jim Moeller  
**David Ornstein**

**Lance Liotta**

Mary Relling  
William Evans

**Vincent Fusaro**  
Sally Ross

Tim Veenstra  
Director, NCI Biomedical Proteomics Program  
Tom Conrads  
Deputy Director, NCI Biomedical Proteomics Program

**Elise Kohn**  
Monica Brown

**Cloud Paweletz**  
Verena Bichsel  
Ali Ardekani

**Michael Emmert-Buck**  
Michael Flaig  
Mamoun Ahram  
John Gillespie

Mark Raffeld  
Connie Hebeda  
Elaine Jaffe  
**David Fishman**  
**Gordon Mills**  
George Coukos

**Valerie Calvert**  
Rene Krieg  
Mike Kimmell

Marston Linehan

**Frank Sistare**  
Jun Zhang  
Eugene Herman

David Berman  
**Lu Charboneau**  
**Mary Winters**  
Paul Herrmann  
Nicole Simone  
Henry Krutzsch  
Lee Leak

Phil Taylor  
Mark Roth

Pat Steeg  
Julie Wulfkuhl  
Kelley McLean

**Kathryn Zoon**  
**Director CBER, FDA**  
**Amy Rosenberg**  
**Director DTP/OTRR CBER, FDA**

Bruce Trock  
Baljit Singh  
Marc Lippman

Dennis Sgroi  
Bruce Trock  
Fred Li

**Jay Siegel**  
**Director , OTRR CBER, FDA**

**J. Carl Barrett**  
**Director CCR, NCI**