

cobas® HPV

Qualitative nucleic acid test for use on the cobas[®] 6800/8800 Systems

For in vitro diagnostic use

cobas[®] HPV P/N: 07460155190

cobas[®] HPV Positive Control Kit P/N: 07460171190

cobas[®] Buffer Negative Control Kit P/N: 07002238190

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Intended use

cobas° HPV for use on the **cobas**° 6800/8800 Systems (**cobas**° HPV) is a qualitative *in vitro* test for the detection of Human Papillomavirus in clinician-collected cervical specimens using an endocervical brush/spatula or broom and placed in the ThinPrep° Pap Test™ PreservCyt° Solution. This test detects the high-risk HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68.

cobas® HPV is indicated for use for routine cervical cancer screening as per professional medical guidelines, including triage of ASC-US cytology, co-testing (or adjunctive screen) with cytology, and HPV primary screening of women to assess the risk for cervical precancer and cancer. Patients should be followed-up in accordance with professional medical guidelines, results from prior screening, medical history, and other risk factors.

Warning

cobas® HPV is NOT intended:

- for use in determining the need for treatment (i.e., excisional or ablative treatment of the cervix) in the absence of high-grade cervical dysplasia. Patients who are HPV16/18 positive should be monitored carefully for the development of high-grade cervical dysplasia according to current practice guidelines.
- for women who have undergone hysterectomy.
- for use with samples other than those collected by a clinician using an endocervical brush/spatula or a cervical broom and placed in the ThinPrep® Pap Test™ PreservCyt® Solution.

HPV-negative cancers of the cervix do occur in rare circumstances. Also, no cancer screening test is 100% sensitive. Use of this device for primary cervical cancer screening should be undertaken after carefully considering the performance characteristics put forth in this label, as well as recommendations of professional guidelines.

The use of this test has not been evaluated for the management of women with prior ablative or excisional therapy, hysterectomy, who are pregnant or who have other risk factors (e.g., HIV+, immunocompromised, history of sexually transmitted infections).

Summary and explanation of the test

Background and rationale for HPV testing

Human papillomavirus (HPV) is a small, non-enveloped, double-stranded DNA virus, with a genome of approximately 8000 nucleotides. There are more than 140 different HPV genotypes^{3,4} and approximately 40 different genotypes can infect the human anogenital mucosa. Fourteen HPV genotypes are classified as carcinogenic or highrisk (HR): 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68. Please note that one of these, HPV66, was recently categorized as "possibly carcinogenic" based on its relatively low prevalence in invasive cervical carcinomas. 99

Persistent infection with these high risk HPV genotypes is the central cause of cervical cancer and its precursor cervical intraepithelial neoplasia (CIN).⁶ Sexually transmitted infections with HPV is extremely common, with estimates of up to 75% of all women experiencing exposure to HPV at some point.¹⁰ However, most infections clear within 1-2 years.¹⁰ Most cervical cancers and deaths from cervical cancer can be prevented through early detection of pre-cancerous lesions in the cervix, leading to timely treatment. In developed countries with cervical cancer screening programs, the Pap smear has been used since

the mid-1950s as the primary tool to detect early precursors to cervical cancer. Although it has decreased the death rates due to cervical cancer dramatically in those countries, the Pap smear and subsequent liquid based cytology methods require interpretation by highly trained cytopathologists and have a high rate of false negatives. Cytological abnormalities are primarily due to infection with HPV; however, various inflammatory or sampling variations can result in false positive cytology results. Triage of an abnormal cytology result involves repeat testing, colposcopy and biopsy to rule out the presence of high-grade precancerous lesions, (cervical intraepithelial neoplasia of grade 2 or higher; ≥CIN2). Therefore, tests that detect infection with these HR HPV genotypes are now being used increasingly in cervical cancer screening programs to improve the prevention of cervical cancers and clinical patient management.¹² Nucleic acid (DNA) testing by PCR is a non-invasive method for determining the presence of a cervical HPV infection. Proper implementation of nucleic acid testing for HPV may increase the sensitivity of cervical cancer screening programs by detecting high-risk lesions earlier in women 25 years and older and reducing the need for unnecessary colposcopy and treatment in patients 21 and older with atypical squamous cells of undetermined significance (ASC-US) cytology. Therefore, tests that detect infection with these HR HPV genotypes are now being used increasingly in cervical cancer screening programs to improve the prevention of cervical cancers.¹²

The 2006 Consensus Guidelines for the Management of Women with Abnormal Cervical Cancer Screening Tests recognized the utility of using a combination of cervical cytology, tests for HPV detection, and type-specific HPV testing for women undergoing screening for cervical cancer. One of the earliest and most common utilization of HPV testing has been for the management (referral to colposcopy) of women with equivocal cervical cytologic abnormalities (ASC-US). Further, revised and updated guidelines now recommend the combination of cytology and HPV testing (cotesting) as the preferred method of screening in women \geq 30 years, with HPV 16/18 genotype-specific testing as an added option to triage women with negative cytology to colposcopy. A later revision provided the option when cotesting to follow up women with low grade squamous intraepithelial lesion (LSIL)/HPV negative results in 12 months rather than refer to colposcopy. Most recently, interim guidance has been issued for HR HPV DNA testing to be used as a first-line primary screening test in women \geq 25 years.

Nucleic acid (DNA) testing by PCR is a non-invasive method for determining the presence of a cervical HPV infection. Proper implementation of nucleic acid testing for HPV may increase the sensitivity of cervical cancer screening programs by detecting high-risk lesions earlier in women 25 years and older and reducing the need for unnecessary colposcopy and treatment in patients 21 and older with ASC-US cytology.

Explanation of the test

cobas° HPV is a qualitative real-time 17,18 PCR test that detects 14 high-risk HPV genotypes. **cobas**° HPV uses primers to define a sequence of approximately 200 nucleotides within the polymorphic L1 region of the HPV genome. A pool of HPV primers present in the Master Mix is designed to amplify HPV DNA from 14 high-risk types (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68). 7,19,20,21,22,23,24 **cobas**° HPV utilizes β -globin DNA as an internal control to monitor the entire sample preparation and PCR amplification process so an additional primer pair targets the human β -globin gene (330 base pair amplicon). Fluorescent oligonucleotide probes bind to polymorphic regions within the sequence defined by these primers. In addition, the test utilizes a low titer positive and a negative control.

Principles of the procedure

cobas° HPV is based on fully automated sample preparation (nucleic acid extraction and purification) followed by PCR amplification²⁵ and detection. The **cobas**° 6800/8800 Systems consist of the sample supply module, the transfer module, the processing module, and the analytic module. Automated data management is performed by the **cobas**° 6800/8800 software which assigns test results for all tests as positive, negative or invalid. Results can be reviewed directly on the system screen,

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exported, or printed as a report.

Nucleic acid (DNA) from patient samples is extracted. In summary, nucleic acid is released by addition of proteinase and lysis reagent to the sample. The released nucleic acid binds to the silica surface of the added magnetic glass particles. Unbound substances and impurities, such as denatured protein, cellular debris and potential PCR inhibitors are removed with subsequent wash steps and purified nucleic acid is eluted from the magnetic glass particles with elution buffer at elevated temperature. External controls (positive and negative) are processed in the same way with each **cobas**® HPV run.

A thermostable DNA polymerase enzyme is used for PCR amplification. The HPV and β -globin sequences are amplified simultaneously utilizing a universal PCR amplification profile with predefined temperature steps and number of cycles. The master mix includes deoxyuridine triphosphate (dUTP), instead of deoxythimidine triphosphate (dTTP), which is incorporated into the newly synthesized DNA (amplicon). Any contaminating amplicon from previous PCR runs are eliminated by the AmpErase enzyme, which is included in the PCR master mix, during the first thermal cycling step. However, newly formed amplicon are not eliminated since the AmpErase enzyme is inactivated once exposed to temperatures above 55°C.

cobas* HPV master mix contains detection probes specific for twelve High Risk HPV target sequences, one detection probe specific for the HPV16 target sequence, one detection probe specific for the HPV18 target sequence and one for β -globin. The amplified signal from twelve high-risk HPV types (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68) is detected using the same fluorescent dye while HPV16, HPV18 and β -globin signals are each detected with their own dedicated fluorescent dye. When not bound to the target sequence, the fluorescent signal of the intact probes is suppressed by a quencher dye. During the PCR amplification step, hybridization of the probes to the specific single-stranded DNA template results in cleavage of the probe by the 5' to 3' exonuclease activity of the DNA polymerase resulting in separation of the reporter and quencher dyes and the generation of a fluorescent signal. With each PCR cycle, increasing amounts of cleaved probes are generated and the cumulative signal of the reporter dye increases concomitantly. Real-time detection and discrimination of PCR products is accomplished by measuring the fluorescence of the released reporter dyes for the HPV targets and β -globin, respectively.

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Reagents and materials

cobas® HPV reagents and controls

Table 1 cobas® HPV

cobas[®] HPV Store at 2-8°C

480 test cassette (P/N 07460155190)

Kit components	Reagent ingredients	Quantity per kit 480 tests	
Proteinase Solution (PASE) Tris buffer, < 0.05% EDTA, Calcium chloride, Calcium acetate, 8% Proteinase EUH210: Safety data sheet available on request. EUH208: Contains Subtilisin. May produce an allergic reaction.		38 mL	
Empty Vessel (EV)	N/A	1	
Elution Buffer (EB)	Tris buffer, 0.2% Methyl-4 hydroxibenzoate	38 mL	
Master Mix Reagent 1 (MMX-R1)	Manganese acetate, Potassium hydroxide, < 0.1% Sodium azide	14.5 mL	
HPV Master Mix Reagent 2 (HPV MMX-R2)	Tricine buffer, Potassium acetate, EDTA, Glycerol, < 18% Dimethyl sulfoxide, <0.12% dATP, dCTP, dGTP, dUTPs, < 0.1% Tween 20, < 0.1% Sodium azide, < 0.1% Z05 DNA polymerase, < 0.10% AmpErase (uracil N-glycosylase) enzyme (microbial), < 0.1% Upstream and downstream HPV primers, < 0.01% Upstream and downstream β -globin primers, < 0.01% Fluorescent-labeled oligonucleotide probes specific for HPV and β -globin, < 0.01% Oligonucleotide aptamer	17.5 mL	

Table 2 cobas[®] HPV Positive Control Kit

cobas® HPV Positive Control Kit

Store at 2-8°C (P/N 07460171190)

Kit components	Reagent ingredients	Quantity per kit
HPV Positive Control (HPV (+) C)	Tris buffer, < 0.05% EDTA, < 0.1% Sodium azide, < 0.01% Non-infectious plasmid DNA (microbial) containing HPV16, HPV18 and HPV39 sequences, < 0.01% Non-infectious plasmid DNA (microbial) containing β -globin sequences, < 0.002% Poly rA RNA (synthetic)	16 mL (16 x 1mL)

Table 3 cobas® Buffer Negative Control Kit

cobas® Buffer Negative Control Kit Store at 2-8°C

(P/N 07002238190)

Kit components	Reagent ingredients	Quantity per kit
cobas® Buffer Negative Control (BUF (-) C)	Tris buffer, < 0.1% sodium azide, EDTA, < 0.002% Poly rA RNA (synthetic)	16 mL (16 x 1 mL)

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cobas omni reagents for sample preparation

Table 4 cobas omni reagents for sample preparation*

Reagents	Reagent ingredients	Quantity per kit	Safety symbol and warning**
cobas omni MGP Reagent (MGP)	Magnetic glass particles, Tris buffer, 0.1% methyl-4 hydroxybenzoate, < 0.1% sodium azide	480 tests	Not applicable
Store at 2–8°C (P/N 06997546190)			
cobas omni Specimen Diluent (SPEC DIL)	Tris buffer, 0.1% methyl-4 hydroxybenzoate, < 0.1% sodium azide	4 x 875 mL	Not applicable
Store at 2–8°C (P/N 06997511190)			
cobas omni Lysis Reagent (LYS) Store at 2–8°C	42.56% (w/w) guanidine thiocyanate***, 5% (w/v) polydocanol***, 2% (w/v) dithiothreitol***, dihydro sodium citrate	4 x 875 mL	
(P/N 06997538190)			DANGER
			H302 + H332 Harmful if swallowed or if inhaled.
			H314 Causes severe skin burns and eye damage.
			H412 Harmful to aquatic life with long lasting effects.
			EUH032 Contact with acids liberates very toxic gas.
			P261 Avoid breathing dust/fume/gas/mist/vapours/spray.
			P273 Avoid release to the environment.
			P280 Wear protective gloves/ protective clothing/ eye protection/ face protection.
			P303 + P361 + P353 IF ON SKIN (or hair): Take off
			immediately all contaminated clothing. Rinse skin with water.
			P304 + P340 + P310 IF INHALED: Remove person to fresh air and keep comfortable for breathing. Immediately call a POISON CENTER/doctor.
			P305 + P351 + P338 + P310 IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. Immediately call a POISON CENTER/doctor.
			593-84-0 guanidinium thiocyanate
			9002-92-0 Polidocanol
			3483-12-3 (R*,R*)-1,4-dimercaptobutane-2,3-diol
cobas omni Wash Reagent (WASH)	Sodium citrate dihydrate, 0.1% methyl-4 hydroxybenzoate	4.2 L	Not applicable
Store at 15–30°C (P/N 06997503190)			

^{*} These reagents are not included in the **cobas**® HPV kit. See listing of additional materials required (Table 7).

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^{**} Product safety labeling primarily follows EU GHS guidance

^{***} Hazardous substance

Reagent storage and handling requirements

Reagents shall be stored and handled as specified in Table 5 and Table 6.

When reagents are not loaded on the **cobas**° 6800/8800 Systems, store them at the corresponding temperature specified in Table 5.

Table 5 Reagent storage (when reagent is not on the system)

Reagent	Storage temperature
cobas® HPV	2–8°C
cobas® HPV Positive Control Kit	2-8°C
cobas® Buffer Negative Control Kit	2-8°C
cobas omni Lysis Reagent	2-8°C
cobas omni MGP Reagent	2-8°C
cobas omni Specimen Diluent	2-8°C
cobas omni Wash Reagent	15-30°C

Reagents loaded onto the **cobas**° 6800/8800 Systems are stored at appropriate temperatures and their expiration is monitored by the system. The **cobas**° 6800/8800 Systems allow reagents to be used only if all of the conditions shown in Table 6 are met. The system automatically prevents use of expired reagents. Table 6 describes the reagent handling conditions enforced by the **cobas**° 6800/8800 Systems.

Table 6 Reagent expiry conditions enforced by the cobas® 6800/8800 Systems

Reagent	Open-kit stability	Number of runs for which this kit can be used	On-board stability (cumulative time on board outside refrigerator)
cobas® HPV	90 days from first usage	Max 20 runs	Max 20 hours
cobas® HPV Positive Control Kit	Not applicable	Max 16 runs	Max 10 hours
cobas® Buffer Negative Control Kit	Not applicable	Max 16 runs	Max 10 hours
cobas omni Lysis Reagent	30 days from loading*	Not applicable	Not applicable
cobas omni MGP Reagent	30 days from loading*	Not applicable	Not applicable
cobas omni Specimen Diluent	30 days from loading*	Not applicable	Not applicable
cobas omni Wash Reagent	30 days from loading*	Not applicable	Not applicable

^{*} Time is measured from the first time that reagent is loaded onto the **cobas**® 6800/8800 Systems.

Additional equipment and materials required but not provided

Table 7 Equipment, materials and consumables required for use with cobas® HPV

Material	P/N
cobas omni Processing Plate	05534917001
cobas omni Amplification Plate	05534941001
cobas omni Pipette Tips	05534925001
cobas omni Liquid Waste Container	07094388001
cobas omni Lysis Reagent	06997538190
cobas omni MGP Reagent	06997546190
cobas omni Specimen Diluent	06997511190
cobas omni Wash Reagent	06997503190
Solid Waste Bag with Insert	08030073001
Kit Drawer Solid Waste Update	08387281001
Tubes, 13 mL Round Base, for use as secondary sample tubes*	07958048190
Caps, neutral color	07958056190
Heat-resistant barcode labels**	RACO Industries, RAC-225075-9501
Vortex Mixer (single tube)	Any vendor
MPA RACK 16 MM LIGHT GREEN 7001-7050 ^{a,b}	03143449001
RD5 RACK – RD Standard rack 0001-0050 LR ^{a,b}	11902997001

^{*} Use of tubes other than those recommended above must be verified by user prior to implementation into **cobas**® HPV workflow in the laboratory.

^{**} For further details on barcode specifications refer to the **cobas**® 6800/8800 Systems User Guide. Use of barcode labels other than those recommended above must be verified by user prior to implementation into **cobas**® HPV workflow in the laboratory.

^a MPA 16mm and RD5 racks are required to use **cobas**[®] HPV. Contact your local Roche representative for a detailed order list for sample racks, racks for clotted tips and rack trays accepted on the instruments.

^b MPA 16mm rack is the preferred rack. If RD5 racks are used, ensure sample tubes are filled with the recommended minimum sample input volume. The tubes sit higher in an RD5 rack because of the rubber gasket at the bottom of each tube position. It is therefore possible that when using RD5 racks, the system could accept tubes containing less than the minimum sample input volume and cause pipetting errors later in the run.

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Table 8 Specimen collection kits for use with cobas® HPV

Collection Kit	P/N
ThinPrep® Pap Test TM Physician's Kit (500 vials & Broom-like collection devices) ThinPrep® Pap Test TM Physician's Kit (500 vials & Cytobrush/spatula collection devices)	Hologic 70136-001 Hologic 70136-002
Rovers® Cervex-Brush® Combi (500/Box)	VWR 89171-022
Cytobrush Plus GT - 25 Bags, 100 Brushes each (2,500/Box)	Medscand C0105
Cytobrush Plus GT - 2 Bags, 500 Brushes each (1,000/Box)	Medscand C0121
Cytobrush Plus GT - 10 Bags, 10 Brushes each (100/Box)	Medscand C0104
Cytobrush Plus GT Sterile - 1 Brush per Pouch (40/Box)	Medscand C0112
Cytobrush Plus GT Scored – 25 Bags, 100 Brushes each (2,500/Box)	Medscand C0305
Pap-Perfect Plastic Spatulas (500/Box)	Medscand 11080
42mm Replacement Caps for Vials (8 trays of 48/box)	07682247001 (optional)

Instrumentation and software required

The **cobas**° 6800/8800 software and **cobas**° HPV analysis packages shall be installed on the instrument(s). The Instrument Gateway (IG) server will be provided with the system(s).

Table 9 Instrumentation

Equipment	P/N
cobas® 6800 System (Moveable Platform)	05524245001 and 06379672001
cobas® 6800 System (Fixed Platform)	05524245001 and 06379664001
cobas® 8800 System	05412722001
Sample Supply Module	06301037001
Instrument Gateway	06349595001

Precautions and handling requirements

Warnings and precautions

As with any test procedure, good laboratory practice is essential to the proper performance of this assay. Due to the high sensitivity of this test, care should be taken to keep reagents and amplification mixtures free of contamination.

- For *in vitro* diagnostic use only.
- For prescription use only.
- All patient samples should be handled as if infectious, using good laboratory procedures as outlined in Biosafety in Microbiological and Biomedical Laboratories²⁷ and in the CLSI Document M29-A4.²⁸ Only personnel proficient in handling infectious materials and the use of cobas* HPV and cobas* 6800/8800 Systems should perform this procedure.
- All human-sourced materials should be considered potentially infectious and should be handled with universal precautions. If spillage occurs, immediately disinfect with a freshly prepared solution of 0.5% sodium hypochlorite in distilled or deionized water (dilute household bleach 1:10) or follow appropriate site procedures.
- Do not freeze any samples stored in primary or secondary tubes.
- Use only supplied or specified required consumables to ensure established test performance.
- Safety Data Sheets (SDS) are available on request from your local Roche representative.
- Closely follow procedures and guidelines provided to ensure that the test is performed correctly. Any deviation from the procedures and guidelines may affect established test performance.
- False positive results may occur if carryover of samples is not adequately controlled during sample handling and processing.

Reagent handling

- Handle all reagents, controls, and samples according to good laboratory practice in order to prevent carryover of samples, reagents, or controls.
- Before use, visually inspect each reagent cassette, diluent, lysis reagent, and wash reagent to ensure that there are no signs of leakage. If there is any evidence of leakage, do not use that material for testing.
- **cobas omni** Lysis Reagent contains guanidine thiocyanate, a potentially hazardous chemical. Avoid contact of reagents with the skin, eyes, or mucous membranes. If contact does occur, immediately wash with generous amounts of water; otherwise, burns can occur.
- Do not allow **cobas omni** Lysis Reagent, which contains guanidine thiocyanate, to contact sodium hypochlorite (bleach) solution. This mixture can produce a highly toxic gas.
- Expended control kits contain pierced vials with residual reagent; special care should be taken during disposal to avoid spills and contact.

- **cobas**° HPV Kit, **cobas**° HPV Positive Control Kit, **cobas**° Buffer Negative Control Kit, **cobas omni** MGP Reagent, and **cobas omni** Specimen Diluent contain sodium azide as a preservative. Avoid contact of reagents with the skin, eyes, or mucous membranes. If contact does occur, immediately wash with generous amounts of water; otherwise, burns can occur. If these reagents are spilled, dilute with water before wiping dry.
- Dispose of all materials that have come in contact with samples and reagents in accordance with country, state, and local regulations.

Good laboratory practice

- Do not pipette by mouth.
- Do not eat, drink, or smoke in designated work areas.
- Wear laboratory gloves, laboratory coats, and eye protection when handling samples and reagents. Avoid
 contaminating gloves when handling samples and controls. Gloves must be changed between handling
 samples and cobas* HPV Kit, cobas* HPV Positive Control Kit, cobas* Buffer Negative Control Kit and
 cobas omni reagents to prevent contamination.
- Wash hands thoroughly after handling samples and reagents, and after removing the gloves.
- Thoroughly clean and disinfect all laboratory work surfaces with a freshly prepared solution of 0.5% sodium hypochlorite in distilled or deionized water (dilute household bleach 1:10). Follow by wiping the surface with 70% ethanol.
- If spills occur on the instrument, follow the instructions in the **cobas*** 6800/8800 Systems User Guide to properly clean and decontaminate the surface of instrument(s).

Specimen collection, transport, and storage

Note: Handle all samples and controls as if they are capable of transmitting infectious agents.

Specimen collection

Cervical specimens collected in PreservCyt* Solution have been validated for use with **cobas*** HPV. Follow the manufacturer's instructions for collecting cervical specimens.

Specimen transport

Cervical specimens collected in PreservCyt* Solution can be transported at 2-30°C. Transportation of HPV specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.²⁹

Specimen storage

Cervical specimens collected in PreservCyt* Solution may be stored at 2-30°C for up to 3 months after the date of collection prior to performing **cobas*** HPV. See PreservCyt* Solution labeling for medium storage requirements. PreservCyt* specimens should not be frozen.

Instructions for use

Procedural notes

- Do not use **cobas*** HPV Kit, **cobas*** HPV Positive Control Kit, **cobas*** Buffer Negative Control Kit, or **cobas omni** reagents after their expiry dates.
- Do not reuse consumables. They are for one-time use only.
- Ensure that specimen barcode labels on sample tubes are visible through the openings on the side of the sample racks. Refer to the **cobas*** 6800/8800 Systems User Guide for proper barcode specifications and additional information on loading sample tubes.
- Refer to the **cobas*** 6800/8800 Systems User Guide for proper maintenance of instruments.

Running cobas® HPV

cobas° HPV can be run with a minimum required sample volume of 1.0 mL. The operation of the instrument is described in detail in the **cobas**° 6800/8800 Systems User's Guide.

Figure 1 summarizes the procedure.

It is necessary to aliquot specimens into barcoded 13 mL round-bottom secondary tubes for processing on the **cobas**° 6800/8800 Systems. Use pipettes with aerosol-barrier or positive-displacement tips to handle specimens.

- A single run can have specimen tested with either the HPV High Risk (HPV-HR) or HPV High Risk Plus Genotyping (HPV-GT) ASAPs.
- Specimens should be processed using the "PreservCyt" sample type selection in the user interface (UI) of **cobas** HPV.

Specimen preparation

- 1. Prepare a barcoded 13 mL round-bottom secondary tube for each PreservCyt® specimen to be tested.
- 2. With clean gloved hands, **vortex** each PreservCyt* primary specimen vial for **10 seconds** immediately prior to transfer.
- 3. Uncap a primary vial and transfer at least **1.0 mL** but no more than **4.0 mL** into the prepared barcoded secondary tube from step 1. *Always use caution when transferring specimens from primary containers to secondary tube*. *Always use a new pipette tip for each specimen*. Transfer tube to a rack (or cap the secondary tube if testing will be performed at a future time).
- 4. Re-cap the primary vial with a replacement cap before moving to the next specimen. Store the primary vial upright.
- Load the racks of uncapped secondary tubes into the Sample Supply Module and process on the cobas* 6800/8800 Systems for HPV testing.

Figure 1 cobas® HPV procedure

- 1 Log onto the system
 - Press Start to Prepare the system
 - Order Tests

Choose "PreservCyt" for ordering specimens collected in PreservCyt® Solution

- 2 Refill reagents and consumables as prompted by the system
 - · Load test specific reagent cassette
 - Load control cassettes
 - · Load pipette tips
 - · Load processing plates
 - Load MGP Reagent
 - · Load amplification plates
 - Refill Specimen Diluent
 - Refill Lysis Reagent
 - Refill Wash Reagent
- 3 Loading specimens onto the system
 - For each primary PreservCyt[®] specimen vial:
 - o Vortex for 10 seconds
 - o Aliquot a minimum of 1 mL of PreservCyt® specimen into a 13 mL round-bottom secondary tube
 - o Transfer tube to rack
 - Load sample rack and clotted tip racks into the sample supply module
 - Confirm samples have been accepted into the transfer module
- 4 Start run
- 5 Review and export results
- Remove sample tubes. If needed, cap any sample tubes meeting the minimum volume requirements for future use. Clean up instrument
 - Unload empty control cassettes
 - · Empty amplification plate drawer
 - Empty liquid waste
 - · Empty solid waste

Results

cobas° HPV automatically detects 14 high risk HPV genotypes (HPV-HR) and/or 12 high risk genotypes with individual typing of HPV16 and HPV18 simultaneously (HPV-GT).

Quality control and validity of results

- One **cobas*** Buffer Negative Control [(-) Ctrl] and one HPV Positive Control [HPV (+) C] are processed with each batch of a requested result type (HPV-HR or HPV-GT).
- In the cobas° 6800/8800 software and/or report, check for flags and their associated results to ensure batch validity.
- All flags are described in the **cobas**° 6800/8800 Systems User Guide.
- The batch is valid if no flags appear for all controls. If the batch is invalid, repeat testing of the entire batch.

Validation of results is performed automatically by the **cobas**° 6800/8800 software based on negative and positive control performance.

Interpretation of results

Display examples for **cobas**[®] HPV are shown in Figure 2 and Figure 3.

Figure 2 Example of cobas® HPV result display for the HPV-HR result request

Test	Sample ID	Valid	Flags	Sample type	Overall result	Target 1	Target 2	Target 3
HPV-HR	C161420284084194727902	Yes		HPV (+) C	Valid	Valid		
HPV-HR	C161420284090428825772	Yes		(-) Ctrl	Valid	Valid		
HPV-HR 400 ul	HPVHRinv_01	NA	Y40T	PreservCyt [®]	NA	Invalid		
HPV-HR 400 ul	HPVHRneg_01	NA		PreservCyt [®]	NA	HR HPV Negative		
HPV-HR 400 ul	HPVHRpos_01	NA		PreservCyt [®]	NA	HR HPV Positive		

Note: The Target 2 and Target 3 columns are reserved for HPV16 and HPV18 results with HPV-GT request, respectively.

Figure 3 Example of cobas® HPV result display for the HPV-GT result request

Test	Sample ID	Valid	Flags	Sample type	Overall result	Target 1	Target 2	Target 3
HPV-GT 400 ul	HPVGTpos_01	NA		PreservCyt [®]	NA	Other HR HPV Negative	HPV 16 Negative	HPV 18 Positive
HPV-GT 400 ul	HPVGTpos_02	NA		PreservCyt [®]	NA	Other HR HPV Negative	HPV 16 Positive	HPV 18 Positive
HPV-GT 400 ul	HPVGTpos_03	NA		PreservCyt [®]	NA	Other HR HPV Positive	HPV 16 Negative	HPV 18 Positive
HPV-GT 400 ul	HPVGTpos_04	NA		PreservCyt [®]	NA	Other HR HPV Positive	HPV 16 Negative	HPV 18 Negative
HPV-GT 400 ul	HPVGTpos_05	NA		PreservCyt [®]	NA	Other HR HPV Positive	HPV 16 Positive	HPV 18 Negative
HPV-GT 400 ul	HPVGTpos_06	NA		PreservCyt [®]	NA	Other HR HPV Positive	HPV 16 Positive	HPV 18 Positive
HPV-GT 400 ul	HPVGTpos_07	NA		PreservCyt [®]	NA	Other HR HPV Negative	HPV 16 Negative	HPV 18 Positive
HPV-GT 400 ul	HPVGTneg_01	NA		PreservCyt®	NA	Other HR HPV Negative	HPV 16 Negative	HPV 18 Negative
HPV-GT 400 ul	HPVGTpos_08	NA	C02H1	PreservCyt [®]	NA	Invalid	HPV 16 Positive	HPV 18 Positive
HPV-GT 400 ul	HPVGTpos_09	NA	C02H1	PreservCyt [®]	NA	Invalid	HPV 16 Positive	Invalid
HPV-GT 400 ul	C161420284090390657451	Yes		HPV (+) C	Valid	Valid	Valid	Valid
HPV-GT 400 ul	C161420284090419645071	Yes		(-) Ctrl	Valid	Valid	Valid	Valid

For a valid batch, check each individual sample for flags in the **cobas**° 6800/8800 software and/or report. The result interpretation should be as follows:

- A valid batch may include both valid and invalid sample results.
- The "Valid" and "Overall Result" columns are not applicable (NA) to sample results for **cobas*** HPV and are marked with "NA". Values reported in these columns **do not** impact the validity of results reported within individual target result columns.
- Reported target results for individual samples are valid unless indicated as "Invalid" within the individual target result column.
- Invalid results for one or more target combinations are possible with the HPV-GT result request and are reported out specifically for each channel. Refer to retesting instructions for the respective specimen type below.
- For invalid target results from PreservCyt® specimens, the original specimen should be re-tested no more than two times to obtain valid results. If the results are still invalid a new specimen should be obtained.

Results and their corresponding interpretation for detecting HR HPV only and Other HR HPV, HPV16 and HPV18 are shown in Table 10 and Table 11.

Table 10 cobas® HPV results and interpretation for the HPV-HR result request

Target 1	Target 2	Target 3	Interpretation
HR HPV Positive			Specimen is positive for the DNA of any one of, or combination of, the following high risk HPV types: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68.
HR HPV Negative	<blank></blank>	<blank></blank>	HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68 DNA were undetectable or below the pre-set threshold.
Invalid			The result for HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68 is invalid.

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Table 11 cobas® HPV results and interpretation for the HPV-GT result request

Target 1	Target 2	Target 3	Interpretation
Other HR HPV Positive	HPV 16 Positive.	HPV 18 Positive.	Specimen is positive for the DNA of any one of, or combination of the following high risk HPV types: 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68.
Other HR HPV Negative	HPV 16 Negative, or Invalid	HPV 18 Negative, or Invalid	HPV types 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68 were undetectable or below the pre-set threshold.
Invalid			The result for HPV types 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68 is invalid.
Other HR HPV Positive.	HPV 16 Positive	HPV 18 Positive.	Specimen is positive for HPV type 16 DNA.
Other HR HPV Negative,	HPV 16 Negative	HPV 18 Negative,	HPV type 16 DNA was undetectable or below the pre-set threshold.
or Invalid	Invalid	or Invalid	The result for HPV type 16 is invalid.
Other HR HPV Positive,	HPV 16 Positive,	HPV 18 Positive	Specimen is positive for HPV type 18 DNA.
Other HR HPV Negative,	HPV 16 Negative,	HPV 18 Negative	HPV type 18 DNA was undetectable or below the pre-set threshold.
or Invalid	or Invalid	Invalid	The result for HPV type 18 is invalid.

Procedural limitations

- cobas° HPV has been evaluated only for use in combination with the cobas° HPV Positive Control Kit, cobas° Buffer Negative Control Kit, cobas omni MGP Reagent, cobas omni Lysis Reagent, cobas omni Specimen Diluent, and cobas omni Wash Reagent for use on the cobas° 6800/8800 Systems.
- cobas® HPV has only been validated for use with cervical specimens collected by a clinician using an endocervical brush/spatula or a cervical broom and placed in the ThinPrep® Pap Test™ PreservCyt® Solution. Assay performance has not been validated for use with other collection media and/or specimen types. Use of other collection media and/or specimen types may lead to false positive, false negative or invalid results.
- Products containing carbomer(s), including vaginal lubricants, creams and gels may interfere with the test and should not be used during or prior to collecting cervical specimens. See Interference results (Table 20) for further details.
- Use of over-the-counter products Replens™, RepHresh™ Vaginal Gel and RepHresh™ Clean Balance™ Kit has been associated with false-negative results.
- Use of Metronidazole Vaginal Gel has been associated with false-negative results.
- cobas[®] HPV detects DNA of the high-risk types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68. This test does not detect DNA of HPV low-risk types (e.g. 6, 11, 42, 43, 44) since there is no clinical utility for testing of low-risk HPV types.¹³
- cobas* HPV is not recommended for evaluation of suspected sexual abuse and for other medico-legal indications.
- Detection of high-risk HPV is dependent on the number of copies present in the specimen and may be affected by specimen collection methods, patient factors, stage of infection and the presence of interfering substances.
- Prevalence of HPV infection in a population may affect performance. Positive predictive values decrease when testing populations with low prevalence or individuals with no risk of infection.

- Infection with HPV is not an indicator of cytologic HSIL or underlying high-grade CIN, nor does it imply that CIN2-3 or cancer will develop. Most women infected with one or more high-risk HPV types do not develop CIN2-3 or cancer.
- A negative high-risk HPV result does not exclude the possibility of future cytologic HSIL or underlying CIN2-3 or cancer.
- Human β-globin amplification and detection is included in **cobas**° HPV to differentiate HPV negative specimens from those that do not exhibit HPV signal due to insufficient cell mass in the specimen. All HPV negative specimens must have a valid β-globin signal within a pre-defined range to be identified as valid negatives.
- Reliable results depend on proper sample collection, storage and handling procedures.
- The addition of AmpErase enzyme into the **cobas*** HPV Master Mix enables selective amplification of target DNA; however, good laboratory practices and careful adherence to the procedures specified in this Instructions For Use are necessary to avoid contamination of reagents.
- Use of this product must be limited to personnel trained in the techniques of PCR and the use of the **cobas**° 6800/8800 Systems.
- Due to inherent differences between technologies, it is recommended that, prior to switching from one technology to the next, users perform method correlation studies in their laboratory to qualify technology differences. One hundred percent agreement between the results should not be expected due to aforementioned differences between technologies and normal variability of the tests.
- The effects of other potential variables such as vaginal discharge, use of tampons, douching, etc. and specimen collection variables have not been evaluated.
- Though rare, mutations within the highly conserved regions of the genomic DNA of Human papillomavirus covered by **cobas**° HPV's primers and/or probes may result in failure to detect the presence of the viral DNA.
- The presence of PCR inhibitors may cause false negative or invalid results.
- HPV negative results are not intended to prevent women from proceeding to colposcopy.
- Positive test results indicate the presence of any one or more of the high risk types, but since patients may be co-infected with low-risk types it does not rule out the presence of low-risk types in patients with mixed infections.
- Results of this test should only be interpreted in conjunction with information available from clinical evaluation of the patient and patient history.
- Use of tubes other than those recommended in Table 7 must be verified by user prior to implementation into **cobas*** HPV workflow in the laboratory.
- Use of barcodes other than those recommended in Table 7 must be verified by user prior to implementation into cobas° HPV workflow in the laboratory.
- Residual post-cytology specimens evaluated were processed on the ThinPrep® 2000 Processor.

Non-clinical performance evaluation

Key performance characteristics

Limit of Detection (LoD) at the clinical cutoff

The LoD at the clinical cutoff for HPV16 and HPV18 was assessed using SiHa and HeLa cell lines in the background of pooled HPV negative patient specimens. Cell lines were diluted to concentrations below, above and at the expected LoD levels. A minimum of 24 replicates were tested for each cell line level using 3 reagent lots with an equal number of runs performed on the **cobas*** 6800 and the **cobas*** 8800 Systems. The LoD was defined as the level of HPV DNA in the sample that has positive test results at least 95% of the time with concentration above the clinical cutoff.

The LoD for SiHa and HeLa was 16 cells/mL. Table 12 and Table 13 contain results from the reagent lot producing the most conservative (highest) LoD in the analysis for HPV16 and HPV18.

Table 12 Limit of Detection levels for HPV16 (SiHa Cell Line)

SiHa Concentration (cells/mL)	Number of Positive/Tested	% Positive	95% Confidence Interval
32	24 / 24	100%	86.2% - 100%
16	24 / 24	100%	86.2% - 100%
8	22 / 24	91.7%	74.2% - 97.7%

Table 13 Limit of Detection levels for HPV18 (HeLa Cell Line)

HeLa Concentration (cells/mL)	Number of Positive/Tested	% Positive	95% Confidence Interval
32	24 / 24	100%	86.2% - 100%
16	24 / 24	100%	86.2% - 100%
8	22 / 24	91.7%	74.2% - 97.7%

Inclusivity

Plasmids for high risk genotypes 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68 were tested in the background of pooled HPV negative patient specimens. All 12 of the high risk genotypes tested close to the LoD at the clinical cutoff were detected by the assay.

Precision

Within-laboratory precision was examined using a panel composed of either HPV cell lines or HPV positive clinical samples diluted into a pool of negative cervical specimen matrix. Additionally, a "zero concentration" HPV negative sample composed of HCT-15 cells in PreservCyt Solution was also included.

The precision panel was designed to include members with high negative, very low (< LoD), low (~ LoD) and moderate (> LoD) concentrations of HPV as well as an HPV negative. Testing was performed with three lots of **cobas*** HPV reagents on two instruments. There was an equal number of runs performed on the **cobas*** 6800 and the **cobas*** 8800 Systems over 12 days for a total of 24 runs for each panel member. A description of the precision panels and the observed hit rates are shown in Table 14.

All panel members exhibited the expected hit rates when tested using cobas° HPV.

Analysis of Ct values variability in positive panel members yielded overall CV (%) ranges from 4.32% to 6.19% for Other High Risk HPV (Table 15), 1.09% to 4.61% for HPV16 (Table 16) and 1.23% to 3.76% for HPV18 (Table 17).

Table 14 Summary of within laboratory precision

D I I I	Target	HPV	T 1	NI Taraka d	N D 't' -	III D. I	Hit Rate	95% CI
Panel Level	Source	Concentration	on Target Channel N Tested		N Positive	HIT Kate	LL	UL
Negative	N/A		Other HR HPV	72	0	0%	0%	5%
Negative	N/A	N/A	HPV16	72	0	0%	0%	5%
Negative	N/A		HPV18	72	0	0%	0%	5%
High Negative	Clinical sample		Other HR HPV	72	0	0%	0%	5%
High Negative	Clinical sample	N/A	HPV16	72	0	0%	0%	5%
High Negative	Clinical sample		HPV18	72	5	7%	3%	15%
< 1 x LoD	Clinical sample	N/A	Other HR HPV	72	30	42%	31%	53%
< 1 x LoD	Clinical sample	N/A	HPV16	71	33	47%	35%	58%
< 1 x LoD	Clinical sample	N/A	HPV18	72	49	68%	57%	78%
< 1 x LoD	SiHa cell line	4.8 cells/mL	HPV16	72	44	61%	50%	72%
< 1 x LoD	HeLa cell line	4.8 cells/mL	HPV18	72	49	68%	57%	78%
~ 1 x LoD	Clinical sample	N/A	Other HR HPV	72	72	100%	95%	100%
~ 1 x LoD	SiHa cell line	16 cells/mL	HPV16	72	72	100%	95%	100%
~ 1 x LoD	HeLa cell line	16 cells/mL	HPV18	72	72	100%	95%	100%
> 1 x LoD	Clinical sample	N/A	Other HR HPV	72	72	100%	95%	100%
> 1 x LoD	SiHa cell line	48 cells/mL	HPV16	72	72	100%	95%	100%
> 1 x LoD	HeLa cell line	48 cells/mL	HPV18	72	72	100%	95%	100%

CI= Confidence interval, LL= Lower limit, UL= Upper limit

Table 15 Overall mean, standard deviations and coefficients of variation (%) for cycle threshold - 12 Other High Risk HPV

	Between-Day		en-Day	Between- Instrument		Between- Operator		Between-Lot		Between-Run		Within-Run		Total		
Level	Hit Rate	Mean Ct	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV
< LoD ¹	41.7%	33.2	0	0	0	0	0	0	0	0	0.47	1.43	1.72	5.18	1.78	5.37
~ LoD ¹	100%	32.4	0	0	0	0	0.49	1.50	0.16	0.51	0	0	1.94	5.98	2.01	6.19
> LoD ¹	100%	30.7	0	0	0	0	0	0	0.27	0.88	0	0	1.30	4.23	1.33	4.32

¹ 12 Other HR HPV positive clinical sample diluted in pooled negative cervical specimen matrix

Table 16 Overall mean, standard deviations and coefficients of variation (%) for cycle threshold - HPV16

			Betwe	en-Day		reen- ıment	_	reen-	Betwe	en-Lot	Betwe	en-Run	Withi	n-Run	To	tal
Level	Hit Rate	Mean Ct	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV
< LoD ¹	46.5%	35.7	0.84	2.34	0.29	0.80	0.85	2.39	0	0	0	0	1.10	3.07	1.65	4.61
< LoD ²	61.1%	36.1	0.44	0.67	0	0	0.16	0.45	0.21	0.57	0	0	0.49	1.36	0.61	1.68
~ LoD ²	100%	35.0	0	0	0.02	0.06	0.02	0.07	0.38	1.09	0	0	0.45	1.28	0.59	1.69
> LoD ²	100%	34.0	0.03	0.09	0.04	0.12	0	0	0.27	0.78	0	0	0.25	0.74	0.37	1.09

¹ HPV16 positive clinical sample diluted in pooled negative cervical specimen matrix.

Table 17 Overall mean, standard deviations and coefficients of variation (%) for cycle threshold - HPV18

			Betwe	en-Day		reen- iment	_	rator	Betwe	en-Lot	Betwe	en-Run	Withi	n-Run	То	tal
Level	Hit Rate	Mean Ct	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV	SD	%CV
< LoD ¹	68.1%	35.9	0	0	0.55	1.52	0	0	0.18	0.51	0.17	0.49	1.21	3.37	1.35	3.76
< LoD ²	68.1%	35.3	0.19	0.54	0	0	0.02	0.06	0	0	0	0	0.97	2.75	0.99	2.80
~ LoD ²	100%	33.8	0	0	0	0	0	0	0.37	1.11	0	0	0.73	2.17	0.82	2.44
> LoD ²	100%	32.2	0	0	0	0	0	0	0.22	0.68	0.03	0.10	0.33	1.02	0.39	1.23

¹ HPV18 positive clinical sample diluted in pooled negative cervical specimen matrix.

² HPV16 cell line diluted in pooled negative cervical specimen matrix.

² HPV18 cell line diluted in pooled negative cervical specimen matrix.

Analytical specificity

A panel of bacteria, fungi and viruses, including those commonly found in the female urogenital tract, as well as several human papillomavirus types classified as low or undetermined risk were tested with $cobas^*$ HPV to assess analytical specificity. The organisms listed in Table 18 were spiked at concentrations of approximately 1 x 10⁶ units*/mL for bacteria and approximately 1 x 10⁵ units*/mL for viruses (except Adenovirus Type 40 which was tested at 2.82×10^4 units*/mL) into pools of HPV negative cervical specimens. Testing was performed with each potential interfering organism alone as well as with each organism mixed with HPV31 plasmid, SiHa (HPV16) and HeLa (HPV18) cell lines at approximately 3×10^{10} cobas* HPV. Results indicated that none of these organisms interfered with detection of Other High Risk HPV, HPV16 and HPV18 DNA or produced a false positive result in the HPV negative specimen.

* All bacteria were quantified as Colony Forming Units (CFU) except *Chlamydia trachomatis* which was quantified as Inclusion Forming Units (IFU). *Trichomonas vaginalis* was quantified as cells/mL. All viruses were quantified as units/mL as determined by TCID₅₀ Endpoint Dilution Assay except Epstein Barr virus which was in copies/mL.

Table 18 Microorganisms tested for analytical specificity

Adenovirus Type 40	Herpes Simplex Virus 2	HPV84
Bacteroides caccae	HPV6	HPV85
Bacteroides ureolyticus	HPV11	HPV89
Bifidobacterium adolescentis	HPV26	Klebsiella oxytoca
Bifidobacterium breve	HPV30	Klebsiella pneumoniae
Bifidobacterium longum	HPV34	Lactobacillus acidophillus
Candida albicans	HPV40	Neisseria gonorrhoeae
Chlamydia trachomatis	HPV42	Peptostreptococcus anaerobius
Clostridioides difficile	HPV53	Peptostreptococcus asaccharolyticus
Clostridium perfringens	HPV54	Proteus mirabilis
Corynebacterium genitalium	HPV55	Proteus penneri
Cytomegalovirus	HPV61	Proteus vulgaris
Enterobacter aerogenes	HPV62	Pseudomonas aeruginosa
Enterobacter cloacae	HPV64	Pseudomonas fluorescens
Enterococcus avium	HPV67	Pseudomonas putida
Enterococcus casseliflavus	HPV69	Staphylococcus aureus
Enterococcus faecalis	HPV70	Staphylococcus epidermidis
Enterococcus faecium	HPV71	Streptococcus agalactiae
Epstein Barr Virus	HPV72	Streptococcus pyogenes
Escherichia coli	HPV73	Treponema pallidum
Finegodia magna*	HPV81	Trichomonas vaginalis
Fusobacterium nucleatum	HPV82	
Herpes Simplex Virus 1	HPV83	

^{*}formerly Peptostreptococcus magnus

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Interference

The effects of endogenous and exogenous substances that may be present in cervical specimens were tested for potential interference. All testing for interference was performed with each potential interfering substance alone as well as with the substance mixed with SiHa (HPV16) and HeLa (HPV18) cell lines at approximately 3 x LoD of **cobas**° HPV in HPV negative samples.

Endogenous substances tested were cervical mucus, peripheral blood mononuclear cells and whole blood. Levels of endogenous substances tolerated by the assay are shown in Table 19. Exogenous substance testing included 18 over-the-counter (OTC) feminine hygiene and prescription products that are listed in Table 20. Of OTC feminine hygiene and prescription products tested, Metronidazole Vaginal Gel, Replens™, RepHresh™ Odor Eliminating Vaginal Gel and RepHresh™ Clean Balance™ Feminine Freshness Kit produced false negative results.

Potential interference from the presence of glacial acetic acid was also tested in pools of HPV negative and HPV positive cervical specimens in PreservCyt* Solution. Concentrations up to and including 5% (v/v) of glacial acetic acid were tolerated by the assay.

Table 19 Summary of endogenous substance concentrations that do not interfere with performance

Endogenous Substance	PreservCyt [®]
Mucus	Presence*
Peripheral Blood Mononuclear Cells (PBMCs as cells/mL)	1.00E+06
Whole Blood (% v/v)	10%

^{*} Presence refers to the amount of cervical mucus normally removed from the cervix prior to sampling.

Table 20 List of substances with concentrations that do not interfere with performance in cervical specimens

Product Name	Concentration
Clindamycin Phosphate Vaginal Cream	1.40 mg/mL
CVS Tioconazole 1 (Equate [™] tioconazole 1)	8.02 mg/mL
Equate [™] Vagicaine Anti-Itch Cream	5.87 mg/mL
Estrace® Cream	4.38 mg/mL
K-Y [®] Ultra Gel	6.59 mg/mL
Metronidazole Vaginal Gel [§]	*
Monistat® 3 Vaginal Antifungal Combination Pack	1.57 mg/mL
Monistat® Complete Care Itch Relief Cream	4.76 mg/mL
Gyne-Lotrimin® 7	3.13 mg/mL
Norforms [®] Suppositories	1.10 mg/mL
Premarin® Vaginal Cream	3.65 mg/mL
Replens [™] Long-Lasting Vaginal Moisturizer [§]	†
RepHresh [™] Odor Eliminating Vaginal Gel [§]	‡
RepHresh [™] Clean Balance [™] Feminine Freshness Kit [§]	‡
Summer's Eve [®] Feminine Deodorant Spray	0.90 mg/mL
VCF® - Vaginal Contraceptive Foam	1.42 mg/mL
Yeast Gard Advanced®	3.04 mg/mL
ZOVIRAX® (acyclovir) Cream 5%	10.37 mg/mL
Glacial acetic acid	5% (v/v)

^{*} Concentration of product that did not cause interference with test performance was 0.20 mg/mL.

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[†] Concentration of product that did not cause interference with test performance was 0.96 mg/mL.

[‡] Concentrations of product that did not interfere with test performance were not determined.

[§] Products containing carbomer(s) have been shown to cause interference.

Competitive inhibition

Competitive inhibition of HPV16 and HPV18 detection was assessed by testing samples containing low concentrations of HPV16 and HPV18 along with high concentration of non-targeted low risk HPV and targeted other high risk HPV. The HPV16 and HPV18 were spiked to concentrations close to $\sim 1 \times \text{LoD}$; each of the 25 low risk and 12 other high risk HPV tested were at a concentration were spiked to a concentration 1000-fold ($3\log_{10}$) higher than that of HPV16 and HPV18.

Results confirmed that **cobas**° HPV can detect low concentrations of HPV16 and HPV18 in the presence of any of the 25 non-targeted low risk and 12 other high risk HPV at a concentration that is 1000-fold (3log₁₀) higher in concentration.

Cross contamination

Studies were performed to evaluate potential cross contamination on the **cobas** $^{\circ}$ 6800/8800 Systems using **cobas** $^{\circ}$ HPV. In this performance study the sample to sample cross-contamination rate of **cobas** $^{\circ}$ HPV has been determined to be 0% (0/288, 95% CI= 0.00% – 1.27%) when alternating very high positive sample representing more than 95% of the positives in the intended use population with negative samples over multiple runs. Run to run cross-contamination has been determined to be 0% (0/187, 95% CI= 0.00% – 1.95%).

Clinical performance evaluation

Expected results

A multicenter, prospective study (IMPACT trial, <u>IM</u>proved <u>Primary screening And Colposcopy Triage</u>) was conducted to evaluate the performance of **cobas*** HPV performed on the **cobas*** 6800/8800 Systems (Roche Molecular Systems, Inc., hereafter referred to as **cobas*** 6800/8800 HPV test) as a triage test to stratify women with ASC-US Pap cytology results for colposcopy, as an adjunctive test to cervical cytology to guide management decisions in women with NILM Pap cytology, and as a first-line primary test for cervical cancer screening.

In total, 35,263 women 25-65 years were enrolled from September 2017 to October 2018 at 32 clinical sites in the United States. A total of 34,914 women met study eligibility criteria. One woman was excluded due to insufficient sample volume for HPV testing (eligible n=34,913). The percent of invalid **cobas**° 6800/8800 HPV test results was 0.04% (13/34,913) with 95% CI: 0.02% to 0.06%.

Table 21 shows HPV positivity by the **cobas*** 6800/8800 HPV test by testing site and study population. The overall HPV positivity was 35.20% in the ASC-US (25-65 years) population, 10.16% in the NILM (30-65 years) population and 15.08% in the Primary Screening (25-65 years) population.

Table 21	HPV positivity by the cobas®	6800/8800 HPV test by testing sites and study population
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Testing Site	esting Site Evaluable ASC-US Evaluable Site Population P (25-65 Years) (30		Evaluable Primary Screening Population (25-65 Years)
1	38.28% (116/303)	8.83% (454/5,139)	13.29% (910/6,846)
2	39.53% (204/516)	10.47% (636/6,074)	15.10% (1,229/8,138)
3	32.40% (335/1,034)	10.78% (902/8,364)	16.85% (2,051/12,171)
4	34.53% (144/417)	10.10% (580/5,745)	13.85% (1,060/7,652)
Overall	35.20% (799/2,270)	10.16% (2,572/25,322)	15.08% (5,250/34,807)

Table 22 shows HPV positivity of the **cobas*** 6800/8800 HPV test by age and study population. HPV positivity decreased with age in each study population. In the ASC-US population, HPV positivity decreased from 52.15% in 25-29 year olds to 38.20% in 30-39 year olds and remained ~24% in women 40-65 years old. In the NILM population, HPV positivity was 12.67% in 30-39 year old women and remained about 8% in 40-65 year old women. In the primary screening population, HPV positivity decreased from 24.01% in 25-29 year olds to 16.44% in 30-39 year olds and remained relatively constant at ~10-11% in 40-65 year old women.

Table 22 HPV positivity by the cobas® 6800/8800 HPV test by age and study population

	cobas® 6800/8800 HPV Test Result					
Age Group (Years)	ASC-US Population (25-65 Years)	NILM Population (30-65 Years)	Primary Screening Population (25-65 Years)			
25-29	52.15% (267/512)	Not Applicable	24.01% (1,568/6,530)			
30-39	38.20% (288/754)	12.67% (1,328/10,482)	16.44% (1,944/11,826)			
40-49	24.00% (129/538)	8.54% (632/7,397)	11.05% (914/8,271)			
50-65	24.68% (115/466)	8.22% (612/7,443)	10.07% (824/8,180)			
Overall	35.20% (799/2,270)	10.16% (2,572/25,322)	15.08% (5,250/34,807)			

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cobas° 6800/8800 HPV test results stratified by age for the ASC-US, NILM, and primary screening populations are presented in Table 23. In all populations, 12 Other HR HPV positive results were more frequent than HPV16 and HPV18 positive results in general and within age groups.

Table 23 cobas® 6800/8800 HPV test results by age group for the evaluable populations

		cobas [®] 6800/8	800 HPV Test Result		
Age Group (Years)	HPV16 Positive n (%)	HPV18 Positive n (%)	12 Other HR HPV Positive n (%)	HPV Negative n (%)	Total N
	ASC-	-US Population (25-	65 Years)	1	
Overall	6.34% (144/2,270)	2.78% (63/2,270)	26.08% (592/2,270)	64.80% (1,471/2,270)	2,270
25-29	8.01% (41/512)	2.34% (12/512)	41.80% (214/512)	47.85% (245/512)	512
30-39	7.43% (56/754)	3.18% (24/754)	27.59% (208/754)	61.80% (466/754)	754
40-49	5.02% (27/538)	2.42% (13/538)	16.54% (89/538)	76.02% (409/538)	538
50-65	4.29% (20/466)	3.00% (14/466)	17.38% (81/466)	75.32% (351/466)	466
	NIL	M Population (30-6	5 Years)		
Overall	2.20% (556/25,322)	1.21% (306/25,322)	6.75% (1,710/25,322)	89.84% (22750/25,322)	25,32
30-39	2.57% (269/10,482)	1.43% (150/10,482)	8.67% (909/10,482)	87.33% (9,154/10,482)	10,48
40-49	2.16% (160/7,397)	1.04% (77/7,397)	5.34% (395/7,397)	91.46% (6,765/7,397)	7,397
50-65	1.71% (127/7,443)	1.06% (79/7,443)	5.45% (406/7,443)	91.78% (6,831/7,443)	7,443
	Primary So	creening Population	(25-65 Years)		
Overall	3.06% (1,064/34,807)	1.42% (493/34,807)	10.61% (3,693/34,807)	84.92% (29557/34,807)	34,80
25-29	3.61% (236/6,530)	1.23% (80/6,530)	19.17% (1,252/6,530)	75.99% (4,962/6,530)	6,530
30-39	3.57% 1.70% 11.17% 83.56% (422/11,826) (201/11,826) (1,321/11,826) (9,882/11,826)		11,82		
40-49	2.88% (238/8,271)	1.27% (105/8,271)	6.90% (571/8,271)	88.95% (7,357/8,271)	8,27
50-65	2.05% (168/8,180)	1.31% (107/8,180)	6.71% (549/8,180)	89.93% (7,356/8,180)	8,180

Clinical performance

The study enrolled 35,263 women aged 25–65 years undergoing routine cervical cancer screening in the US from September 2017 to October 2018 at 32 clinical sites in the Baseline Phase. A total of 34,914 women were eligible to participate in the study.

Following written informed consent, demographic information and gynecologic histories were obtained. All women had one cervical sample collected using a brush/spatula for approximately half of the subjects and a broom-type device for the other half. Cervical samples were collected for HPV testing and ThinPrep® Pap Test™ liquid based cytology (LBC). Two HPV tests were used: the FDA-approved HPV Test and **cobas®** 6800/8800 HPV test, performed according to manufacturer's instructions. HPV testing was performed on pre-aliquoted samples in secondary vials prior to cytology processing at four testing laboratories. LBC testing was conducted at the same four laboratories. Cytology samples were classified according to the criteria of the 2001 Bethesda System. Pap cytology, FDA approved HPV Test, and **cobas®** 6800/8800 HPV test results were used to inform referral to colposcopy.

To determine the clinical study endpoint, a subset of non-pregnant women identified at the enrollment visit was selected to undergo colposcopy and biopsy/endocervical curettage (ECC). The subset included women aged 25-65 years with ≥ASC-US cytology and women aged 25-65 years with positive **cobas**° HPV Test results (positive by the FDA-approved HPV Test and/or **cobas**° 6800/8800 HPV test). In addition, 59 women with unsatisfactory Pap cytology and HPV-negative results (negative by both the FDA-approved HPV Test and **cobas**° 6800/8800 HPV test), and a randomly selected subset of subjects with NILM Pap cytology and HPV-negative results (negative by both the FDA-approved HPV Test and **cobas**° 6800/8800 HPV test) were referred to colposcopy (approximately 1:50). In order to avoid bias, study participants and colposcopists were blinded to all HPV test and cytology results until after the colposcopy was completed.

Colposcopy was conducted according to a standardized protocol following the principles recommended by the American Society for Colposcopy and Cervical Pathology (ASCCP) as follows: biopsies were obtained on all visible lesions; endocervical curettage was performed in all patients in whom the squamocolumnar junction was not visualized and a single random cervical biopsy was obtained if no lesions were visible. All biopsies were examined by a Central Pathology Review (CPR) process consisting of three expert pathologists, and discordant results adjudicated according to a predefined protocol. The slides that were prepared from the biopsies were stained using conventional hematoxylin and eosin (H&E) staining, and H&E with p16 IHC assay (CINtec Histology, Ventana Medical Systems, Inc.). The expert pathologist first evaluated H&E-stained slides to establish the CPR_{H&E} reference diagnosis, then evaluated both H&E- and p16 histology-stained slides for that case to establish the CPR_{H&E+p16} reference diagnosis. Additionally, CPR results were derived using results from CPR_{H&E} and CPR_{H&E+p16} diagnoses, where diagnosis was based on the H&E-stained slides with adjunctive interpretation of the p16-stained slides only when a case met LAST (Lower Anogenital Squamous Terminology) criteria (excluding ASC-US/HPV16+ as a LAST criterion).

Clinical performance of the **cobas*** 6800/8800 HPV test is presented using interpretation of H&E-stained slides with adjunctive use of p16-stained slides in accordance with the 2012 Lower Anogenital Squamous Terminology Standardization Project for HPV-Associated Lesions (LAST)³⁰ excluding ASC-US/HPV16+ as a LAST criterion (CPR_{H&E+p16 per LAST}) at the clinical endpoints \geq CIN2 and \geq CIN3.

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ASC-US (25-65 years) population - performance evaluation

In IMPACT, all women 25-65 years old with \geq ASC-US Pap cytology, regardless of HPV results, were invited to undergo colposcopy. The clinical performance of **cobas*** 6800/8800 HPV test was measured against histology results of \geq CIN2 and \geq CIN3 by CPR.

Eligible women in IMPACT who had an ASC-US Pap cytology result and valid **cobas** $^{\circ}$ 6800/8800 HPV test results were considered evaluable for the analyses of the ASC-US objectives. Of the 34,807 evaluable women in the overall population, 2,270 women had an ASC-US Pap cytology (6.5%) and valid **cobas** $^{\circ}$ 6800/8800 HPV test result. The results of the **cobas** $^{\circ}$ 6800/8800 HPV test reported as HPV Positive or HPV Negative by CPR histologic diagnosis are presented in Table 24. Of the 1,814 women with ASC-US and valid CPR diagnosis, 124 women were diagnosed with \geq CIN2 (prevalence of 6.8%) and 36 women were diagnosed with \geq CIN3 (prevalence of 2.0%).

Table 24 cobas® 6800/8800 HPV test results and central pathology review diagnoses in the evaluable ASC-US population (25-65 years)

cobas® 6800/8800	Central Pathology Review Diagnoses					Total	
HPV Test Result	Undetermined ¹	Normal ²	CIN1	CIN2	≥CIN3	Total	
HPV Positive	42	440	84	75	32	673	
HPV Negative	66	1,101	65	13	4	1,249	
Total	108	1,541	149	88	36	1,922	

¹Undetermined includes: biopsy sample inadequate for analysis, subject/colposcopist unblinded to HPV or Pap cytology result at colposcopy visit or biopsy sample taken out of window.

The performance of **cobas**° 6800/8800 HPV test and the FDA-approved HPV Test in detecting high grade cervical disease (≥CIN2 and ≥CIN3) is presented in Table 25. The sensitivity for detecting ≥CIN2 was 86.29% (95% CI: 79.14, 91.26) for the **cobas**° 6800/8800 HPV test, similar to the performance of the FDA-approved HPV Test (86.18%, 95% CI: 78.98, 91.19). The specificity for detecting ≥CIN2 was 68.99% (95% CI: 66.75, 71.15) for the **cobas**° 6800/8800 HPV test and 69.47% (95% CI: 67.23, 71.62) for the FDA-approved HPV Test.

The sensitivity for detecting ≥CIN3 was 88.89% (95% CI: 74.69, 95.59) for the **cobas*** 6800/8800 HPV test, and 86.11% (95% CI: 71.34, 93.92) for the FDA-approved HPV Test. The specificity for detecting ≥CIN3 was 66.31% (95% CI: 64.08, 68.47) for the **cobas*** 6800/8800 HPV test, and 66.74% (95% CI: 64.52, 68.89) for the FDA-approved HPV Test.

²Normal includes: Negative or normal histology and atypical squamous cells or glandular changes indefinite for neoplasia.

Table 25 Performance of the cobas® 6800/8800 HPV test and the FDA-approved HPV Test in the evaluable ASC-US population (25-65 years)

Performance	Prevalence (95% C	CIN2 I)=6.84% (124/1814) 6, 8.09)	≥CIN3 Prevalence (95% CI)=1.98% (36/1814) (1.44, 2.74)		
Parameters	cobas® 6800/8800 HPV Test	FDA-approved HPV Test	cobas® 6800/8800 HPV Test	FDA-approved HPV Test	
Sensitivity (%) (95% CI)	86.29 (107/124) (79.14, 91.26)	86.18 (106/123) (78.98, 91.19)	88.89 (32/36) (74.69, 95.59)	86.11 (31/36) (71.34, 93.92)	
Specificity (%) (95% CI)	68.99 (1166/1690) (66.75, 71.15)	69.47 (1174/1690) (67.23, 71.62)	66.31 (1179/1778) (64.08, 68.47)	66.74 (1186/1777) (64.52, 68.89)	
PPV (%) (95% CI)	16.96 (107/631) (15.60, 18.41)	17.04 (106/622) (15.66, 18.52)	5.07 (32/631) (4.47, 5.75)	4.98 (31/622) (4.33, 5.73)	
NPV (%) (95% CI)	98.56 (1166/1183) (97.78, 99.07)	98.57 (1174/1191) (97.80, 99.08)	99.66 (1,179/1,183) (99.15, 99.87)	99.58 (1,186/1,191) (99.06, 99.81)	
PLR (95% CI)	2.78 (107/124) / (524/1690) (2.52, 3.08)	2.82 (106/123) / (516/1690) (2.55, 3.12)	2.64 (32/36) / (599/1778) (2.31, 3.01)	2.59 (31/36) / (591/1777) (2.24, 3.00)	
NLR (95% CI)	0.20 (17/124) / (1166/1690) (0.13, 0.31)	0.20 (17/123) / (1174/1690) (0.13, 0.31)	0.17 (4/36) / (1179/1778) (0.07, 0.42)	0.21 (5/36) / (1186/1777) (0.09, 0.47)	

PPV=Positive predictive value; NPV=Negative predictive value; PLR=Positive likelihood ratio; NLR=Negative likelihood ratio

The performance of the **cobas** $^{\circ}$ 6800/8800 HPV test and the FDA-approved HPV Tests for detecting high grade cervical disease (\geq CIN2 and \geq CIN3) stratified by age group is presented in Table 26. The sensitivity for detecting \geq CIN2 ranged from 80.77% to 89.36% for the **cobas** $^{\circ}$ 6800/8800 HPV test and from 84.62% to 89.36% for the FDA-approved HPV Test; the specificity ranged from 51.35% to 77.72% for the **cobas** $^{\circ}$ 6800/8800 HPV test and from 52.25% to 78.43% for the FDA-approved HPV Test.

The sensitivity for ≥CIN3 of both the **cobas**° 6800/8800 HPV test and the FDA-approved HPV Test ranged from 66.67% to 93.33%; the specificity ranged from 47.95% to 76.22% for the **cobas**° 6800/8800 HPV test and from 48.77% to 76.79% for the FDA-approved HPV Test.

Table 26 Performance of the cobas[®] 6800/8800 HPV test and the FDA-approved HPV Test in detecting ≥CIN2 and ≥CIN3 in the evaluable ASC-US population, stratified by age group

Statistic	cobas [®] 6800/8800 HPV Test	FDA-approved HPV Test	cobas® 6800/8800 HPV Test	FDA-approved HPV Test	cobas [®] 6800/8800 HPV Test	FDA-approved HPV Test
	25	-29 Years	30-	39 Years	40-0	65 Years
	•		≥CIN2		·	
Prevalence (%) (95% CI)		37 (47/380) 43, 16.06)		(51/591) 2, 11.17)		(26/843) 1, 4.48)
Sensitivity (%) (95% CI)	89.36 (42/47) (77.41, 95.37)	89.36 (42/47) (77.41, 95.37)	86.27 (44/51) (74.28, 93.19)	84.00 (42/50) (71.49, 91.66)	80.77 (21/26) (62.12, 91.49)	84.62 (22/26) (66.47, 93.85)
Specificity (%) (95% CI)	51.35 (171/333) (46.00, 56.67)	52.25 (174/333) (46.89, 57.56)	66.67 (360/540) (62.59, 70.51)	66.54 (360/541) (62.46, 70.39)	77.72 (635/817) (74.74, 80.44)	78.43 (640/816) (75.48, 81.12)
PPV (%) (95% CI)	20.59 (42/204) (18.27, 23.11)	20.90 (42/201) (18.53, 23.47)	19.64 (44/224) (17.21, 22.32)	18.83 (42/223) (16.38, 21.56)	10.34 (21/203) (8.42, 12.65)	11.11 (22/198) (9.20, 13.36)
NPV (%) (95% CI)	97.16 (171/176) (93.69, 98.75)	97.21 (174/179) (93.79, 98.77)	98.09 (360/367) (96.27, 99.03)	97.83 (360/368) (95.96, 98.84)	99.22 (635/640) (98.30, 99.64)	99.38 (640/644) (98.48, 99.75)
			≥CIN3			
Prevalence (%) (95% CI)		5 (15/380) 2.41, 6.41)		(15/591) 54, 4.15)		(6/843) 3, 1.54)
Sensitivity (%) (95% CI)	93.33 (14/15) (70.18, 98.81)	93.33 (14/15) (70.18, 98.81)	93.33 (14/15) (70.18, 98.81)	86.67 (13/15) (62.12, 96.26)	66.67 (4/6) (30.00, 90.32)	66.67 (4/6) (30.00, 90.32)
Specificity (%) (95% CI)	47.95 (175/365) (42.87, 53.07)	48.77 (178/365) (43.68, 53.88)	63.54 (366/576) (59.53, 67.37)	63.54 (366/576) (59.53, 67.37)	76.22 (638/837) (73.22, 78.98))	76.79 (642/836) (73.81, 79.53)
PPV (%) (95% CI)	6.86 (14/204) (5.87, 8.01)	6.97 (14/201) (5.95, 8.14)	6.25 (14/224) (5.31, 7.34)	5.83 (13/223) (4.71, 7.20)	1.97 (4/203) (1.11, 3.46))	2.02 (4/198) (1.14, 3.55)
NPV (%) 95% CI (%)	99.43 (175/176) (96.33, 99.91)	99.44 (178/179) (96.39, 99.92)	99.73 (366/367) (98.22, 99.96)	99.46 (366/368) (98.05, 99.85)	99.69 (638/640) (99.04, 99.90)	99.69 (642/644) (99.04, 99.90)

PPV=Positive predictive value; NPV=Negative predictive value.

Table 27 presents CPR diagnosis by all possible **cobas**° 6800/8800 HPV test results in the evaluable ASC-US women who completed colposcopy.

Table 27 All possible cobas® 6800/8800 HPV test results and central pathology review diagnoses in the evaluable ASC-US population (25-65 years)

	Central Pathology Review Diagnoses					
cobas [®] 6800/8800 HPV Test Result (12 Other HR HPV; HPV16; HPV18)	Undetermined ¹	Normal ²	CIN1	CIN2	≥CIN3	Total
12 Other HR HPV Negative;						
HPV16 Negative;	66	1,101	65	13	4	1,249
HPV18 Negative						
12 Other HR HPV Negative;						
HPV16 Negative;	1	18	5	2	1	27
HPV18 Positive						
12 Other HR HPV Negative;						
HPV16 Positive;	3	31	4	10	9	57
HPV18 Negative						
12 Other HR HPV Negative;						
HPV16 Positive;	0	1	0	1	0	2
HPV18 Positive						
12 Other HR HPV Positive;						
HPV16 Negative;	31	344	67	49	13	504
HPV18 Negative						
12 Other HR HPV Positive;						
HPV16 Negative;	2	15	3	3	0	23
HPV18 Positive						
12 Other HR HPV Positive;						
HPV16 Positive;	5	28	4	10	9	56
HPV18 Negative						
12 Other HR HPV Positive;						
HPV16 Positive;	0	3	0	0	0	3
HPV18 Positive						
12 Other HR HPV Positive;						
Invalid;	0	0	1	0	0	1
Invalid						
Overall	108	1,541	149	88	36	1,922

¹Undetermined includes: biopsy sample inadequate for analysis, subject/colposcopist unblinded to HPV or Pap cytology result at colposcopy visit or biopsy sample taken out of window.

Likelihood ratios (LRs) for the **cobas**° 6800/8800 HPV test are presented in Table 28 for the ASC-US (25-65 years) population.

Likelihood ratio of 14 HR HPV positive results associated with \geq CIN2 and \geq CIN3 was 2.78 and 2.64, respectively, indicating an overall increased probability of disease in women with HPV positive results.

For \geq CIN2, the LR of HPV16 positive and/or HPV18 positive was 5.48, indicating that an HPV16 positive and/or an HPV18 positive result is \sim 5.5 times more likely to occur in a subject with \geq CIN2 than in a subject without. The LR of a negative **cobas**° 6800/8800 HPV test result was 0.20, indicating that a negative result was 5 times more likely to occur in a subject without <CIN2 than in a subject with \geq CIN2.

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²Normal includes: Negative or normal histology and atypical squamous cells or glandular changes indefinite for neoplasia.

For ≥CIN3, LR of HPV16 positive and/or HPV18 positive was 6.80, and the LR of an HPV negative result was 0.17.

Table 28 Likelihood ratios of disease (≥CIN2 and ≥CIN3) by the cobas® 6800/8800 HPV test results in the evaluable ASC-US population (25-65 years)

cobas [®] 6800/8800	Likelihood Ratio (95% CI)		
HPV Test Result	≥CIN2 vs <cin2< th=""><th>≥CIN3 vs <cin3< th=""></cin3<></th></cin2<>	≥CIN3 vs <cin3< th=""></cin3<>	
HPV Positive	2.78 (2.52, 3.08)	2.64 (2.31, 3.01)	
HPV16 Positive	7.49 (5.30, 10.58)	9.66 (6.59, 14.17)	
HPV18 Positive	1.99 (0.86, 4.61)	1.07 (0.15, 7.57)	
HPV16/18 Positive	5.48 (4.08, 7.35)	6.80 (4.80, 9.63)	
12 Other HR HPV Positive	2.05 (1.69, 2.49)	1.39 (0.90, 2.17)	
HPV Negative	0.20 (0.13, 0.31)	0.17 (0.07, 0.42)	

ASC-US (25-65 years) population - absolute and relative risk estimates

The absolute risk of disease among women with positive HPV results was 16.96% and 5.07% for \geq CIN2 and \geq CIN3, respectively (Table 29). For both \geq CIN2 and \geq CIN3, the risk of disease was highest for women with HPV positive results, HPV16 and/or HPV18 positive results, and 12 Other HR HPV positive results and lowest for an HPV negative result.

The absolute risk of disease (\geq CIN2 and \geq CIN3) by **cobas*** 6800/8800 HPV test results stratified by age group in the evaluable ASC-US population is presented in Table 30. For all age groups, absolute risks were higher for women with any HPV positive results, and lowest for an HPV negative result.

Table 29 Absolute risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable ASC-US population (25-65 years)

cobas [®] 6800/8800 HPV Test Result	Absolute Risk % (n/N) (95% Cl)			
	≥CIN2	≥CIN3		
LIDV Desitive	16.96 (107/631)	5.07 (32/631)		
HPV Positive	(14.23, 20.08)	(3.61, 7.07)		
LIDV40440 D. W.	28.66 (45/157)	12.10 (19/157)		
HPV16/18 Positive	(22.17, 36.18)	(7.89, 18.13)		
LIDVAC Decitive	35.45 (39/110)	16.36 (18/110)		
HPV16 Positive	(27.14, 44.75)	(10.61, 24.39)		
LIDV10 Decitive	12.77 (6/47)	2.13 (1/47)		
HPV18 Positive	(5.98, 25.17)	(0.38, 11.11)		
10 Odbar LID LIDV Danishira	13.08 (62/474)	2.74 (13/474)		
12 Other HR HPV Positive	(10.34, 16.41)	(1.61, 4.64)		
LIDV No motive	1.44 (17/1183)	0.34 (4/1183)		
HPV Negative	(0.90, 2.29)	(0.13, 0.87)		

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Table 30 Absolute risk of disease (≥CIN2 and ≥CIN3) by HPV Genotype from the cobas® 6800/8800 HPV test in the evaluable ASC-US population (25-65 years), stratified by age group

cobas® 6800/8800 HPV Test Result	Absolute Risk,% (95% CI)			
	≥CIN2	≥CIN3		
	25–29 Years			
HPV Positive	20.59 (15.61, 26.66)	6.86 (4.13, 11.19)		
HPV16/18 Positive	34.21 (21.21, 50.11)	15.79 (7.44, 30.42)		
HPV16 Positive	38.71 (23.73, 56.18)	19.35 (9.19, 36.28)		
HPV18 Positive	14.29 (2.57, 51.31)	0.00 (0.00, 35.43)		
12 Other HR HPV Positive	17.47 (12.45, 23.96)	4.82 (2.46, 9.22)		
HPV Negative	2.84 (1.22, 6.48)	0.57 (0.10, 3.15)		
	30-39 Years			
HPV Positive	19.64 (14.97, 25.34)	6.25 (3.76, 10.22)		
HPV16/18 Positive	40.00 (28.57, 52.63)	18.33 (10.56, 29.92)		
HPV16 Positive	46.51 (32.51, 61.08)	23.26 (13.15, 37.74)		
HPV18 Positive	23.53 (9.55, 47.26)	5.88 (1.05, 26.98)		
12 Other HR HPV Positive	12.20 (8.03, 18.09)	1.83 (0.62, 5.24)		
HPV Negative	1.91 (0.93, 3.88)	0.27 (0.05, 1.53)		
	40-65 Years			
HPV Positive	10.34 (6.87, 15.30)	1.97 (0.77, 4.96)		
HPV16/18 Positive	13.56 (7.03, 24.54)	3.39 (0.93, 11.54)		
HPV16 Positive	19.44 (9.75, 35.03)	5.56 (1.54, 18.14)		
HPV18 Positive	4.35 (0.77, 20.99)	0.00 (0.00, 14.31)		
12 Other HR HPV Positive	9.03 (5.35, 14.83)	1.39 (0.38, 4.92)		
HPV Negative	0.78 (0.33, 1.82)	0.31 (0.09, 1.13)		

The relative risk (RR) of disease (\geq CIN2 and \geq CIN3) by **cobas**° 6800/8800 HPV test results in the evaluable ASC-US population is presented in Table 31.

The RRs of \geq CIN2 and \geq CIN3 for women with positive vs. negative **cobas**° 6800/8800 HPV test results were 11.78 (95% CI: 7.14, 19.50) and 14.91 (95% CI: 5.33, 42.22), respectively, indicating that women with a positive HPV result were \sim 12 times more likely to have \geq CIN2 and \sim 15 times more likely to have \geq CIN3 than women with a HPV negative test result.

Similarly, women who were HPV16 and/or HPV18 positive were significantly more likely to have \geq CIN2 than women with (i) a positive result for 12 Other HR HPV types (2.19), or (ii) a HPV negative result (19.90). Women with a positive 12 Other HR HPV result were significantly more likely to have \geq CIN2 than women with a HPV negative result (9.08). Similar results were observed for \geq CIN3 histology i.e. women with HPV positive results were more likely to have \geq CIN3 compared to HPV negative results (8.06).

Table 31 Relative risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable ASC-US population (25-65 years)

cobas® 6800/8800 HPV Test Result	Relative	Risk (95% CI)
	≥CIN2	≥CIN3
HPV Positive vs HPV Negative	11.78 (7.14, 19.50)	14.91 (5.33, 42.22)
HPV16/18 Positive vs HPV Negative	19.90 (11.71, 33.97)	35.59 (12.33, 103.86)
HPV16/18 Positive vs 12 Other HR HPV Positive	2.19 (1.56, 3.07)	4.42 (2.23, 8.73)
12 Other HR HPV Positive vs HPV Negative	9.08 (5.38, 15.40)	8.06 (2.66, 24.75)

The relative risk of disease (\geq CIN2 and \geq CIN3) by **cobas*** 6800/8800 HPV test results stratified by age group in the evaluable ASC-US population is presented in Table 32 . For all age groups, similar patterns of increased risks associated with any HPV positive results were observed as those presented for the overall population in Table 32.

Table 32 Relative risk of disease (≥CIN2 and ≥CIN3) by HPV Genotype from the cobas® 6800/8800 HPV test in the evaluable ASC-US population (25-65 years), stratified by age group

cobas® 6800/8800 HPV Test Result	Relative R	isk (95% CI)
	≥CIN2	≥CIN3
25-	29 Years	
HPV Positive vs HPV Negative	7.25 (2.93, 17.92)	12.04 (1.60, 90.94)
HPV16/18 Positive vs HPV Negative	12.05 (4.56, 31.77)	27.70 (3.44, 224.18)
HPV16/18 Positive vs 12 Other HR HPV Positive	1.96 (1.13, 3.40)	3.28 (1.21, 8.89)
12 Other HR HPV Positive vs HPV Negative	6.15 (2.44, 15.51)	8.46 (1.07, 67.09)
30-	39 Years	
HPV Positive vs HPV Negative	10.28 (4.72, 22.47)	23.15 (3.04, 173.25)
HPV16/18 Positive vs HPV Negative	20.94 (9.46, 46.51)	67.89 (8.85, 511.73)
HPV16/18 Positive vs 12 Other HR HPV Positive	3.28 (1.96, 5.49)	10.02 (2.89, 34.70)
12 Other HR HPV Positive vs HPV Negative	6.39 (2.76, 14.82)	6.78 (0.70, 64.06)
40-	65 Years	
HPV Positive vs HPV Negative	13.26 (5.06, 34.67)	6.35 (1.16, 34.17)
HPV16/18 Positive vs HPV Negative	17.38 (5.86, 51.37)	10.94 (1.56, 75.62)
HPV16/18 Positive vs 12 Other HR HPV Positive	1.50 (0.66, 3.43)	2.44 (0.35, 16.92)
12 Other HR HPV Positive vs HPV Negative	11.58 (4.19, 31.90)	4.48 (0.63, 31.29)

Use of the cobas® 6800/8800 HPV test in ASC-US triage of women 21-24 years

In order to evaluate the performance of the **cobas**° 6800/8800 HPV test in 21- to 24-year old women with ASC-US Pap cytology, 140 refrigerated residual cervical samples collected in PreservCyt° were identified from participants in the ATHENA trial who were 21-24 years old and diagnosed with ASC-US Pap cytology. Samples were tested in 2018 using both **cobas**° 6800/8800 HPV test and the FDA-approved HPV Test (Table 33). One sample had invalid result by the FDA-approved HPV test and the number of evaluable samples was 139. Agreements for HPV16, HPV18, and 12 Other HR HPV are shown in Table 33, respectively.

Table 33 Cross-tabulation of cobas® 6800/8800 HPV test results and the FDA-approved HPV Test results using residual ATHENA samples

cobas [®] 6800/8800	FDA-approved HPV Test Result				
HPV Test Result	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	33	0	2	0	35
HPV18 Positive	0	8	3	1	12
12 Other HR HPV Positive	0	0	81	0	81
HPV Negative	0	0	2	9	11
Total	33	8	88	10	139
Genotype Specific PPA	100.00% (33/33)	100.00% (8/8)	92.05% (81/88)		
(95% CI)	(89.57, 100.00)	(67.55, 100.00)	(84.48, 96.09)		
14 HR HPV		PPA: 98.45% (127/129	9)	NPA: 90.00% (9/10)	
Percent Agreement (95% CI)		(94.52, 99. 57)		(59.50, 98.21)	

PPA: Positive percent agreement; NPA: Negative percent agreement.

Note: HPV16 positive implies HPV16 positive, HPV18 positive or negative and 12 Other HR HPV positive or negative.

HPV18 positive implies HPV16 negative, HPV18 Positive and 12 Other HR HPV positive or negative.

12 Other HR HPV positive implies HPV16 negative, HPV18 negative and 12 Other HR HPV positive.

NILM population (30-65 years) - performance characteristics

Of the 34,807 evaluable women in the overall population, 25,322 had NILM Pap cytology and were 30-65 years of age. Among these, a total of 3,335 (13.17%) were selected or randomized for colposcopy for histologic diagnosis. Of those identified for colposcopy, 2,805 completed the procedure, and thereof 2,632 had a valid CPR and cobas[®] 6800/8800 HPV test result. Table 34 summarizes the distribution of verified and unverified CPR diagnoses by the cobas[®] 6800/8800 HPV test results. A total of 151 subjects were diagnosed with ≥CIN2 by CPR including 54 cases with ≥CIN3.

Table 34 cobas® 6800/8800 HPV test results and central pathology review diagnoses in the evaluable NILM population (30-65 years)

cobas® 6800/8800	Centra	Patholog	y Review Di	agnoses	5. 2. 3	
HPV Test Result	Normal ¹	CIN1	CIN2	≥CIN3	Unknown Disease Status ²	Total
HPV Positive	1,797	100	94	54	527	2,572
HPV Negative	571	13	3	0	22,163	22,750
Total	2,368	113	97	54	22,690	25,322

¹Normal includes: Negative or normal histology, and atypical squamous cells or glandular changes indefinite for neoplasia.

Unadjusted and adjusted performance characteristics for the NILM (30-65) population are shown in Table 35. The

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²Unknown disease status includes: biopsy sample inadequate for analysis or subject/colposcopist unblinded to HPV or Pap cytology result at colposcopy visit or biopsy sample taken out of window or subjects not selected for colposcopy.

unadjusted estimates of sensitivity and specificity for detection of \geq CIN2 were 98.01% (95% CI: 94.32, 99.32) and 23.54% (95% CI: 21.91, 25.25) respectively; for detection of \geq CIN3 estimates were 100% (95% CI: 93.36, 100) and 22.77% (95% CI: 21.19, 24.43). The verification bias adjusted sensitivities for \geq CIN2 and \geq CIN3 were 72.51% and 100%, respectively; adjusted specificities were 90.48% and 90.08%, respectively. The adjusted estimates of PPV for \geq CIN2 and \geq CIN3 were 7.20% and 2.61%, respectively; NPVs were 99.69% and 100%, respectively.

The adjusted estimates of prevalence for ≥CIN2 and ≥CIN3 were 1.01% and 0.27%, respectively.

Table 35 Performance of the cobas® 6800/8800 HPV test in the evaluable NILM population (30-65 years)

	Central Pathology Review Diagnoses					
Performance Parameters	≥(CIN2	≥(CIN3		
raiameters	Unadjusted	Adjusted	Unadjusted	Adjusted		
Sensitivity (%)	98.01 (148/151)	72.51	100.00 (54/54)	100.00		
(95% CI)	(94.32, 99.32)	(44.28, 100.00)	(93.36, 100.00)	(93.36, 100.00)		
Specificity (%)	23.54 (584/2481)	90.48	22.77 (587/2578)	90.08		
(95% CI)	(21.91, 25.25)	(90.10, 90.85)	(21.19, 24.43)	(89.72, 90.45)		
PPV (%)	7.24 (148/2045)	7.20	2.64 (54/2045)	2.61		
(95% CI)	(6.19, 8.44)	(6.10, 8.34)	(2.03, 3.43)	(1.93, 3.30)		
NPV (%)	99.49 (584/587)	99.69	100.00 (587/587)	100.00		
(95% CI)	(98.51, 99.83)	(99.02, 100.00)	(99.35, 100.00)	(99.35, 100.00)		
Prevalence (%)	5.74 (151/2632)	1.01	2.05 (54/2632)	0.27		
(95% CI)	(4.91, 6.69)	(0.65, 1.62)	(1.58, 2.67)	(0.19, 0.34)		

PPV=Positive predictive value; NPV=Negative predictive value.

NILM (30-65 years) population - likelihood ratios

Table 36 shows the **cobas**° 6800/8800 HPV test results in the evaluable NILM (30-65) population by CPR diagnoses.

Table 36 All possible cobas® 6800/8800 HPV test results and central pathology review diagnoses in the evaluable NILM population (30-65 years)

	C	entral Patholo	gy Review D	iagnoses		
cobas [®] 6800/8800 HPV Test Result (12 Other HR HPV; HPV16; HPV18)	Undetermined ¹	Normal ²	CIN1	CIN2	≥CIN3	Total
12 Other HR HPV Negative; HPV16 Negative; HPV18 Negative	22,163	571	13	3	0	22,750
12 Other HR HPV Negative; HPV16 Negative; HPV18 Positive	45	189	5	5	5	249
12 Other HR HPV Negative; HPV16 Positive; HPV18 Negative	98	304	13	6	20	441
12 Other HR HPV Negative; HPV16 Positive; HPV18 Positive	3	9	1	0	0	13
12 Other HR HPV Positive; HPV16 Negative; HPV18 Negative	351	1,193	73	70	22	1,709
12 Other HR HPV Positive; HPV16 Negative; HPV18 Positive	10	39	4	3	1	57
12 Other HR HPV Positive; HPV16 Positive; HPV18 Negative	19	56	4	10	6	95
12 Other HR HPV Positive; HPV16 Positive; HPV18 Positive	0	7	0	0	0	7
12 Other HR HPV Positive ; Invalid; Invalid	1	0	0	0	0	1
Overall	22,690	2,368	113	97	54	25,322

¹Undetermined includes: biopsy sample inadequate for analysis, or subject/colposcopist unblinded to HPV or Pap cytology result at colposcopy visit or biopsy sample taken out of window and subjects not identified for colposcopy

Table 37 presents the likelihood ratio (LR) of disease (\geq CIN2 and \geq CIN3) in women 30-65 years old with NILM Pap cytology by the **cobas*** 6800/8800 HPV test results. Adjusted likelihood ratios of HR HPV positive results associated with \geq CIN2 and \geq CIN3 were 7.62 and 10.08, respectively, indicating an overall increased probability of disease associated with HPV positive result.

For \geq CIN3, positive HPV16 results had the highest positive LR of 23.78 (adjusted), indicating that a positive HPV16 result is approximately 23 times more likely to come from those with \geq CIN3 than without. There were no cases of \geq CIN3 observed among women with a negative **cobas*** 6800/8800 HPV test result. Similar patterns of high positive likelihood associated with HPV positive results and low negative likelihoods associated with HPV negative results were observed for \geq CIN2.

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²Normal includes: Negative or normal histology and atypical squamous cells or glandular changes indefinite for neoplasia.

Table 37 Likelihood ratios of disease (≥CIN2 and ≥CIN3) by the cobas® 6800/8800 HPV test results in the evaluable NILM population (30-65 years)

	Likelihood Ratio (95% CI)						
cobas [®] 6800/8800 HPV Test Result	≥CIN2 v	s <cin2< th=""><th colspan="3">≥CIN3 vs <cin3< th=""></cin3<></th></cin2<>	≥CIN3 vs <cin3< th=""></cin3<>				
	Unadjusted	Adjusted	Unadjusted	Adjusted			
HPV Positive	1.28 (1.24 ,1.32)	7.62 (4.63, 10.78)	1.29 (1.27, 1.32)	10.08 (9.64, 10.50)			
HPV16 Positive	1.75 (1.33, 2.30)	10.14 (5.34, 16.58)	3.03 (2.26, 4.05)	23.78 (13.94, 30.90)			
HPV18 Positive	0.97 (0.58, 1.62)	5.78 (2.19, 11.65)	1.17 (0.54, 2.51)	8.82 (1.44, 16.05)			
HPV16/18 Positive	1.46 (1.17, 1.81)	8.55 (5.07, 12.89)	2.33 (1.85, 2.94)	18.34 (12.49, 22.22)			
12 Other HR HPV Positive	1.19 (1.04, 1.36)	7.15 (4.37, 11.05)	0.79 (0.57,1.09)	6.05 (4.02, 8.95)			
HPV Negative	0.08 (0.03, 0.24)	0.30 (0.00, 0.60)	0.00* (0.00, 0.29)	0.00* (0.00, 0.29)			

^{*}No ≥CIN3 cases observed among women with negative cobas® 6800/8800 HPV test results

NILM (30-65 years) population - absolute risk and relative risk estimates

The adjusted absolute risk (AR) of disease (\geq CIN2 and \geq CIN3) in the NILM 30-65 population by the **cobas**° 6800/8800 HPV test results are presented in Table 38. The adjusted AR of \geq CIN2 was 7.19% among women with a positive HPV test result; highest in women with a positive HPV16 result (9.35%), followed by women with a positive 12 Other HR HPV result (6.78%), and women with an HPV18 positive result (5.56%). The adjusted AR of \geq CIN3 was 2.60% among women with a positive HPV test result; highest in women with a positive HPV16 result (5.94%), followed by women with a positive HPV18 result (2.29%), and women with a positive 12 Other HR HPV result (1.58%).

The risks of \geq CIN2 and \geq CIN3 were low among women with HPV negative results (adjusted ARs: 0.31%, and 0.15%, respectively). Age stratified absolute risks are presented in Table 39 and Table 40.

Table 38 Absolute risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable NILM population (30-65 years)

		Absolute Risk % (95% CI)						
cobas [®] 6800/8800 HPV Test Result	≥CIN	2	≥CINS	3				
	Unadjusted	Adjusted	Unadjusted	Adjusted				
LIDV Dopitivo	7.24 (148/2045)	7.19	2.64 (54/2045)	2.60				
HPV Positive	(6.19, 8.44)	(6.10, 8.33)	(2.03, 3.43)	(1.93, 3.31)				
LIDVA O Description	9.63 (42/436)	9.35	5.96 (26/436)	5.94				
HPV16 Positive	(7.21, 12.77)	(6.66, 12.09)	(4.10, 8.59)	(3.60, 8.19)				
LIDV/10 Desiking	5.58 (14/251)	5.56	2.39 (6/251)	2.29				
HPV18 Positive	(3.35, 9.14)	(2.68, 8.40)	(1.10, 5.12)	(0.63, 4.38)				
LIDV /10 /10 Dec. '1'	8.15 (56/687)	8.00	4.66 (32/687)	4.64				
HPV16/18 Positive	(6.33, 10.44)	(5.99, 10.06)	(3.32, 6.50)	(3.11, 6.29)				
10 Other LID LIDV Decition	6.77 (92/1358)	6.78	1.62 (22/1358)	1.58				
12 Other HR HPV Positive	(5.56, 8.24)	(5.54, 8.21)	(1.07, 2.44)	(0.96, 2.30)				
LIDV/ Nonetice	0.51 (3/587)	0.31	0.09 (0.5*/587)	0.15				
HPV Negative	(0.17, 1.49)	(0.00, 0.98)	(0.01, 0.81)	(0.00; 0.18)				

^{*}No ≥CIN3 cases observed among women with negative **cobas**® 6800/8800 HPV test results, 0.5 case was used in order to estimate risk.

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Table 39 Absolute risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable NILM population (30-39 years)

		Absolute Risk % (95% CI)						
cobas [®] 6800/8800 HPV Test Result	≥CIN	l2	≥CIN3					
	Unadjusted	Adjusted	Unadjusted	Adjusted				
LIDV Danisir.	9.44 (98/1038)	9.26	3.47 (36/1038)	3.39				
HPV Positive	(7.81, 11.37)	(7.64, 10.93)	(2.52, 4.76)	(2.31,4.44)				
HPV16 Positive	16.27 (34/209)	15.61	9.57 (20/109)	9.29				
HPV 16 POSITIVE	(11.88, 21.87)	(11.21,20.38)	(6.28, 14.32)	(5.55,13.25)				
LIDV/10 Desixing	5.74 (7/122)	5.33	1.64 (2/122)	1.33				
HPV18 Positive	(2.81, 11.37)	(2.03,9.83)	(0.45, 5.78)	(0.00,4.21)				
LIDV/10/10 Desitive	12.39 (41/331)	11.93	6.65 (22/331)	6.44				
HPV16/18 Positive	(9.26, 16.37)	(8.95,15.35)	(4.43, 9.86)	(4.00,9.10)				
10 Other IID IID// Desitive	8.06 (57/707)	8.03	1.98 (14/707)	1.98				
12 Other HR HPV Positive	(6.27, 10.30)	(6.11,9.95)	(1.18, 3.30)	(1.04,2.96)				
LIDV/ No mation	0.42 (1/236)	0.74	0.21(0.5/236)	0.37				
HPV Negative	(0.07, 2.36)	(0.00,2.55)	(0.02, 2.00)	(0.00,0.43)				

Table 40 Absolute risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable NILM population (40-65 years)

	Absolute Risk % (95% CI)					
cobas [®] 6800/8800 HPV Test Result	≥CIN	2	≥CIN3			
	Unadjusted	Adjusted	Unadjusted	Adjusted		
LIDV Dopitivo	4.97 (50/1007)	4.98	1.79 (18/1007)	1.85		
HPV Positive	(3.79, 6.49)	(3.78,6.43)	(1.13, 2.81)	(0.97,2.65)		
LIDV/10 Deciking	3.52 (8/227)	3.48	2.64 (6/227)	2.79		
HPV16 Positive	(1.80, 6.80)	(1.44,6.11)	(1.22, 5.65)	(0.66,5.03)		
LIDV/10 Donitivo	5.43 (7/129)	5.77	3.10 (4/129)	3.21		
HPV18 Positive	(2.65, 10.78)	(1.95,9.90)	(1.21, 7.70)	(0.56,6.62)		
LIDV10/10 Docitivo	4.21 (15/356)	4.29	2.81 (10/356)	2.93		
HPV16/18 Positive	(2.57, 6.83)	(2.23,6.47)	(1.53, 5.09)	(1.12,4.59)		
12 Other LID LIDV Desitive	5.38 (35/651)	5.37	1.23 (8/651)	1.25		
12 Other HR HPV Positive	(3.89, 7.39)	(3.83,7.08)	(0.62, 2.41)	(0.50,2.11)		
LIDV/ Nogotivo	0.57 (2/351)	0.02	0.14 (0.5/351)	0.01		
HPV Negative	(0.16, 2.05)	(0.00,0.05)	(0.01, 1.35)	(0.00,0.01)		

The relative risk (RR) of \geq CIN2 and \geq CIN3 is shown in Table 41. Relative risk of \geq CIN2 and \geq CIN3 for women with HPV positive result compared with HPV negative result was 14.21 and 29.33, respectively. Women with HPV16 and/or HPV18 positive results had the highest relative risk compared to women with HPV negative results (\geq CIN2: 15.98 and \geq CIN3: 51.78).

Table 41 Relative risk of disease (≥CIN2 and ≥CIN3) by HPV genotype from the cobas® 6800/8800 HPV test in the evaluable NILM population (30-65 years)

	Central Pathology Review Diagnoses			
cobas® 6800/8800 HPV Test Result	≥CIN2	≥CIN3		
HPV Positive vs. Negative	14.20 (4.53, 44.25)	29.33 (1.92, 501.27)		
HPV16/18 Positive vs. Negative	15.98 (5.02, 50.69)	51.78 (3.35, 891.43)		
12 Other HR HPV Positive vs. Negative	13.27 (4.21, 41.69)	18.00 (1.15, 313.24)		
HPV16/18 Positive vs. 12 Other HR HPV positive	1.20 (0.87, 1.66)	2.88 (1.68, 4.91)		

Note: Unadjusted estimates shown

Agreement between the cobas $^{\rm @}$ 6800/8800 HPV test results and the FDA-approved HPV Test results for women 25-65 years

The analytical agreement of the **cobas**° 6800/8800 HPV test was also evaluated by estimating the percent agreements, along with 95% confidence intervals (CIs), for different results of the FDA-approved HPV Test in prequot samples (Table 42 and Table 43). Genotype specific percent agreements were: PPA for HPV16 positive was 97.07% (95% CI: 95.64, 98.04); PPA for HPV18 positive was 97.21% (95% CI: 94.60, 98.658), PPA for 12 Other HR HPV positive was 85.96% (95% CI: 84.8, 87.00) and NPA for HPV negative was 97.73% (95% CI: 97.55, 97.89).

Table 42 Cross-tabulation of the cobas® 6800/8800 HPV test results and the FDA-approved HPV test results

	FDA-approved HPV Test Result				
cobas® 6800/8800 HPV Test Result	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	762	6	46	250	1,064
HPV18 Positive	2	279	47	165	493
12 Other HR HPV Positive	13	1	3,409	260	3,683
HPV Negative	8	1	464	29,011	29,484
Total	785	287	3,966	29,686	34,724

Note: HPV16 positive implies HPV16 positive, HPV18 positive or negative and 12 Other HR HPV positive or negative.

HPV18 positive implies HPV16 negative, HPV18 Positive and 12 Other HR HPV positive or negative.

12 Other HR HPV positive implies HPV16 negative, HPV18 negative and 12 Other HR HPV positive.

Table 43 Agreement between the cobas® 6800/8800 HPV test results and the FDA Approved HPV test results for the detection of HPV genotypes

HPV Genotypes	Positive Percent Agreement % (n/N; 95% CI)	Negative Percent Agreement % (n/N; 95% CI)
HPV16 Positive	97.07% (762/785; 95.64%, 98.04%)	99.11% (33,637/33,939; 99.00%, 99.20%)
HPV18 Positive	97.21% (279/287; 94.60%, 98.58%)	99.38% (34,223/34,437; 99.29%, 99.46%)
12 Other HR HPV Positive	85.96% (3,409/3,966; 84.84%, 87.00%)	99.11% (30,484/30,758; 99.00%, 99.21%)
HPV Positive	90.61% (4,565/5,038; 89.77%, 91.39%)	97.73% (29,011/29,686; 97.55%, 97.89%)

Note: HPV16 positive implies HPV16 positive, HPV18 positive or negative and 12 Other HR HPV positive or negative.

HPV18 positive implies HPV16 negative, HPV18 Positive and 12 Other HR HPV positive or negative.

12 Other HR HPV positive implies HPV16 negative, HPV18 negative and 12 Other HR HPV positive.

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Agreement with a composite comparator for the ASC-US (25-65 years) and the NILM (30-65 years) populations

The agreement of the **cobas**° 6800/8800 HPV test was evaluated by comparing results from the test with a composite comparator composed of HPV DNA sequencing and an FDA-approved HR HPV DNA test.

Representative cervical samples were selected from 2 subsets of women from the IMPACT study: women \geq 25 years who had ASC-US Pap cytology results (n=590), and women 30-65 years with NILM Pap cytology results (n=3,167).

The analytical agreement of the **cobas**° 6800/8800 HPV test results were compared with the composite comparator for the detection of 14 HR HPV genotypes and the positive percent agreement (PPA), negative percent agreement (NPA) and 95% confidence intervals (CIs) were calculated. The composite comparator for the detection of 14 HR HPV was indeterminate if results were discordant between HPV DNA sequencing result and the FDA-approved HR HPV DNA test result. All results including the indeterminate results for the compososite comparator for the detection of 14 HR HPV are presented in Table 44.

Table 44 Agreement between the cobas® 6800/8800 HPV test results and the composite comparator for the detection of 14 HR HPV

	cobas [®] 6800/8800	HPV	Composite Compa	arator		Agreement (%)	
Population	HPV Test Result	Positive Negative Indeterminate		Total	(95% CI)		
ASC-US	Positive	420	0	12	432	PPA: 98.36% (420/427) (96.66%, 99.20%)	
≥25 Years	Negative	7	134	17	158	NPA: 100.0% (134/134) (97.21%, 100.00%)	
	Total	427	134	29	590	-	
	Positive	1153	31	79	1263	PPA: 90.57% (1153/1273) (88.84%, 92.06%)	
NILM ≥30 Years	Negative	120	1635	149	1904	NPA: 98.14% (1635/1666) (97.37%, 98.69%)	
	Total	1273	1666	228	3167	-	

PPA=positive percent agreement, NPA=negative percent agreement.

The analytical agreement of the **cobas*** 6800/8800 HPV test results were compared with the composite comparator for HPV genotyping and the corresponding percent agreements (PA) along with 95% confidence intervals (CIs) were calculated: PA for HPV16 positive, PA for HPV18 positive, PA for 12 Other HR HPV positive and PA for HPV negative. The composite comparator for HPV genotyping was indeterminate if results were discordant between HPV DNA sequencing result and the FDA-approved HR HPV DNA test result. All results including the indeterminate results for the compososite comparator for HPV genotyping are presented in Table 45 and Table 46 for ASC-US 25-65 years and NILM 30-65 years, respectively.

Table 45 Agreement between the cobas® 6800/8800 HPV test results and composite comparator in the ASC-US population (25-65 years)

	Composite Comparator for HPV Genotyping								
cobas® 6800/8800 HPV Test Result	FDA-approved= HPV16 Positive, DNA Sequencing= HPV16 Positive	FDA-approved= HPV18 Positive, DNA Sequencing= HPV18 Positive	FDA-approved= 12 Other HR HPV Positive, DNA Sequencing= 12 Other HR HPV Positive	FDA-approved= HPV Negative DNA Sequencing= HPV Negative	Indeterminate	Total			
HPV16 Positive	68	0	2	0	6	76			
HPV18 Positive	0	21	2	0	8	31			
12 Other HR HPV Positive	0	0	317	0	8	325			
HPV Negative	0	0	7	134	17	158			
Total	68	21	328	134	39	590			
Percent Agreement (95% CI)	100.0% (68/68) (94.65%, 100.0%)	100.0% (21/21) (84.54%, 100.0%)	96.65% (317/328) (94.10%, 98.12%)	100.0% (134/134) (97.21%, 100.0%)					

Note: Indeterminate includes results where FDA approved and DNA Sequencing results are discordant.

Table 46 Agreement between the cobas® 6800/8800 HPV test results and composite comparator in the NILM population (30-65 years)

	Composite Comparator for HPV Genotyping								
cobas® 6800/8800 HPV Test Result	FDA-approved= HPV16 Positive, DNA Sequencing= HPV 16 Positive	FDA-approved= HPV18 Positive, DNA Sequencing= HPV18 Positive	FDA-approved=12 Other HR HPV Positive, DNA Sequencing=12 Other HR HPV Positive	FDA-approved= HPV Negative, DNA Sequencing= HPV Negative	Indeterminate	Total			
HPV16 Positive	171	1	13	19	30	234			
HPV18 Positive	0	74	11	5	15	105			
12 Other HR HPV Positive	0	0	853	7	64	924			
HPV Negative	1	1	113	1635	154	1904			
Total	172	76	990	1666	263	3167			
Percent Agreement (95% CI)	99.42% (171/172) (96.78%, 99.90%)	97.37% (74/76) (90.90%, 99.28%)	86.16% (853/990) (83.87%, 88.17%)	98.14% (1635/1666) (97.37%, 98.69%)					

Note: Indeterminate includes results where FDA approved and DNA Sequencing results are discordant.

Primary screening population (25-65 years) - performance evaluation

Among the 35,263 women enrolled in the study, 34,914 met study eligibility criteria. One woman was excluded due to insufficient sample volume for **cobas**° 6800/8800 HPV testing and from the 34,913 eligible women, a total of 34,807 were evaluable for the analyses of the primary screening population. To be evaluable, women must have been eligible for study at enrollment and have a valid **cobas**° 6800/8800 HPV test result. The percent of invalid **cobas**° 6800/8800 HPV test results was 0.04% (13/34,913) with 95% CI: 0.02% to 0.06%.

The median age of the evaluable women in the primary screening population was 39 years with 18.8% women in the age group 25-29 years and 34% in the age group 30-39 years; the remaining 47.2% women were 40-65 years old.

Approximately 12% self-reported that they had received the HPV vaccine, 91.5% reported Pap cytology screening within

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the previous 5 years, and 8.4% reported having colposcopy procedure within 5 years prior to study enrollment. Among the evaluable women 42.4% reported having an HPV screening test in the previous 5 years and among them 12.7% reported having a positive HPV result.

A total of 6,826 women proceeded to colposcopy and of these 6,776 subjects completed colposcopy. Thereof, biopsy samples for 3 subjects were lost/misplaced during transport. Diagnosis of \geq CIN2 was observed in 595 of 6,773 (8.8%) women who went to colposcopy and had valid CPR results at colposcopy.

The number of women with colposcopy results for each combination of the **cobas*** 6800/8800 HPV test and Pap cytology results is shown in Table 47. A correction for verification bias was applied due to the different rate of colposcopy in each category. Disease status was imputed for those women without histology results from the women who went to colposcopy based on their HPV result (from both **cobas*** 6800/8800 HPV test and FDA-approved HPV Test), Pap cytology, and age.

Table 47 Number of subjects with adjudicated histology and pap cytology and cobas® 6800/8800 HPV test results in the evaluable primary screening population (25-65 years)

cobas [®] 6800/8800	Number of	Pap Cytology				
HPV Test Result	Subjects	NILM	ASC-US	>ASC-US	Unsatisfactory	Total
	Total	1,061	207	270	19	1,557
HPV16/18 Positive	With adjudicated colposcopy	884	168	230	12	-
12 Other HR HPV	Total	2,518	592	545	38	3,693
Positive	With adjudicated colposcopy	2,099	506	447	26	-
	Total	27,326	1,471	323	437	29,557
HPV Negative	With adjudicated colposcopy	804	1,250	285	65	-
Total	-	30,905	2,270	1,138	494	34,807

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Screening algorithms

The use of the **cobas**° 6800/8800 HPV test as a first line screening method was evaluated by comparing the Primary Screening algorithm (Figure 4) with the Cytology Alone algorithm (Figure 5).

Figure 4 Primary screening algorithm

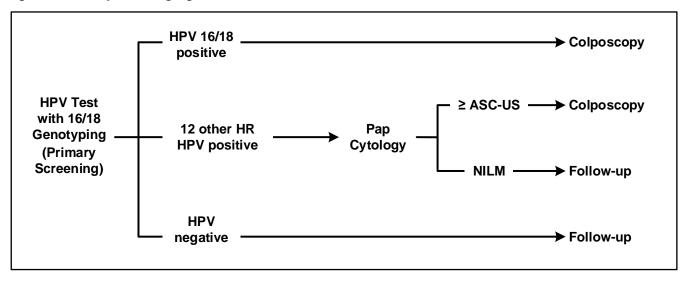
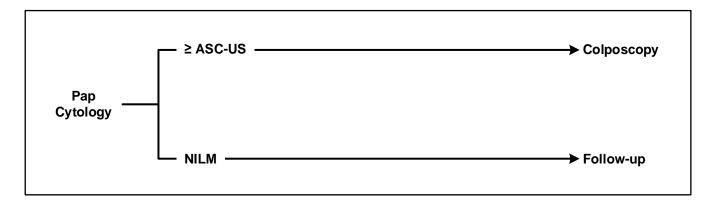


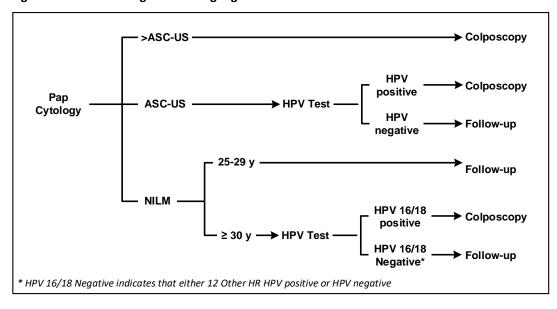
Figure 5 Cytology alone algorithm



The peformance of the **cobas**° 6800/8800 HPV test as a first line screening method was also evaluated by comparing the Primary Screening algorithm with the ASC-US Triage/Co-testing algorithm which represents the screening strategy endorsed by the 2012 American Society of Colposcopy and Cervical Pathology guidelines. According to these guidelines, women 25-29 years of age with >ASC-US Pap cytology results or ASC-US and high risk HPV positive results are referred to colposcopy, as are women ≥30 years with >ASC-US Pap cytology results or ASC-US and high risk HPV positive results, as well as those with NILM Pap cytology and HPV16 and/or HPV18 positive results (Figure 6).

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Figure 6 ASC-US triage/co-testing algorithm



Performance of the Primary Screening algorithm and the Cytology Alone algorithm was evaluated by estimating the sensitivity, specificity, PLR, NLR, prevalence, PPV, and NPV in the identification of high-grade cervical disease (\geq CIN2 and \geq CIN3); results of the comparison are presented in Table 48.

The performance of the Primary Screening algorithm was significantly better than the Cytology Alone algorithm for both ≥CIN2 and ≥CIN3 clinical endpoints in that the Primary Screening algorithm had significantly higher sensitivity, specificity, PPV, NPV and PLR, and also significantly lower NLR compared with the Cytology Alone algorithm. Also, the Primary Screening algorithm improved disease detection (13.82% increase in ≥CIN3 sensitivity) and required 2.05% fewer colposcopy referrals compared to the Cytology Alone algorithm.

Table 48 Adjusted performance of the primary screening and cytology alone algorithms in the evaluable primary screening population (25-65 years)

	Prevalence	≥CIN2 e (95% CI)=2.34 (2.03, 2.83)	≥CIN3 Prevalence (95% CI)=0.87 (0.77, 0.98)			
Performance Parameters	Primary Screening Algorithm	Cytology Alone Algorithm	Difference	Primary Screening Algorithm	Cytology Alone Algorithm	Difference	
Sensitivity (%)	62.41	56.39	6.02	79.93	66.12	13.82	
(95% CI)	(52.39, 70.24)	(47.27, 64.21)	(2.85, 9.21)	(74.36, 84.80)	(59.71, 72.37)	(8.42, 19.46)	
Specificity (%)	93.57	91.32	2.24	92.90	90.71	2.19	
(95% CI)	(93.31, 93.85)	(91.04, 91.63)	(1.93, 2.54)	(92.62, 93.19)	(90.41, 91.00)	(1.89, 2.50)	
PPV (%)	18.86	13.47	5.39	9.02	5.90	3.12	
(95% CI)	(17.15, 20.55)	(12.13, 14.85)	(4.35, 6.23)	(7.90, 10.20)	(5.06, 6.81)	(2.51, 3.81)	
NPV (%)	99.05	98.87	0.18	99.81	99.67	0.14	
(95% CI)	(98.57, 99.32)	(98.39, 99.16)	(0.09, 0.25)	(99.75, 99.86)	(99.60, 99.74)	(0.09, 0.19)	
PLR	9.70	6.50	3.20	11.25	7.11	4.14	
(95% CI)	(8.09, 11.11)	(5.38, 7.47)	(2.51, 3.86)	(10.35, 12.12)	(6.42, 7.83)	(3.37, 4.92)	
NLR	0.40	0.48	-0.08	0.22	0.37	-0.16	
(95% CI)	(0.32, 0.51)	(0.39, 0.58)	(-0.11, -0.04)	(0.16, 0.28)	(0.31, 0.44)	(-0.22, -0.10)	
Colposcopy Referral (%)	7.74	9.79	-2.05	7.74	9.79	-2.05	
(95% CI)	(7.45, 8.02)	(9.48, 10.09)	(-2.35, -1.74)	(7.45, 8.02)	(9.48, 10.09)	(-2.35, -1.74)	

PPV=Positive predictive value; NPV=Negative predictive value; PLR=Positive likelihood ratio; NLR= Negative likelihood ratio.

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The comparisons of the Primary Screening algorithm to the ASC-US Triage/Co-testing algorithm are shown in Table 49. For both \geq CIN2 and \geq CIN3 clinical endpoints, Primary Screening algorithm had significantly higher specificity, PPV and PLR compared with the ASC-US Triage/Co-testing algorithm. Also, the Primary Screening algorithm required 0.35% fewer colposcopy referrals compared to the ASC-US Triage/Co-testing algorithm. For detecting \geq CIN2 and \geq CIN3, sensitivity, NPV, and NLRs were similar between the two algorithms.

Table 49 Adjusted performance of the primary screening algorithm and the ASC-US triage/co-testing algorithm in the evaluable primary screening population (25-65 years)

	Prevalence	≥CIN2 (95% CI)=2.34%	(2.03, 2.83)	≥CIN3 Prevalence (95% CI)=0.87% (0.77, 0.98)			
Performance Parameters	Primary Screening Algorithm	ASC-US Triage /Co-testing Algorithm	Difference	Primary Screening Algorithm	ASC-US Triage /Co-testing Algorithm	Difference	
Sensitivity (%)	62.41	62.53	-0.12	79.93	77.63	2.30	
(95% CI)	(52.39, 70.24)	(52.65, 70.78)	(-2.18, 1.55)	(74.36, 84.80)	(71.90, 82.69)	(-0.96, 5.95)	
Specificity (%)	93.57	93.21	0.36	92.90	92.52	0.37	
(95% CI)	(93.31, 93.85)	(92.96, 93.50)	(0.21, 0.48)	(92.62, 93.19)	(92.25, 92.80)	(0.24, 0.50)	
PPV (%)	18.86	18.08	0.78	9.02	8.38	0.64	
(95% CI)	(17.15, 20.55)	(16.51, 19.83)	(0.14, 1.26)	(7.90, 10.20)	(7.34, 9.57)	(0.27, 1.05)	
NPV (%)	99.05	99.05	0.00	99.81	99.79	0.02	
(95% CI)	(98.57, 99.32)	(98.57, 99.33)	(-0.05, 0.04)	(99.75, 99.86)	(99.72, 99.84)	(-0.01, 0.06)	
PLR	9.70	9.21	0.49	11.25	10.38	0.87	
(95% CI)	(8.09, 11.11)	(7.71, 10.56)	(0.09, 0.81)	(10.35, 12.12)	(9.52, 11.20)	(0.36, 1.40)	
NLR	0.40	0.40	-0.00	0.22	0.24	-0.03	
(95% CI)	(0.32, 0.51)	(0.31, 0.51)	(-0.02, 0.02)	(0.16, 0.28)	(0.19, 0.30)	(-0.07, 0.01)	
Colposcopy Referral (%)	7.74	8.09	-0.35	7.74	8.09	-0.35	
(95% CI)	(7.45, 8.02)	(7.80, 8.38)	(-0.48, -0.22)	(7.45, 8.02)	(7.80, 8.38)	(-0.48, -0.22)	

PPV=Positive predictive value; NPV=Negative predictive value; PLR=Positive likelihood ratio; NLR= Negative likelihood ratio.

Table 50 presents the performance of the Primary Screening algorithm, Cytology Alone algorithm and the ASC-US Triage/Co-testing algorithm stratified by age groups for detection of ≥CIN3.

Table 50 Adjusted performance of the primary screening algorithm, cytology alone algorithm and ASC-US triage/co-testing algorithm for detection of ≥CIN3, stratified by age group

Performance Parameters	Sensitivity (%) (95% CI)	Specificity (%) (95% CI)	PPV (%) (95% CI)	NPV (%) (95% CI)	PLR (95% CI)	NLR (95% CI)	Colposcopy Referral (%) (95% CI)				
25–29 Years Prevalence (%) (95% Cl)=1.50 (1.19,1.87)											
Primary Screening Algorithm	76.53 (65.77, 85.86)	89.60 (88.81,90.35)	10.08 (7.55, 12.71)	99.60 (99.38, 99.78)	7.36 (6.18, 8.42)	0.26 (0.16, 0.38)	11.39 (10.64,12.20)				
Cytology Alone Algorithm	65.31 (54.12, 75.77)	87.66 (86.87, 88.49)	7.46 (5.54, 9.56)	99.40 (99.15, 99.61)	5.29 (4.36, 6.22)	0.40 (0.28, 0.52)	13.14 (12.33, 13.97)				
ASC-US Triage/Co-Testing Algorithm	64.29 (52.94, 73.96)	91.45 (90.73, 92.15)	10.28 (7.61, 13.27)	99.41 (99.15, 99.61)	7.52 (6.04, 8.99)	0.39 (0.28, 0.52)	9.39 (8.68, 10.11)				
		Prevalen	30-39 \ nce (%) (95% (/ears (1)=1.23 (1.02,	1.44)						
Primary Screening Algorithm	84.14 (77.23, 90.48)	92.30 (91.78, 92.79)	11.94 (9.70, 14.29)	99.79 (99.69, 99.88)	10.92 (9.84, 12.12)	0.17 (0.10, 0.25)	8.64 (8.15, 9.17)				
Cytology Alone Algorithm	68.28 (59.41, 76.34)	90.75 (90.24, 91.27)	8.39 (6.62, 10.02)	99.57 (99.42, 99.70)	7.38 (6.31, 8.40)	0.35 (0.26, 0.45)	9.98 (9.46, 10.51)				
ASC-US Triage/Co-Testing Algorithm	86.21 (79.43, 92.39)	91.30 (90.76, 91.80)	10.96 (8.90, 13.05)	99.81 (99.71, 99.90)	9.91 (8.95, 10.85)	0.15 (0.08, 0.23)	9.65 (9.13, 10.19)				
		Prevalen	40–65 \ ice (%) (95% (/ears Cl)=0.37 (0.27,	0.47)						
Primary Screening Algorithm	77.05 (64.81, 87.67)	94.62 (94.27, 94.95)	5.06 (3.66, 6.58)	99.91 (99.85, 99.95)	14.33 (11.91, 16.67)	0.24 (0.13, 0.37)	5.64 (5.32, 6.00)				
Cytology Alone Algorithm	62.30 (50.75, 76.36)	91.87 (91.46, 92.30)	2.77 (1.96, 3.79)	99.85 (99.78, 99.91)	7.67 (6.22, 9.53)	0.41 (0.26, 0.54)	8.33 (7.89, 8.75)				
ASC-US Triage/Co-Testing Algorithm	80.33 (68.94, 90.65)	93.82 (93.47, 94.17)	4.61 (3.39, 6.02)	99.92 (99.87, 99.97)	13.00 (11.09, 15.00)	0.21 (0.10, 0.33)	6.46 (6.11, 6.83)				

PPV=Positive predictive value; NPV=Negative predictive value; PLR=Positive likelihood ratio; NLR= Negative likelihood ratio.

Primary Screening Population (25-65 years) – risk estimates

The risks of high-grade cervical disease using the Primary Screening algorithm are presented in Table 51. Women positive for HPV16 and/or HPV18 (4.47%) and 12 Other HR HPV positive with ≥ASC-US cytology (3.27%) are referred for immediate colposcopy by the Primary Screening algorithm. The risks of ≥CIN2 were 18.63% (95% CI: 16.60, 20.70) for HPV16 and/or HPV18 positive, and 19.09% (95% CI: 16.60, 21.69) for 12 Other HR HPV positive with ≥ASC-US cytology (Table 51).

Women with 12 Other HR HPV positive and NILM cytology had a risk of 7.47% for \geq CIN2. The majority of women (84.92%) were HPV-negative and had a risk of 0.39% for \geq CIN2.

Table 51 Adjusted risk of disease in HPV and cytology categories by the primary screening algorithm (25-65 years)

cobas® 6800/8800 HPV Test Result	Proportion of women with Result (%)	Risk of ≥CIN2 (%) (95% CI)	Risk of ≥CIN3 (%) (95% CI)
HPV Positive	15.08	13.30 (12.26, 14.39)	5.56 (4.91, 6.18)
HPV16/18 Positive	4.47	18.63 (16.60, 20.70)	10.85 (9.27, 12.44)
HPV16 Positive	3.06	22.65 (19.99, 25.41)	14.00 (11.78, 16.09)
HPV18 Positive	1.42	9.94 (7.25, 12.99)	4.06 (2.42, 6.22)
12 Other HR HPV Positive and ≥ASC-US	3.27	19.09 (16.60, 21.69)	6.60 (5.11, 8.31)
12 Other HR HPV Positive and NILM Cytology	7.34	7.47 (6.25, 8.71)	1.88 (1.27, 2.52)
HPV Negative	84.92	0.39 (0.15, 0.88)	0.05 (0.02, 0.08)

The risks of high-grade cervical disease using the Primary Screening algorithm stratified by age group are presented in Table 52. The risks of \geq CIN2 were all above 10% in each age group for women with HPV16 and/or HPV18 positive results and women with 12 Other HR HPV positive result and \geq ASC-US cytology. The risk of \geq CIN3 was no more than 0.10% in each age group for women with a HPV negative test result (ranged from 0.03 to 0.10%).

Table 52 Risk of disease in HPV and cytology categories by the primary screening algorithm, stratified by age group

cobas [®] 6800/8800	Proportion of women	Risk of ≥CIN2 (%)	Risk of ≥CIN3 (%)	
HPV Test Result	with Result (%)	(95% CI)	(95% CI)	
	25-29 Years	•		
HPV Positive	24.01	15.18 (13.30, 17.26)	5.99 (4.71, 7.41)	
HPV16/18 Positive	4.84	22.15 (17.49, 27.60)	12.97 (8.87, 17.33)	
HPV16 Positive	3.61	27.12 (20.80, 34.03)	16.95 (11.60, 22.68)	
HPV18 Positive	1.23	7.50 (1.39, 15.79)	1.25 (0.00, 5.59)	
12 Other HR HPV Positive and ≥ASC-US	6.55	22.43 (17.81, 26.97)	7.94 (5.09, 11.23)	
12 Other HR HPV Positive and NILM Cytology	12.62	8.74 (6.65, 11.02)	2.31 (1.13, 3.55)	
HPV Negative	75.99	0.40 (0.22, 0.61)	0.10 (0.02, 0.22)	
	30-39 Years			
HPV Positive	16.44	15.74 (13.86, 17.65)	7.25 (6.04, 8.58)	
HPV16/18 Positive	5.27	24.08 (20.45, 28.09)	14.93 (11.92, 18.15)	
HPV16 Positive	3.57	29.62 (25.00, 35.02)	19.19 (15.10, 23.65)	
HPV18 Positive	1.70	12.44 (7.23, 17.47)	5.97 (2.78, 9.61)	
12 Other HR HPV Positive and ≥ASC-US	3.37	20.30 (15.97, 24.68)	7.27 (4.45, 10.26)	
12 Other HR HPV Positive and NILM Cytology	7.80	8.24 (6.29, 10.25)	2.06 (1.10, 3.14)	
HPV Negative	83.56	0.82 (0.13, 2.26)	0.04 (0.00, 0.08)	
	40-65 Years			
HPV Positive	10.56	8.86 (7.46, 10.34)	3.28 (2.42, 4.17)	
HPV16/18 Positive	3.76	11.33 (8.67, 13.84)	5.66 (3.90, 7.75)	
HPV16 Positive	2.47	12.81 (9.48, 16.23)	6.90 (4.40, 9.77)	
HPV18 Positive	1.29	8.49 (4.52, 12.53)	3.30 (0.91, 5.97)	
12 Other HR HPV Positive and ≥ASC-US	1.88	12.90 (9.10,17.47)	3.87 (1.84, 6.46)	
12 Other HR HPV Positive and NILM Cytology	4.92	5.31 (3.74, 7.21)	1.23 (0.50, 2.12)	
HPV Negative	89.44	0.10 (0.05, 0.17)	0.03 (0.01, 0.07)	

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Primary screening population (25-65 years) - risks of disease in women with NILM cytology and negative cobas[®] 6800/8800 HPV test results

Table 53 presents the absolute risks (AR) of disease (\geq CIN2 and \geq CIN3) for women with NILM Pap cytology, HPV-negative, and NILM with HPV-negative results. Risk of \geq CIN3 in women with NILM Pap cytology was 0.33% compared with 0.05% among women with negative **cobas*** 6800/8800 HPV test results. This indicates that women with NILM Pap cytology have 6.6 (0.33/0.05) times higher risk of \geq CIN3 compared with women with HPV-negative results. The addition of a NILM cytology result to a negative **cobas*** 6800/8800 HPV test result marginally decreased \geq CIN3 risk.

Table 53 Adjusted risk of disease in women with NILM cytology and negative cobas® 6800/8800 HPV test results

Cytology and cobas® 6800/8800 HPV Test Result			Risk of ≥CIN3 (%) (95% CI)
NILM	90.21% (31,399/34,807)	1.13 (0.84, 1.61)	0.33 (0.26, 0.40)
HPV Negative	84.92% (29,557/34,807)	0.39 (0.15, 0.88)	0.05 (0.02, 0.08)
NILM and HR HPV Negative	79.76% (27,763/34,807)	0.25 (0.01, 0.76)	0.00 (0.00, 0.01)

Primary screening population (25-65 years) - benefit and risk per 10,000 women

Benefits and risks per 10,000 women using the Primary Screening, Cytology Alone, and ASC-US Triage/Co-testing algorithms are presented in Table 54. Per 10,000 women, the Primary Screening algorithm correctly identified the highest number of true positive ≥CIN3 cases (70) compared to the Cytology Alone algorithm (58) and ASC-US Triage/Co-testing algorithm (68). The Primary Screening algorithm was associated with fewer colposcopies compared to the Cytology Alone algorithm and the ASC-US Triage/Co-testing algorithm (775 vs. 980 and 810, respectively). Fewer cases of ≥CIN3 high grade disease were missed by the Primary Screening algorithm (17 vs. 29 and 19) as well as fewer false positive CIN2 (<CIN2) were identified with the Primary Screening algorithm compared with Cytology Alone, and ASC-US Triage/Co-testing algorithms (628 vs. 848 and 663, respectively).

Table 54 Benefit and risk of using the primary screening, cytology alone, and ASC-US triage/co-testing algorithms in the primary screening population (25-65 years) per 10,000 women

				Benefit		Risk		
	Num	ber of Tests and Pro	True Po	ositives	False No	egatives	False Positives	
Algorithm	Pap Cytology	cobas [®] 6800/8800 HPV	Colposcopy	≥CIN3	CIN2	≥CIN3	CIN2	<cin2< th=""></cin2<>
Primary Screening	1,061	10,000	775	70	77	17	70	628
Cytology Alone	10,000	0	980	58	74	29	73	848
ASC-US Triage/Co-testing	10,000	8,043	810	68	79	19	68	663

Primary screening population (25-65 years) - benefits and risk per 100 colposcopy procedures

Benefits and risks per 100 colposcopy procedures when using the Primary Screening, Cytology Alone, and ASC-US Triage/Co-testing algorithms are presented in Table 55. For the Primary Screening algorithm, the number of screening tests that had to be performed to select 100 women for colposcopy was 1,427 (137+1,290); 1,020 were required for the Cytology alone algorithm, while 2,228 (1,235+993) were required for the ASC-US Triage/Co-testing algorithm. The number of true positives (≥CIN2) by the Primary Screening algorithm was 19 per 100 colposcopies compared to 14 for Cytology alone, and 18 for the ASC-US Triage/Co-testing algorithm. The probability of ≥CIN3 among women not referred to colposcopy was 0.17% (2/1,190) by the Primary Screening algorithm, 0.30% (3/920) by the Cytology alone algorithm and 0.18% (2/1,135) by the ASC-US Triage/Co-testing algorithm.

Table 55 Benefit and risks of the primary screening, cytology alone and ASC-US triage/co-testing algorithms in the primary screening population (25-65 years) per 100 colposcopy procedures

			Ber	nefit	Ri			
	Nun	nber of Tests and Prod	edures	True Po	ositives	False No	False Positives	
Algorithm	Pap Cytology	cobas [®] 6800/8800 HPV	Colposcopy	≥CIN3	CIN2	≥CIN3	CIN2	<cin2< th=""></cin2<>
Primary Screening	137	1,290	100	9	10	2	9	81
Cytology Alone	1,020	0	100	6	8	3	7	86
ASC-US Triage/Co-testing	1,235	993	100	8	10	2	8	82

Performance by vaccination status

The performance of the **cobas**° 6800/8800 HPV test was also evaluated by self-reported vaccination status in the 25-29 year age group. Among the 25-29 year participants, 39% self-reported having received the HPV vaccine. The performance of **cobas**° 6800/8800 HPV in unvaccinated and vaccinated women with ASC-US cytology (25-29 years old) is presented in Table 56. Results in a subset of the primary screening population (25-29 years old) stratified by self-reported vaccination status is presented in Table 57.

Table 56 Performance of the cobas® 6800/8800 HPV test in detecting disease, stratified by HPV vaccination status in the ASC-US population (25-29 years)

Statistic	Overall	Vaccinated	Unvaccinated
·		≥CIN2	
Sensitivity (%)	89.13 (41/46)	78.57 (11/14)	93.75 (30/32)
(95% CI)	(76.96, 95.27)	(52.41, 92.43)	(79.85, 98.27)
Specificity (%)	51.35 (171/333)	52.76 (67/127)	50.49 (104/206)
(95% CI)	(46.00, 56.67)	(44.12, 61.23)	(43.71, 57.24)
PPV (%)	20.20 (41/203)	15.49 (11/71)	22.73 (30/132)
(95% CI)	(15.25, 26.25)	(8.88, 25.65)	(16.41, 30.59)
NPV (%)	97.16 (171/176)	95.71 (67/70)	98.11 (104/106)
(95% CI)	(93.52, 98.78)	(88.14, 98.53)	(93.38, 99.48)
Prevalence (%)	12.14 (46/379)	9.93 (14/141)	13.45 (32/238)
(95% CI)	(9.22 ,15.81)	(6.01 ,15.98)	(9.69, 18.36)

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Statistic	Overall	Vaccinated	Unvaccinated
·		≥CIN3	
Sensitivity (%)	93.33 (14/15)	83.33 (5/6)	100.00 (9/9)
(95% CI)	(70.18, 98.81)	(43.65, 96.99)	(70.09, 100.00)
Specificity (%)	48.08 (175/364)	51.11 (69/135)	46.29 (106/229)
(95% CI)	(42.99, 53.20)	(42.77, 59.40)	(39.94, 52.75)
PPV (%)	6.90 (14/203)	7.04 (5/71)	6.82 (9/132)
(95% CI)	(4.15, 11.24)	(3.05, 15.45)	(3.63, 12.45)
NPV (%)	99.43 (175/176)	98.57 (69/70)	100.00 (106/106)
(95% CI)	(96.85, 99.90)	(92.34, 99.75)	(96.50, 100.00)
Prevalence (%)	3.96 (15/379)	4.26 (6/141)	3.78 (9/238)
(95% CI)	(2.41 ,6.43)	(1.96 ,8.97)	(2.00 ,7.03)

PPV=Positive predictive value; NPV=Negative predictive value.

Table 57 Performance of the cobas® 6800/8800 HPV test in detecting disease, stratified by HPV vaccination status in the primary screening population (25-29 years)

	Unac	ljusted	Adjı	usted
Statistics	Vaccinated	Unvaccinated	Vaccinated	Unvaccinated
		≥CIN2		
Sensitivity (%)	55.17 (32/58)	69.12 (94/136)	54.67	68.51
(95% CI)	(42.45, 67.25)	(60.92, 76.27)	(42.22, 68.01)	(60.98, 76.60)
Specificity (%)	75.56 (405/536)	64.21(531/827)	93.04	89.29
(95% CI)	(71.75, 79.01)	(60.88, 67.40)	(91.99, 94.06)	(88.24, 90.39)
PPV (%)	19.63 (32/163)	24.10 (94/390)	19.07	23.48
(95% CI)	(15.64, 24.35)	(21.55, 26.85)	(13.67, 25.44)	(19.60, 27.86)
NPV (%)	93.97 (405/431)	92.67 (531/573)	98.56	98.34
(95% CI)	(92.10, 95.41)	(90.73, 94.23)	(97.98, 99.07)	(97.85, 98.82)
PLR	2.26 (32/58)/(131/536)	1.93 (94/136)/(296/827)	7.85	6.40
(95% CI)	(1.71, 2.97)	(1.67, 2.23)	(5.87, 10.51)	(5.51, 7.47)
NLR	0.59 (26/58)/(405/536)	0.48 (42/136)/(531/827)	0.49	0.35
(95% CI)	(0.44, 0.79)	(0.37, 0.62)	(0.34, 0.62)	(0.26, 0.44)
Colposcopy Referral (%)	27.44 (163/594)	40.50 (390/963)	8.35	13.35
(95% CI)	(24.01, 31.17)	(37.44, 43.63)	(7.34, 9.42)	(12.29, 14.48)
Prevalence (%)	9.76 (58/594)	14.12 (136/963)	2.91	4.58
(95% CI)	(7.63, 12.42)	(12.07, 16.46)	(2.25, 3.65)	(3.88, 5.36)
		≥CIN3		
Sensitivity (%)	65.38 (17/26)	81.63 (40/49)	66.67	80.00
(95% CI)	(46.22, 80.59)	(68.64, 90.02)	(45.83, 83.87)	(69.78, 92.91)
Specificity (%)	74.30 (422/568)	61.71 (564/914)	92.41	87.76
(95% CI)	(70.55, 77.72)	(58.51, 64.80)	(91.32, 93.42)	(86.68, 88.90)
PPV (%)	10.43 (17/163)	10.26 (40/390)	10.23	9.85
(95% CI)	(7.85, 13.73)	(8.90, 11.79)	(5.94, 15.23)	(7.24, 13.19)
NPV (%)	97.91 (422/431)	98.43 (564/573)	99.53	99.62
(95% CI)	(96.50, 98.76)	(97.20, 99.13)	(99.15, 99.79)	(99.39, 99.88)
PLR	2.54 (17/26)/(146/568)	2.13 (40/49)/(350/914)	8.78	6.54
(95% CI)	(1.86, 3.48)	(1.82, 2.49)	(5.85, 11.63)	(5.62, 7.77)
NLR	0.47 (9/26)/(422/568)	0.30 (9/49)/(564/914)	0.36	0.23
(95% CI)	(0.27, 0.79)	(0.16, 0.54)	(0.17, 0.59)	(0.08, 0.34)
olposcopy Referral (%)	27.44 (163/594)	40.50 (390/963)	8.35	13.35
(95% CI)	(24.01, 31.17)	(37.44, 43.63)	(7.34, 9.42)	(12.29, 14.48)
Prevalence (%)	4.38 (26/594)	5.09 (49/963)	1.28	1.64
(95% CI)	(3.00, 6.34)	(3.87, 6.66)	(0.85, 1.83)	(1.21, 2.10)

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Comparison of results from the $cobas^{\tiny{(8)}}$ 6800/8800 HPV test for pre-quot vs. post-quot clinical samples

Within IMPACT, a sub-study was designed to compare the performance of the **cobas**° 6800/8800 HPV test on cervical specimens tested prior to Pap cytology processing (pre-quot) and after Pap cytology processing (post-quot) on the ThinPrep° 2000 Processor (Hologic Inc.). The **cobas**° 6800/8800 HPV test was performed on 3,753 paired pre-quot and post-quot samples.

Agreement between the **cobas*** 6800/8800 HPV test results of pre-quot samples and post-quot samples for any HPV and for genotype-specific results are presented in Table 58, Table 59, and Table 60 for each of the three study populations (ASC-US 25-65 years, NILM 30-65 years, and the primary screening population 25-65 years).

Table 58 Agreement of cobas® 6800/8800 HPV test results in pre-quot vs. post-quot samples in the ASC-US population (25-65 years), stratified by CPR diagnosis

		Pre-quo	t Cytology Samples				
		≥CII	N2				
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total		
HPV16 Positive	3	0	0	0	3		
HPV18 Positive	0	0	0	0	0		
12 Other HR HPV Positive	0	0	2	0	2		
HPV Negative	0	0	0	2	2		
Total	3	0	2	2	7		
Genotype Specific PPA (95% CI)	100.0% (3/3) (43.85%, 100.0%)	NC	100.0% (2/2) (34.24%, 100.0%)				
14 HR HPV Percent Agreement (95% CI)	Agreement PPA=100.0% (5/5)						
		<cii< td=""><td>N2</td><td></td><td></td></cii<>	N2				
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total		
HPV16 Positive	6	0	1	1	8		
HPV18 Positive	0	3	0	0	3		
12 Other HR HPV Positive	0	0	32	0	32		
HPV Negative	0	0	4	121	125		
Total	6	3	37	122	168		
Genotype Specific PPA (95% CI)	100.0% (6/6) (60.97%, 100.0%)	100.0% (3/3) (43.85%, 100.0%)	86.49% (32/37) (72.02%, 94.09%)				
14 HR HPV Percent Agreement (95% CI)		PPA=91.30% (42/4 (79.68%, 96.57%		NPA=99.18% (121/122) (95.50%, 99.86%)			

NC=not calculable

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Table 59 Agreement of cobas® 6800/8800 HPV test results in pre-quot vs. post-quot samples in the NILM population (30-65 years), stratified by CPR diagnosis

		Pre-quot	t Cytology Samples		
		≥CII	N2		
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	6	0	0	0	6
HPV18 Positive	0	0	0	0	0
12 Other HR HPV Positive	0	0	8	0	8
HPV Negative	0	0	0	1	1
Total	6	0	8	1	15
Genotype Specific PPA (95% CI)	100.00% (6/6) (60.97%, 100.0%)	NC	100.0% (8/8) (67.56%, 100.0%)		
14 HR HPV Percent Agreement (95% CI)		PPA=100.0% (14/ (78.47%, 100.0%	-	NPA=100.0% (1/1) (20.65%, 100.0%)	
		<cii< td=""><td>N2</td><td></td><td></td></cii<>	N2		
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	41	0	0	2	43
HPV18 Positive	0	17	1	0	18
12 Other HR HPV Positive	1	0	107	5	113
HPV Negative	8	2	19	58	87
Total	50	19	127	65	261
Genotype Specific PPA (95% CI)	82.00% (41/50) (69.20%, 90.23%)	89.47% (17/19) (68.61%, 97.06%)	84.25% (107/127) (76.92%, 89.57%)		
14 HR HPV Percent Agreement (95% CI)		PPA=85.20% (167/ (79.56%, 89.50%		NPA=89.23% (58/65) (79.40%, 94.68%)	

NC=not calculable

Table 60 Agreement of cobas® 6800/8800 HPV test results in pre-quot vs. post-quot samples in the primary screening population (25-65 years), stratified by CPR diagnosis

		Pre-quot	Cytology Samples		
		≥CIN2			
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	18	0	0	0	18
HPV18 Positive	0	5	0	0	5
12 Other HR HPV Positive	0	0	30	1	31
HPV Negative	0	0	0	5	5
Total	18	5	30	6	59
Genotype Specific PPA (95% CI)	100.0% (18/18) (82.41%, 100.0%)	100.0% (5/5) (56.55%, 100.0%)	100.0% (30/30) (88.65%, 100.0%)		
14 HR HPV Percent Agreement (95% CI)		PPA=100.0% (53/5 (93.24%, 100.0%)		NPA=83.33% (5/6) (43.65%, 96.99%)	
		<cin2< td=""><td></td><td></td><td></td></cin2<>			
Post-quot Cytology Samples	HPV16 Positive	HPV18 Positive	12 Other HR HPV Positive	HPV Negative	Total
HPV16 Positive	67	0	2	4	73
HPV18 Positive	0	28	1	0	29
12 Others Positive	1	0	218	7	226
HPV Negative	8	2	26	237	273
Total	76	30	247	248	601
Genotype Specific PPA (95% CI)	88.16% (67/76) (79.00%, 93.64%)	93.33% (28/30) (78.68%, 98.15%)	88.26% (218/247) (83.65%, 91.70%)		
14 HR HPV Percent Agreement (95% CI)		PPA=89.80% (317/3 (86.20%, 92.54%)		NPA=95.56% (237/248) (92.23%, 97.51%)	

Site-to-site reproducibility

To evaluate site-to-site reproducibility testing was performed at three testing sites, using one reagent lot and four **cobas**° Systems (three **cobas**° 6800 Systems at all three testing sites and one **cobas**° 8800 System at one of those site). Each panel member was tested for five days, three replicates per run, on the four Systems. Two operators performed one run per day for five days for each System. A 13-member panel composed of pools made from clinical samples collected into PreservCyt° Solution, and from samples derived from SiHa and HeLa cell lines was tested for reproducibility.

Table 61 summarizes results for the negative panel members by site/instrument, operator/run, and day on the three **cobas*** 6800 Systems and 1 **cobas*** 8800 System. All negative panel members were correctly identified as negative across site/instrument, operator/run and testing day.

Percent of positive results for the positive panel members are presented in Table 62. Analysis of variance of the Ct values from tests performed on positive panel members yielded total CV(%) ranging from 1.1% to 5.6% across all panel members. The CV(%) ranged from 1.1% to 2.7% for the cell line panel members and 2.1% to 5.6% for the pooled clinical panel members. The largest component of variance observed (1.71 for Pooled HPV45 Low Positive at 1 x LoD) among all positive panel members was for within-run (Table 63).

Table 61 Agreement and variability for negative panel member for site/instrument, operator/run, and day on the cobas® 6800/8800 Systems

					Number	of Ne	egatives/Tota	l Number of	Vali	d Results			
			Between-Site/Instrument				etween-Oper	ator/Run		Between-Day			
Panel Member	Ct SD	Ct CV%	ID	Negative Agreement (%)	Negative/ Valid	ID*	Negative Agreement (%)	Negative/ Valid	ID	Negative Agreement (%)	Negative/ Valid		
	n/a	n/a	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24		
			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24		
Negative background			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24		
cell line			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24		
						5	100.0	30/30	5	100.0	24/24		
						6	100.0	30/30					
	n/a	n/a	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24		
			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24		
Negative pooled clinical			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24		
samples			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24		
						5	100.0	30/30	5	100.0	24/24		
						6	100.0	30/30					

^{*}Note: Operators 1 and 2 were at testing site 1; Operators 3 and 4 were at testing site 2; Operators 5 and 6 were at testing site 3.

Table 62 Agreement and variability for positive panel members for site/instrument, operator/run, and day on the cobas® 6800/8800 Systems

				Nu	ımber of Po	sitive	Results/Total	Number of \	/alid	Results		
			Be	tween-Site/I	nstrument	Between-Operator/Run				Between-Day		
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID¹	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid	
	Po	sitive Ce	ell Lir	ne Panel Mem	bers: HPV16	6/18 V	Veak Positive	(0.3 x LoD)				
	0.76	2.1	11	66.7	20/30	1	60.0	9/15	1	58.3	14/24	
			21	76.7	23/30	2	73.3	11/15	2	54.2	13/24	
HPV16 Weak Positive			31	46.7	14/30	3	93.3	14/15	3	62.5	15/24	
(0.3 x LoD)			32	60.0	18/30	4	60.0	9/15	4	83.3	20/24	
						5	53.3	16/30	5	54.2	13/24	
						6	53.3	16/30				
	0.96	2.7	11	53.3	16/30	1	40.0	6/15	1	70.8	17/24	
			21	60.0	18/30	2	66.7	10/15	2	66.7	16/24	
HPV18 Weak Positive			31	60.0	18/30	3	60.0	9/15	3	45.8	11/24	
(0.3 x LoD)			32	70.0	21/30	4	60.0	9/15	4	70.8	17/24	
						5	73.3	22/30	5	50.0	12/24	
						6	56.7	17/30				
	F	Positive (Cell L	ine Panel Me	mbers: HPV	16/18	Low Positive	(1 x LoD)		•		
	0.47	1.3	11	96.7	29/30	1	100.0	15/15	1	95.8	23/24	
	0117		21	96.7	29/30	2	93.3	14/15	2	100.0	24/24	
HPV16 Low Positive			31	100.0	30/30	3	93.3	14/15	3	100.0	24/24	
(1 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
(. × 202)			02	100.0	00/00	5	100.0	30/30	5	95.8	23/24	
						6	100.0	30/30	Ť	00.0	20,21	
	0.63	1.9	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
	0.00	1.0	21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
HPV18 Low Positive			31	96.7	29/30	3	100.0	15/15	3	100.0	24/24	
(1 x LoD)			32	100.0	30/30	4	100.0	15/15	4	95.8	23/24	
(. × 202)			02	100.0	00/00	5	96.7	29/30	5	100.0	24/24	
						6	100.0	30/30	3	100.0	24/24	
		Positiv	e Cel	⊥ I Line Panel N	lembers: HP				1			
	0.37	1.1	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
	0.37	1.1	21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
UDV/16 Docitive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
HPV16 Positive (3 x LoD)			32	100.0	29/29	4	100.0	15/15	4	100.0	23/23	
(O A LOD)			32	100.0	23/23	5	100.0	30/30	5	100.0	24/24	
						1			J	100.0	24/24	
	0.40	1.0	11	100.0	20/20	6	100.0	29/29	1	1000	24/24	
	0.40	1.2	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
LIDI(so D			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
HPV18 Positive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
(3 x LoD)			32	100.0	29/29	4	100.0	15/15	4	100.0	23/23	
						5	100.0	30/30	5	100.0	24/24	
						6	100.0	29/29				

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						sitive	Results/Total	Number of \	/alid	Results		
			Be	tween-Site/I	nstrument	В	etween-Opera	ator/Run		Between-Day		
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID¹	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid	
			I		Clinical Pane	l Men				(70)	<u> </u>	
	1.07	3.2	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
	1.07	3.2	21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV16 Low			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
Positive			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
(1 x LoD)			32	100.0	30/30	5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30	3	100.0	24/24	
	0.89	2.7	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
	0.00	2.7	21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV16 Positive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
3 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
(6 × 202)			02	100.0	00/00	5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30	Ť	100.0	2 1/2 1	
	0.74	2.1	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
	0.7 1		21	96.7	29/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV18 Low			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
Positive			32	100.0	30/30	4	93.3	14/15	4	95.8	23/24	
(1 x LoD)				100.0	00,00	5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30	Ť	100.0		
	0.92	2.7	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV18 Positive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
(3 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
						5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30				
	1.80	5.6	11	96.7	29/30	1	100.0	15/15	1	100.0	24/24	
			21	100.0	30/30	2	93.3	14/15	2	100.0	24/24	
Pooled HPV45 Low			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
Positive			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
(1 x LoD)						5	100.0	30/30	5	95.8	23/24	
						6	100.0	30/30				
	1.54	5.2	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV45 Positive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
(3 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
						5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30				
	1.04	3.1	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24	
D			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24	
Pooled HPV39 Low			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24	
Positive (1 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24	
(1 X LUD)						5	100.0	30/30	5	100.0	24/24	
						6	100.0	30/30				

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				Number of Positive Results/Total Number of Valid Results									
			Between-Site/Instrument			Between-Operator/Run				Between-Day			
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID¹	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid		
	1.45	4.6	11	100.0	30/30	1	100.0	15/15	1	100.0	24/24		
			21	100.0	30/30	2	100.0	15/15	2	100.0	24/24		
Pooled HPV39 Positive			31	100.0	30/30	3	100.0	15/15	3	100.0	24/24		
(3 x LoD)			32	100.0	30/30	4	100.0	15/15	4	100.0	24/24		
						5	100.0	30/30	5	100.0	24/24		
						6	100.0	30/30					

Note: ¹Operators 1 and 2 were at testing site 1; Operators 3 and 4 were at testing site 2; Operators 5 and 6 were at testing site 3.

Table 63 Overall mean, standard deviation, and coefficients of variation (%) for cycle threshold, estimated from positive panel members

			Stan	dard Deviation	, Coefficient of	Variation (%)	
Panel Member	N	Mean Ct	Between- Site/Instru- ment	Between- Operator/ Run	Between- Day	Within- Run	Total CV
Positive Cell Line Panel Members							
HPV16/18 Weak Positive (0.3 x LoD)							
HPV16 Weak Positive (0.3 x LoD)	77	36.6	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.76, (2.08%)	2.1
HPV18 Weak Positive (0.3 x LoD)	74	35.3	0.00, (0.00%)	0.00, (0.00%)	0.12, (0.34%)	0.95, (2.69%)	2.7
HPV16/18 Low Positive (1 x LoD)							
HPV16 Low Positive (1 x LoD)	118	35.6	0.10, (0.27%)	0.00, (0.00%)	0.15, (0.43%)	0.43, (1.22%)	1.3
HPV18 Low Positive (1 x LoD)	119	34.1	0.00, (0.00%)	0.09, (0.28%)	0.00, (0.00%)	0.63, (1.83%)	1.9
HPV16/18 Positive (3 x LoD)							
HPV16 Positive (3 x LoD)	119	34.7	0.05, (0.16%)	0.00, (0.00%)	0.11, (0.31%)	0.35, (1.01%)	1.1
HPV18 Positive (3 x LoD)	119	32.9	0.05, (0.16%)	0.08, (0.25%)	0.00, (0.00%)	0.39, (1.19%)	1.2
Positive Clinical Panel Members	<u>'</u>		1	,	,	,	
Pooled HPV16 Low Positive (1 x LoD)	120	33.6	0.25, (0.73%)	0.00, (0.00%)	0.00, (0.00%)	1.05, (3.11%)	3.2
Pooled HPV16 Positive (3 x LoD)	120	33.1	0.30, (0.90%)	0.00, (0.00%)	0.00, (0.00%)	0.84, (2.53%)	2.7
Pooled HPV18 Low Positive (1 x LoD)	119	35.1	0.00, (0.00%)	0.00, (0.00%)	0.11, (0.31%)	0.74, (2.09%)	2.1
Pooled HPV18 Positive (3 x LoD)	120	34.0	0.56, (1.64%)	0.00, (0.00%)	0.21, (0.62%)	0.70, (2.06%)	2.7
Pooled HPV45 Low Positive (1 x LoD)	120	31.9	0.56, (1.74%)	0.00, (0.00%)	0.00, (0.00%)	1.71, (5.37%)	5.6
Pooled HPV45 Positive (3 x LoD)	120	29.7	0.00, (0.00%)	0.00, (0.00%)	0.60, (2.04%)	1.42, (4.79%)	5.2
Pooled HPV39 Low Positive (1 x LoD)	120	33.4	0.20, (0.61%)	0.00, (0.00%)	0.33, (0.98%)	0.97, (2.90%)	3.1
Pooled HPV39 Positive (3 x LoD)	120	31.5	0.00, (0.00%)	0.00, (0.00%)	0.62, (1.95%)	1.31, (4.15%)	4.6

Notes: Ct=Cycle Threshold; CV=Coefficient of Variation

² One replicate failed due to processing error and excluded from analysis.

Lot-to-lot variability

Lot-to-lot variability was evaluated at one testing site, using three reagent lots for each of the two Systems separately (**cobas**° 6800 and **cobas**° 8800). This study used the same panel as described in the site-to-site reproducibility study. Each panel member was tested for 15 days (5 days per lot), three replicates per run, for each of the two **cobas**° Systems. Two operators performed one run per day for 5 days for each reagent lot.

Table 64 and Table 65 show results for the negative panel member by reagent lot, operator/run and day on the **cobas*** 6800 System and on the **cobas*** 8800 System, respectively. In both Systems all negative panel members were correctly identified as negative across reagent lot, operator/run and testing day.

Table 64 Agreement and variability for negative panel members by lot, operator/run, and day on the cobas® 6800 System

					Numb	er Ne	gative/Total Nu	mber Valid F	Resul	ts		
			Between-Lot				Between-Opera	tor/Run		Between-Day		
Panel Member	Ct SD	Ct CV%	ID	Negative Agreement (%)	Negative/ Valid	ID	Negative Agreement (%)	Negative/ Valid	ID	Negative Agreement (%)	Negative/ Valid	
	n/a	n/a	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
Negative			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
background cell			3	100.0	30/30				3	100.0	18/18	
line									4	100.0	18/18	
									5	100.0	18/18	
	n/a	n/a	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
.			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Negative pooled clinical samples			3	100.0	30/30				3	100.0	18/18	
ciinicai sanipies									4	100.0	18/18	
									5	100.0	18/18	

Table 65 Agreement and variability for negative panel members by lot, operator/run, and day on the cobas® 8800 System

					Number	of N	egatives/Total N	Number Valid	Res	ults	_	
			Between-Lot				Between-Opera	tor/Run		Between-Day		
Panel Member	Ct SD	Ct CV%	ID	Negative Agreement (%)	Negative/ Valid	ID	Negative Agreement (%)	Negative/ Valid	ID	Negative Agreement (%)	Negative/ Valid	
			1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
Negative			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
background cell	N/A	N/A	3	100.0	30/30				3	100.0	18/18	
line									4	100.0	18/18	
									5	100.0	18/18	
			1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
N			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Negative pooled clinical samples	N/A	N/A	3	100.0	30/30				3	100.0	18/18	
ciiriicai sampies									4	100.0	18/18	
									5	100.0	18/18	

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Table 66 presents the percent of positive results for the positive panel members by reagent lot, operator/run and day on the **cobas**° 6800 System. Analysis of variance of the Ct values from valid tests performed on positive panel members yielded total CV(%) ranging of 0.9% to 5.0% across all panel members. The CV(%) ranged from 0.9% to 2.2% for the cell line panel members and 1.7% to 5.0% for the pooled clinical panel members (Table 66). The largest component of variance observed (1.55 for Pooled HPV45 Low Positive at 1 x LoD) among all positive panel members on the **cobas**° 6800 System was for within-run (Table 67).

Table 68 presents the percent of positive results for the positive panel members by reagent lot, operator/run and day on the **cobas**° 8800 System. Analysis of variance of the Ct values from valid tests performed on positive panel members yielded total CV(%) ranging of 1.1% to 7.4% across all panel members. The CV(%) ranged from 1.1% to 3.0% for the cell line panel members and 2.0% to 7.4% for the pooled clinical panel members (Table 69). The largest component of variance observed (2.16 for Pooled HPV45 Low Positive at 1 x LoD) among all positive panel members on the **cobas**° 8800 System was for within-run (Table 69).

Table 66 Agreement and variability for positive panel members for lot, operator, and day on the cobas® 6800 System

					Number	of Po	ositives/Total	Number Va	alid F	lesults	
				Between-	Lot	В	etween-Opera	ator/Run		Between-E	Day
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID*	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid
	Positi	ve Cell	Line	Panel Memb	ers: HPV16	/18	Weak Positive	e (0.3 x Lo[))		
	0.75	2.0	1	46.7	14/30	5	57.8	26/45	1	61.1	11/18
LIDV10 March Description			2	46.7	14/30	6	53.3	24/45	2	44.4	8/18
HPV16 Weak Positive (0.3 x LoD)			3	73.3	22/30				3	38.9	7/18
(0.0 x 20D)									4	77.8	14/18
									5	55.6	10/18
	0.77	2.2	1	60.0	18/30	5	62.2	28/45	1	66.7	12/18
HDV/19 Wools Dooiting			2	60.0	18/30	6	66.7	30/45	2	66.7	12/18
HPV18 Weak Positive (0.3 x LoD)			3	73.3	22/30				3	66.7	12/18
(,									4	66.7	12/18
									5	55.6	10/18
	Posi	tive Ce	II Lir	ne Panel Men	nbers: HPV	16/18	B Low Positive	e (1 x LoD)			
	0.50	1.4	1	100.0	30/30	5	97.8	44/45	1	94.4	17/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
HPV16 Low Positive (1 x LoD)			3	96.7	29/30				3	100.0	18/18
(1 x 20D)									4	100.0	18/18
									5	100.0	18/18
	0.67	2.0	1	96.7	29/30	5	97.8	44/45	1	100.0	18/18
HPV18 Low Positive			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
(1 x LoD)			3	100.0	30/30				3	100.0	18/18
									4	94.4	17/18
									5	100.0	18/18

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					Number	of Po	ositives/Total	Number Va	alid F	Results	
				Between-		ı —	etween-Oper			Between-E)ay
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID*	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid
	Po	ositive	Cell	Line Panel M	embers: HF	PV16	/18 Positive (3 x LoD)			
	0.31	0.9	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
HPV16 Positive (3 x LoD)			3	100.0	30/30				3	100.0	18/18
(0 X 202)									4	100.0	18/18
									5	100.0	18/18
	0.39	1.2	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
HPV18 Positive (3 x LoD)			3	100.0	30/30				3	100.0	18/18
(0 X 20D)									4	100.0	18/18
									5	100.0	18/18
				Positive Cl	inical Pane	l Me	mbers				
	1.13	3.4	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Pooled HPV16 Low Positive (1 x LoD)			3	100.0	30/30				3	100.0	18/18
(1 X 200)									4	100.0	18/18
									5	100.0	18/18
	1.00	3.0	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Pooled HPV16 Positive (3 x LoD)			3	100.0	30/30				3	100.0	18/18
(3 × L0D)									4	100.0	18/18
									5	100.0	18/18
	0.60	1.7	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Pooled HPV18 Low Positive (1 x LoD)			3	100.0	30/30				3	100.0	18/18
(1 x LOD)									4	100.0	18/18
									5	100.0	18/18
	0.86	2.5	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Pooled HPV18 Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	18/18
									5	100.0	18/18
		1	1	1	1		1	I	1		<u> </u>

					Number	of Po	ositives/Total	Number Va	alid F	Results		
				Between-	Lot	Ве	etween-Opera	ator/Run		Between-Day		
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID*	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid	
	1.60	5.0	1	100.0	30/30	5	97.8	44/45	1	100.0	18/18	
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Pooled HPV45 Low Positive (1 x LoD)			3	96.7	29/30				3	100.0	18/18	
(1 × 200)									4	94.4	17/18	
									5	100.0	18/18	
	1.46	4.9	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Pooled HPV45 Positive (3 x LoD)			3	100.0	30/30				3	100.0	18/18	
(3 x 200)									4	100.0	18/18	
									5	100.0	18/18	
	0.75	2.3	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Pooled HPV39 Low Positive (1 x LoD)			3	100.0	30/30				3	100.0	18/18	
(1 x 200)									4	100.0	18/18	
									5	100.0	18/18	
	0.84	2.6	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18	
			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18	
Pooled HPV39 Positive (3 x LoD)			3	100.0	30/30				3	100.0	18/18	
(0 × 20D)									4	100.0	18/18	
									5	100.0	18/18	

^{*}Note: Operators 5 and 6 were at testing site 3.

Table 67 Overall mean, standard deviation, and coefficients of variation (%) for cycle threshold, estimated from positive panel members on the cobas[®] 6800 System

			Standard Deviation, Coefficient of Variation (%)								
Panel Member	N	Mean Ct	Between- Lot	Between- Operator/Run	Between- Day	Within- Run	Total CV				
Positive Cell Line Panel Members				_							
HPV16/18 Weak Positive (0.3 x LoD)											
HPV16 Weak Positive (0.3 x LoD)	52	36.5	0.07, (0.20%)	0.00, (0.00%)	0.28, (0.78%)	0.69, (1.88%)	2.0				
HPV18 Weak Positive (0.3 x LoD)	58	35.4	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.77, (2.19%)	2.2				
HPV16/18 Low Positive (1 x LoD)											
HPV16 Low Positive (1 x LoD)	89	35.6	0.09, (0.24%)	0.04, (0.13%)	0.00, (0.00%)	0.49, (1.37%)	1.4				
HPV18 Low Positive (1 x LoD)	89	34.1	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.67, (1.97%)	2.0				
HPV16/18 Positive (3 x LoD)											
HPV16 Positive (3 x LoD)	90	34.6	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.31, (0.88%)	0.9				
HPV18 Positive (3 x LoD)	90	32.9	0.00, (0.00%)	0.00, (0.00%)	0.13, (0.41%)	0.36, (1.10%)	1.2				
Positive Clinical Panel Members			-			·					
Pooled HPV16 Low Positive (1 x LoD)	90	33.5	0.11, (0.32%)	0.00, (0.00%)	0.00, (0.00%)	1.12, (3.35%)	3.4				
Pooled HPV16 Positive (3 x LoD)	90	33.1	0.11, (0.33%)	0.00, (0.00%)	0.00, (0.00%)	1.00, (3.01%)	3.0				
Pooled HPV18 Low Positive (1 x LoD)	90	35.1	0.14, (0.41%)	0.00, (0.00%)	0.00, (0.00%)	0.58, (1.67%)	1.7				
Pooled HPV18 Positive (3 x LoD)	90	33.7	0.00, (0.00%)	0.26, (0.76%)	0.14, (0.43%)	0.81, (2.39%)	2.5				
Pooled HPV45 Low Positive (1 x LoD)	90	32.0	0.00, (0.00%)	0.00, (0.00%)	0.42, (1.31%)	1.55, (4.84%)	5.0				
Pooled HPV45 Positive (3 x LoD)	90	29.7	0.18, (0.62%)	0.00, (0.00%)	0.00, (0.00%)	1.45, (4.89%)	4.9				
Pooled HPV39 Low Positive (1 x LoD)	90	33.3	0.00, (0.00%)	0.00, (0.00%)	0.23, (0.69%)	0.71, (2.14%)	2.3				
Pooled HPV39 Positive (3 x LoD)	90	31.6	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.84, (2.65%)	2.6				

Notes: Ct=Cycle Threshold; CV=Coefficient of Variation

Table 68 Agreement and variability for positive panel member by lot, operator/run, and day on the cobas® 8800 System

					Numb	er of P	ositives/Total N	umber Valid F	Result	ts	
				Between-L	.ot		Between-Operat	tor/Run		Between-D	ay
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID¹	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid
			Posit	ive Cell Line Pa	nel Member	s: HPV	/16/18 Weak Pos	sitive (0.3 x L	oD)		
	0.67	1.8	1	60.0	18/30	5	57.8	26/45	1	66.7	12/18
HPV16 Weak			2	63.3	19/30	6	68.9	31/45	2	61.1	11/18
Positive			3	66.7	20/30				3	72.2	13/18
(0.3 x LoD)									4	66.7	12/18
									5	50.0	9/18
	1.07	3.0	1	70.0	21/30	5	73.3	33/45	1	77.8	14/18
HPV18 Weak			2	70.0	21/30	6	64.4	29/45	2	72.2	13/18
Positive			3	66.7	20/30				3	72.2	13/18
(0.3 x LoD)									4	72.2	13/18
									5	50.0	9/18
	1		Pos	itive Cell Line P	anel Memb	ers: HF	PV16/18 Low Pos	sitive (1 x Lol)		
	0.44	1.2	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
HPV16 Low Positive			2	96.7	29/30	6	95.6	43/45	2	100.0	18/18
			3	96.7	29/30				3	94.4	17/18
(1 x LoD)									4	100.0	18/18
									5	94.4	17/18
	0.74	2.2	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
HPV18 Low			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(1 x LoD)									4	100.0	18/18
									5	100.0	18/18
			P	ositive Cell Line	Panel Mem	bers:	HPV16/18 Positi	ve (3 x LoD) ²			
	0.38	1.1	1	100.0	29/29	5	100.0	45/45	1	100.0	18/18
HPV16			2	100.0	30/30	6	100.0	44/44	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	17/17
									5	100.0	18/18
	0.41	1.2	1	100.0	29/29	5	100.0	45/45	1	100.0	18/18
HPV18			2	100.0	30/30	6	100.0	44/44	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	17/17
									5	100.0	18/18

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					Numb	er of P	ositives/Total N	umber Valid F	Result	s	
				Between-L	.ot		Between-Opera	tor/Run		Between-D	ay
Panel Member	Ct SD	Ct CV%	ID	Positive Agreement (%)	Positive/ Valid	ID¹	Positive Agreement (%)	Positive/ Valid	ID	Positive Agreement (%)	Positive/ Valid
		•		P	ositive Clini	cal Pa	nel Members				
	0.91	2.7	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV16			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Low Positive			3	100.0	30/30				3	100.0	18/18
(1 x LoD)									4	100.0	18/18
									5	100.0	18/18
	0.88	2.7	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV16			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	18/18
									5	100.0	18/18
	0.70	2.0	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV18			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Low Positive			3	100.0	30/30				3	100.0	18/18
(1 x LoD)									4	100.0	18/18
									5	100.0	18/18
	1.02	3.0	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV18			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	18/18
									5	100.0	18/18
	2.32	7.4	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV45			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Low Positive			3	100.0	30/30				3	100.0	18/18
(1 x LoD)									4	100.0	18/18
									5	100.0	18/18
	1.74	5.9	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV45			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Positive			3	100.0	30/30				3	100.0	18/18
(3 x LoD)									4	100.0	18/18
									5	100.0	18/18
	1.06	3.2	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV39			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Low Positive			3	100.0	30/30				3	100.0	18/18
(1 x LoD)									4	100.0	18/18
/									5	100.0	18/18
	1.52	4.8	1	100.0	30/30	5	100.0	45/45	1	100.0	18/18
Pooled HPV39			2	100.0	30/30	6	100.0	45/45	2	100.0	18/18
Positive			3	100.0	30/30			15. 10	3	100.0	18/18
(3 x LoD)									4	100.0	18/18
/									5	100.0	18/18

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¹ Operators 5 and 6 were at testing site 3.
² One replicate failed due to processing error and excluded from analysis.

Table 69 Overall mean, standard deviation, and coefficients of variation (%) for cycle threshold, estimated from positive panel members on the cobas[®] 8800

			Standard Deviation, Coefficient of Variation (%)								
Panel		Mean	Between-	Between-	Between-	Within-	Total				
Member	N	Ct	Lot	Operator/Run	Day	Run	CV				
Positive Cell Line Panel Members											
HPV16/18 Weak Positive (0.3 x LoD)											
HPV16 Weak Positive (0.3 x LoD)	58	36.6	0.00, (0.00%)	0.16, (0.45%)	0.16, (0.44%)	0.63, (1.72%)	1.8				
HPV18 Weak Positive (0.3 x LoD)	63	35.5	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	1.07, (3.01%)	3.0				
HPV16/18 Low Positive (1 x LoD)											
HPV16 Low Positive (1 x LoD)	88	35.6	0.00, (0.00%)	0.00, (0.00%)	0.14, (0.40%)	0.42, (1.18%)	1.2				
HPV18 Low Positive (1 x LoD)	90	34.2	0.00, (0.00%)	0.16, (0.46%)	0.30, (0.86%)	0.66, (1.94%)	2.2				
HPV16/18 Positive (3 x LoD)											
HPV16 Positive (3 x LoD)	89	34.6	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.38, (1.10%)	1.1				
HPV18 Positive (3 x LoD)	89	32.7	0.00, (0.00%)	0.00, (0.00%)	0.00, (0.00%)	0.41, (1.24%)	1.2				
Positive Clinical Panel Members											
Pooled HPV16 Low Positive (1 x LoD)	90	33.6	0.07, (0.21%)	0.00, (0.00%)	0.00, (0.00%)	0.91, (2.71%)	2.7				
Pooled HPV16 Positive (3 x LoD)	90	32.9	0.00, (0.00%)	0.00, (0.00%)	0.13, (0.39%)	0.87, (2.64%)	2.7				
Pooled HPV18 Low Positive (1 x LoD)	90	35.0	0.00, (0.00%)	0.05, (0.15%)	0.16, (0.47%)	0.68, (1.94%)	2.0				
Pooled HPV18 Positive (3 x LoD)	90	33.6	0.24, (0.70%)	0.25, (0.75%)	0.18, (0.54%)	0.94, (2.80%)	3.0				
Pooled HPV45 Low Positive (1 x LoD)	90	31.2	0.40, (1.27%)	0.00, (0.00%)	0.74, (2.37%)	2.16, (6.93%)	7.4				
Pooled HPV45 Positive (3 x LoD)	90	29.5	0.00, (0.00%)	0.41, (1.40%)	0.59, (2.00%)	1.59, (5.39%)	5.9				
Pooled HPV39 Low Positive (1 x LoD)	90	33.1	0.00, (0.00%)	0.00, (0.00%)	0.30, (0.92%)	1.02, (3.07%)	3.2				
Pooled HPV39 Positive (3 x LoD)	90	31.4	0.00, (0.00%)	0.00, (0.00%)	0.59, (1.88%)	1.40, (4.46%)	4.8				

Notes: Ct=Cycle Threshold; CV=Coefficient of Variation

Additional information

Key assay features

Sample types Cervical specimen collected in PreservCyt® Solution

Amount of sample processed

 \geq 1000 μ L required in sample tube for PreservCyt[®] samples, instrument processes 400 μ L

Maximum volume of 4 mL in sample tube for PreservCyt® samples

Test duration <3.5 hours to first result

Symbols

The following symbols are used in labeling for Roche PCR diagnostic products.

Table 70 Symbols used in labeling for Roche PCR diagnostics products

© sw	Ancillary Software	LLR	Lower Limit of Assigned Range	CONTROL — Negative Control
EC REP	Authorized representative in the European community	ULR	Upper Limit of Assigned Range	CONTROL + Positive Control
BARCODE	Barcode Data Sheet		Store in the dark	CONTROL Control
LOT	Batch code	\sum	Contains sufficient for < n> tests	Assigned Range [copies/mL] Assigned Range (copies/mL)
\$	Biological risks	X	Temperature limit	Assigned Range [IU/mL] Assigned Range (IU/mL)
REF	Catalogue number	TDF	Test Definition File	Procedure Standard Standard Procedure
	Consult instructions for use		Manufacturer	Procedure UltraSensitive Ultrasensitive Procedure
Cont.	Contents of kit	\geq	Use-by date	QS copies / PCR QS copies per PCR reaction, use the QS copies
D	Distributed by	GTIN	Global Trade Item Number	per PCR reaction in calculation of the results. QS IU/PCR QS IU per PCR reaction, use the QS International Units (IU) per PCR reaction in calculation of the results.
Î	For IVD performance evaluation only	SN	Serial number	This product fulfills the requirements of the European Directive 98/79 EC for <i>in vitro</i> diagnostic medical devices.
Rx Only	US Only: Federal law restricts this device to sale by or on the order of a physician.	~~	Date of manufacture	•
IVD	In Vitro diagnostic medical device	2	Do not reuse	

US Customer Technical Support 1-800-526-1247

Manufacturer and distributors

Table 71 Manufacturer and distributors



Roche Molecular Systems, Inc. 1080 US Highway 202 South Branchburg, NJ 08876 USA www.roche.com



Roche Diagnostics 9115 Hague Road Indianapolis, IN 46250-0457 USA (For Technical Assistance call the Roche Response Center toll-free: 1-800-526-1247) Roche Diagnostics GmbH Sandhofer Strasse 116 68305 Mannheim, Germany

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